

NEWS AND VIEWS

Miniaturized two-photon microscope: seeing clearer and deeper into the brain

Alcino J Silva

Light: Science & Applications (2017) 6, e17104; doi:10.1038/lsa.2017.104; published online 25 August 2017

A miniature two-photon microscope circumvents previous limitations of this technology and now allows imaging of activity not only in the soma but also in the dendrites and spines of neurons in freely moving animals engaged in a wide spectrum of behaviors.

Conventional brain imaging in animal models with multiphoton microscopy usually requires that the animal has its head fixed to a very large table microscope, which severely limits the types of behaviors that can be studied.

To address this limitation, recently, Schnitzer and colleagues at Stanford University^{1,2} developed a complementary technique that involves head-mounted microscopes for fluorescent imaging, leveraging the power of ultrasensitive fluorescent proteins that are widely used for imaging neuronal activity (GCaMP6, Figure 1c). Inspired by Schnitzer's invention, our laboratory helped construct an open-source, head-mounted, integrated miniature fluorescence microscope^{1,2} (the UCLA miniscope) with associated hardware and software to image *in vivo* calcium transients using the calcium indicator GCaMP6f (Figure 1d)⁵. We recently used these head-mounted microscopes to study CA1 neuronal ensembles encoding multiple memories over time⁶.

These head-mounted microscopes for fluorescent imaging, however, have limitations regarding the types of structures and events that can be visualized. For example, with these microscopes, imaging is limited to somas since it is virtually impossible to resolve smaller structures such as dendrites. To address these limitations, in 2001, Denk and colleagues designed a head-mounted two-photon microscope (Figure 1a)³. Although this was an important conceptual development, this microscope and others that followed have had a very limited impact in neuroscience due to technological problems that limited their use. This is due to several problems, including optical limitations of the fibers used, which cannot deliver 920-nm femtosecond laser pulses to a living brain without significant distortion that precludes useful imaging, inadequate sampling rates and significant motion artifacts. Because of these limitations, stable imaging

of calcium dynamics in dendritic spines of freely behaving animals, for example, has remained beyond the capability of available approaches.

To address these problems, Weijian Zong (now at HHMI), Liangyi Chen and Peace Cheng at Peking University developed a fast, high-resolution and miniaturized two-photon microscope capable of resolving activity in single spines in freely moving mice⁴. Weighing only approximately 2 g, this miniaturized two-photon microscope can image with a very high spatiotemporal resolution, and its nifty miniature scanner allows for random-access imaging and fast free-line scanning. A key development that enabled this remarkable approach was the design of a hollow-core photonic crystal fiber, developed by Aimin Wang, that is capable of delivering 920-nm femtosecond laser pulses with essentially no nonlinear pulse-broadening. This allows the miniaturized two-photon microscope to perform comparably to its big, bench-top cousins. The authors tested their miniature microscope in a wide range of behavioral paradigms, and they produced beautiful GCaMP6f imaging of activity not only in the soma but also in dendrites and spines. Importantly, the imaging was not degraded by the unpredictable, often sudden and vigorous head and body movements of the animal, thus circumventing a serious limitation of previous designs.

By every standard, this microscope represents a fundamental development that will change the way neuroscientists image cellular and subcellular structures in actively moving animals. It also inspires further efforts to design optical systems geared toward miniaturized imaging systems. The potential impact of this microscope goes beyond imaging of neuronal and dendritic activity. Systems neuroscience is primed to enter a new era, in which imaging of complex biological events in identified cells and subcellular structures of cell ensembles will bring us closer than ever before to understanding the very organizing principles underlying complex behaviors. This marvelous invention just brought us a step closer to that goal.

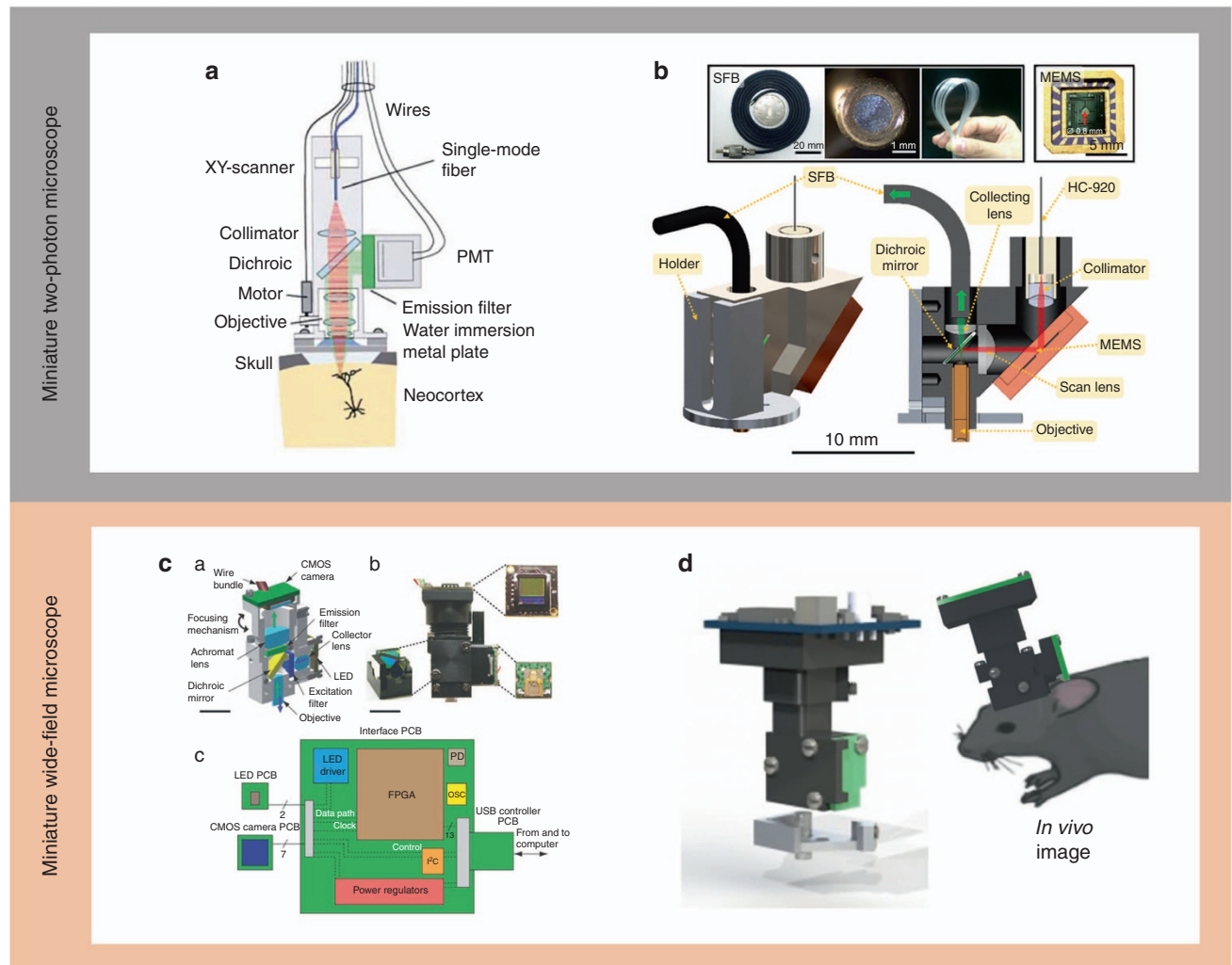



Figure 1 Miniaturized head-mounted microscopes. (a) Miniature two-photon microscope by Denk *et al.* in which the PMY is directly mounted on the microscope³; (b) miniature two-photon microscope by Cheng *et al.* weighing only 2.2 g⁴; (c) miniature wide-field microscope by Schnitzer *et al.*^{1,2}; (d) the UCLA miniature wide-field miniscope⁵.

CONFLICT OF INTEREST

The author declares no conflict of interest.

- 5 Ziv Y, Burns LD, Cocker ED, Hamel EO, Ghosh KK *et al.* Long-term dynamics of CA1 hippocampal place codes. *Nat Neurosci* 2013; **16**: 264–266.
- 6 Zong WJ, Wu RL, Li ML, Hu YH, Li YJ *et al.* Fast high-resolution miniature two-photon microscopy for brain imaging in freely behaving mice. *Nat Methods* 2017; **14**: 713–719.

- 1 Cai DJ, Aharoni D, Shuman T, Shobe J, Biane J *et al.* A shared neural ensemble links distinct contextual memories encoded close in time. *Nature* 2016; **534**: 115–118.
- 2 Chen TW, Wardill TJ, Sun Y, Pulver SR, Renninger SL *et al.* Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* 2013; **499**: 295–300.
- 3 Ghosh KK, Burns LD, Cocker ED, Nimmerjahn A, Ziv Y *et al.* Miniaturized integration of a fluorescence microscope. *Nat Methods* 2011; **8**: 871–878.
- 4 Helmchen F, Fee MS, Tank DW, Denk W. A miniature head-mounted two-photon microscope: high-resolution brain imaging in freely moving animals. *Neuron* 2001; **31**: 903–912.

 This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>

© The Author(s) 2017