³Knight Cancer Institute, Oregon Health & Science University, Portland, OR, USA and ⁴Department of Pathology, Oregon Health & Science University, Portland, OR, USA E-mail: fang@ohsu.edu.

REFERENCES

- 1 Flex E, Petrangeli V, Stella L, Chiaretti S, Hornakova T, Knoops L *et al.* Somatically acquired JAK1 mutations in adult acute lymphoblastic leukemia. *J Exp Med* 2008; **205**: 751–758.
- 2 Mullighan CG, Zhang J, Harvey RC, Collins-Underwood JR, Schulman BA Phillips LA *et al.* JAK mutations in high-risk childhood acute lymphoblastic leukemia. *Proc Nat Acad Sci USA* 2009; **106**: 9414–9418.
- 3 Gaikwad A, Rye CL, Devidas M, Heerema NA, Carroll AJ, Izraeli S et al. Prevalence and clinical correlates of JAK2 mutations in Down syndrome acute lymphoblastic leukaemia. Br J Haematol 2009; 144: 930–932.
- 4 Jeong EG, Kim MS, Nam HK, Min CK, Lee S, Chung YJ et al. Somatic mutations of JAK1 and JAK3 in acute leukemias and solid cancers. Clin Cancer Res 2008; 14: 3716–3721.
- 5 Kameda T, Shide K, Shimoda HK, Hidaka T, Kubuki Y, Katayose K et al. Absence of gain-of-function JAK1 and JAK3 mutations in adult T cell leukemia/lymphoma. Int J Hematol 2010: 92: 320–325.
- 6 Cornejo MG, Kharas MG, Werneck MB, Le Bras S, Moore SA, Ball B *et al.* Constitutive JAK3 activation induces lymphoproliferative syndromes in murine bone marrow transplantation models. *Blood* 2009; **113**: 2746–2754.

OPEN

- 7 Walters DK, Mercher T, Gu TL, O'Hare T, Tyner JW, Loriaux M et al. Activating alleles of JAK3 in acute megakaryoblastic leukemia. *Cancer Cell* 2006; 10: 65–75.
- 8 Beadling C, Heinrich MC, Warrick A, Forbes EM, Nelson D, Justusson E et al. Multiplex mutation screening by mass spectrometry evaluation of 820 cases from a personalized cancer medicine registry. J Mol Diagn 2011; 13: 504–513.
- 9 Shochat C, Tal N, Bandapalli OR, Palmi C, Ganmore I, te Kronnie G et al. Gain-of-function mutations in interleukin-7 receptor-α (IL7R) in childhood acute lymphoblastic leukemias. J Exp Med 2011; 208: 901–908.
- 10 Zenatti PP, Ribeiro D, Li W, Zuurbier L, Silva MC, Paganin M et al. Oncogenic IL7R gain-of-function mutations in childhood T-cell acute lymphoblastic leukemia. Nat Genet 2011; 43: 932–939.
- 11 Hornakova T, Chiaretti S, Lemaire MM, Foà R, Ben Abdelali R, Asnafi V et al. ALL-associated JAK1 mutations confer hypersensitivity to the antiproliferative effect of type I interferon. Blood 2010; 115: 3287–3295.
- 12 Malinge S, Ragu C, Della-Valle V, Pisani D, Constantinescu SN, Perez C *et al.* Activating mutations in human acute megakaryoblastic leukemia. *Blood* 2008; **112**: 4220–4226.
- 13 Sato T, Toki T, Kanezaki R, Xu G, Terui K, Kanegane H *et al.* Functional analysis of JAK3 mutations in transient myeloproliferative disorder and acute megakaryoblastic leukaemia accompanying Down syndrome. *Br J Haematol* 2008; **141**: 681–688.
- 14 Haan C, Rolvering C, Raulf F, Kapp M, Drückes P, Thomas G *et al.* Jak1 has a dominant role over Jak3 in signal transduction through γc-containing cytokine receptors. *Chem Biol* 2011; **18**: 314–323.
- 15 Changelian PS, Moshinsky D, Kuhn CF, Flanagan ME, Munchhof MJ, Harris TM et al. The specificity of JAK3 kinase inhibitors. Blood 2008; 111: 2155–2157.

Increased CD200 expression in acute myeloid leukemia is linked with an increased frequency of FoxP3⁺ regulatory T cells

Leukemia (2012) 26, 2146-2148; doi:10.1038/leu.2012.75

CD200 is a type-1a transmembrane cell-surface glycoprotein that is normally expressed in critical tissues such as the central nervous system and testis, as well as certain leukocytes, including T and B lymphocytes, where its role is to promote peripheral tolerance and protect immune privileged sites.¹ CD200 has no known intracellular signaling motif, but induces immunosuppression through engagement with CD200R, a cell-surface receptor homolog, which is expressed on leukocytes of myeloid lineage, including mast-cells, macrophages, basophils, dendritic cells as well as certain T-cell populations.² CD200, which is frequently overexpressed in acute myeloid leukemia (AML) patient blasts and associated with a worse outcome,³ has the potential to induce the formation of CD4⁺CD25⁺⁺FoxP3⁺ regulatory T cells (Tregs),² a subset of immunosuppressive T cells that are linked with a poor prognosis in $\rm AML.^5$ Most importantly, Tregs have been documented to suppress the anti-leukemia response in vitro from AML patients,⁶ suggesting that these cells have an important role in regulating AML patient tumor immunity. We therefore investigated whether CD200 protein expression on human AML blasts was associated with an increased frequency of Treq cells in AML using a cohort of 40 AML patients (at the point of diagnosis before to any treatment; Supplementary Table S1 for patient demographic). AML blast CD200 protein expression and patient Treg frequency was analyzed by flow cytometry (for full gating strategy, refer to Supplementary Figure S1). The data show that CD200 blast expression level correlated significantly with the frequency of Treg cells (Figure 1a). This association was found to be independent of white blood cell count (Figure 1b), suggesting that CD200 expression on AML blasts promotes Treg formation. We also examined whether CD200 expression was differentially expressed on AML blasts with a putative leukemic stem cell phenotype (CD34⁺CD38⁻),⁷ but could find no evidence for this within the six samples examined (Supplementary Figure S2).

Given that the depletion of Tregs has been shown to improve T-cell-mediated therapy, as well as improving antitumor activity in leukemia patients that are in complete remission;^{8,9} we investigated whether Tregs from CD200^{hi} patients were functionally immunosuppressive. We examined the ability of purified Tregs to suppress naïve responder T-cell (CD4⁺CD25⁻) proliferation following CD3/CD28 co-stimulation (refer to Supplementary Materials and Methods). The data demonstrate that Tregs isolated from CD200^{hi} AML patients were capable of suppressing T-cell proliferation at responder to Treg ratios of >1 to 0.01 (Figure 1c). Reciprocal analysis of Tregs from CD200^{lo} patients could not be carried out because of the extremely low frequencies of these cells in CD200^{lo} patients (Figure 1d). In fact, Treg frequencies in CD200^{lo} AML patients were uniformly lower than in healthy donor controls, suggesting that CD200 has an influence on Treg induction in this context.

Since mouse models have suggested that CD200-induced Tregs are likely to mediate Th1 immunosuppression,¹⁰ a cytokine response with a prognostic link in leukemia,¹¹ we measured the Th1 cytokine response (TNF α , IL2 and IFN γ production) in CD200^{hi} patients before and after Treg depletion. Representative flow cytometric plots confirmed Treg depletion using magnetic separation (Figure 2a). However, removal of Tregs alone was insufficient to significantly improve the Th1 cytokine response as detected by intracellular cytokine staining (Figure 2b). Elsewhere we report that, in addition to its known influence on Treg production, CD200 is also directly immunosuppressive through engagement with CD200R

Accepted article preview online 20 March 2012; advance online publication, 13 April 2012

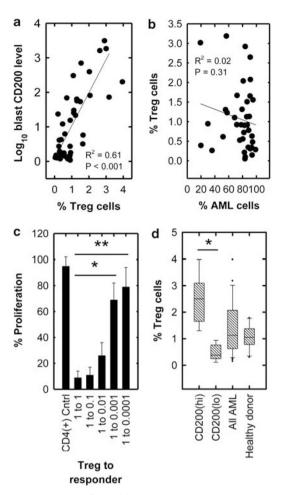


Figure 1. Assessment of AML blast CD200 expression with respect to Treg frequency and function. Treg cells from 40 AML patients at diagnosis were identified by flow cytometry based on the frequency of CD3⁺ CD4⁺CD25⁺ T cells that were > 95% FoxP3⁺ (refer to Supplementary Figure S1). (a) Linear regression analysis comparing AML blast CD200 expression level with the frequency of Tregs from 40 diagnostic AML samples. (b) Correlation of AML blast frequency with Treg cell frequency. To assess the suppressive function, Tregs and naive T cells (CD4⁺CD25⁻) were isolated from CD200^{hi} AML patients by magnetic separation and added together at increasing ratios before CD3/CD28 co-stimulation. Suppression was measured by the ability of Tregs to inhibit naïve CD4⁺ T-cell proliferation (refer to Supplementary Materials and Methods). (**c**) Suppression of naïve CD4⁺ T-cell proliferation by AML Treg cells at indicated Treg to responder ratios (data represent mean \pm 1 s.d., n = 4). (d) Treg cell frequency stratified with respect to AML blast CD200 expression. *P < 0.05 and **P < 0.01 analyzed by one-way analysis of variance with Tukey's multiple comparison test.

on T cells in AML.¹² However, in the present study we show that removal of Treg cells alone is unlikely to show effect in these assays when large numbers of CD200⁺ blast cells are present. These data would predict that Tregs have little impact on immunosuppression at diagnosis when the disease burden is high, as the dominant mode of T-cell suppression would be mediated by CD200^{hi} blasts directly inhibiting Th1 responses.¹² Conversely, following chemotherapy, the reduction of tumor burden would be predicted to make Treg-mediated immunosuppression cells the dominant factor.

In summary, these findings illustrate a clear correlation between blast CD200 expression level and the frequency of immunosuppressive Treg cells. Previous studies of human AML have also reported increased frequencies of Tregs in untreated disease¹³ as well as during regeneration following treatment,¹⁴ however, the

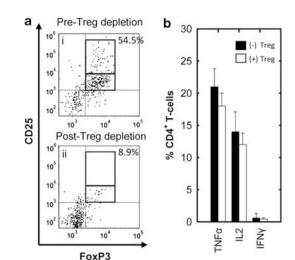


Figure 2. Assessment of AML patient Treg depletion in recovering the Th1 response in CD200^{hi} patients. Tregs cells were depleted from CD200^{hi} AML patients by magnetic separation. (**a**) Representative flow cytometric data depicting T cells (i) before depletion of Tregs (CD3⁺CD4⁺CD25⁺⁺FoxP3⁺) and (ii) following Treg depletion from a CD200^{hi} AML patient. AML cells were stimulated with PMA/ionomycin for 6 h at 37 °C before intracellular Th1 cytokine staining was performed. Flow cytometry was used to assess Th1 cytokine levels within the CD45⁺⁺CD3⁺CD4⁺ fraction. (**b**) Summary data showing the Th1 response from CD200^{hi} AML patients ± Treg cells (*n* = 6).

link with CD200 expression has not previously been established. Our data also indicate that while Tregs in these patients may be functionally immunosuppressive, they may only become influential following cytoreduction. Indeed, several studies have identified that increased frequencies of Tregs are associated with relapse in myeloid malignancy.^{15,16} Furthermore, phase-I studies using anti-CD200 monoclonal antibody immunotherapy have shown that blocking CD200 is sufficient to reduce the Treg frequency in chronic lymphocytic leukemia patients,¹⁷ illustrating that blocking CD200 may be therapeutically advantageous in AML.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

This work was funded by Leukemia and Lymphoma Research UK. We thank Gareth Betts (Cardiff University) for his technical assistance in analyzing Tregs. We are grateful to the patients for access to material enrolled in the NCRI clinical trials. Dr Steve Coles is currently funded by NISCHR, Wales, UK.

AUTHOR CONTRIBUTIONS

SJC designed and performed the experiments/analyzed all data and co-wrote the manuscript. RKH provided statistical guidance. ECYW provided biological insight. AKB provided resources and clinical insight. SM, RLD and AT contributed to experimental design and co-wrote the manuscript.

SJ Coles¹, RK Hills¹, ECY Wang², AK Burnett¹, S Man^{2,3}, RL Darley^{1,3} and A Tonks^{1,3} ¹Department of Haematology, Institute of Cancer and Genetics, Cardiff University, Wales, UK and ²Institute of Infection and Immunity, School of Medicine, Cardiff University, Wales, UK E-mail: Tonksa@cf.ac.uk ³These authors contributed equally to this work.

REFERENCES

- Wright GJ, Jones M, Puklavec MJ, Brown MH, Barclay AN. The unusual distribution of the neuronal/lymphoid cell surface CD200 (OX2) glycoprotein is conserved in humans. *Immunology* 2001; **102**: 173–179.
- 2 Wright GJ, Cherwinski H, Foster-Cuevas M, Brooke G, Puklavec MJ, Bigler M et al. Characterization of the CD200 receptor family in mice and humans and their interactions with CD200. J Immunol 2003; **171**: 3034–3046.
- 3 Tonks A, Hills R, White P, Rosie B, Mills KI, Burnett AK et al. CD200 as a prognostic factor in acute myeloid leukaemia. *Leukemia* 2007; **21**: 566–568.
- 4 Gorczynski RM, Lee L, Boudakov I. Augmented induction of CD4 + CD25 + Treg using monoclonal antibodies to CD200R. *Transplantation* 2005; **79**: 488–491.
- 5 Shenghui Z, Yixiang H, Jianbo W, Kang Y, Laixi B, Yan Z et al. Elevated frequencies of CD4(+)CD25(+)CD127(lo) regulatory T cells is associated to poor prognosis in patients with acute myeloid leukemia. Int J Cancer 2010; 129: 1373–1381.
- 6 Curti A, Trabanelli S, Onofri C, Aluigi M, Salvestrini V, Ocadlikova D et al. Indoleamine 2,3-dioxygenase-expressing leukemic dendritic cells impair a leukemia-specific immune response by inducing potent T regulatory cells. *Haematologica* 2010; **95**: 2022–2030.
- 7 Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med* 1997; **3**: 730–737.
- 8 Zhou Q, Bucher C, Munger ME, Highfill SL, Tolar J, Munn DH et al. Depletion of endogenous tumor-associated regulatory T cells improves the efficacy of adoptive cytotoxic T-cell immunotherapy in murine acute myeloid leukemia. Blood 2009; **114**: 3793–3802.
- 9 Maury S, Lemoine FM, Hicheri Y, Rosenzwajg M, Badoual C, Cherai M et al. CD4 + CD25 + regulatory T cell depletion improves the graft-versus-tumor effect of donor lymphocytes after allogeneic hematopoietic stem cell transplantation. *Sci Transl Med* 2010; 2: 41ra52.
- 10 Gorczynski L, Chen Z, Hu J, Kai Y, Lei J, Ramakrishna V et al. Evidence that an OX-2positive cell can inhibit the stimulation of type 1 cytokine production by bone marrow-derived B7-1 (and B7-2)-positive dendritic cells. J Immunol 1999; 162: 774–781.

- 11 Gallego A, Vargas JA, Castejon R, Citores MJ, Romero Y, Millan I et al. Production of intracellular IL-2, TNF-alpha, and IFN-gamma by T cells in B-CLL. Cytometry B Clin Cytom 2003; 56: 23–29.
- 12 Coles SJ, Hills RK, Wang ECY, Burnett AK, Man S, Darley RL *et al.* Expression of CD200 on AML blasts directly suppresses memory T-cell function. *Leukemia* 2012; 26: 2148–2151.
- 13 Szczepanski MJ, Szajnik M, Czystowska M, Mandapathil M, Strauss L, Welsh A *et al.* Increased frequency and suppression by regulatory T cells in patients with acute myelogenous leukemia. *Clin Cancer Res* 2009; **15**: 3325–3332.
- 14 Ersvaer E, Liseth K, Skavland J, Gjertsen BT, Bruserud O. Intensive chemotherapy for acute myeloid leukemia differentially affects circulating TC1, TH1, TH17 and TREG cells. *BMC Immunol* 2010; **11**: 38.
- 15 Delluc S, Hachem P, Rusakiewicz S, Gaston A, Marchiol-Fournigault C, Tourneur L et al. Dramatic efficacy improvement of a DC-based vaccine against AML by CD25 T cell depletion allowing the induction of a long-lasting T cell response. Cancer Immunol Immunother 2009; 58: 1669–1677.
- 16 Nadal E, Garin M, Kaeda J, Apperley J, Lechler R, Dazzi F. Increased frequencies of CD4(+)CD25(high) T(regs) correlate with disease relapse after allogeneic stem cell transplantation for chronic myeloid leukemia. *Leukemia* 2007; 21: 472–479.
- 17 Mahadevan D, Lanasa MC, Whelden M, Faas SJ, Ulery TL, Kukreja A et al. First-In-Human Phase I Dose Escalation Study of a Humanized Anti-CD200 Antibody (Samalizumab) In Patients with Advanced Stage B Cell Chronic Lymphocytic Leukemia (B-CLL) or Multiple Myeloma (MM). 52nd ASH Annu Meet Exposition. 2465; Orlando, FL; 2012.

This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-sa/3.0/

Supplementary Information accompanies the paper on the Leukemia website (http://www.nature.com/leu)

OPEN

Expression of CD200 on AML blasts directly suppresses memory T-cell function

Leukemia (2012) 26, 2148-2151; doi:10.1038/leu.2012.77

Previous studies have shown that immunosuppression in acute myeloid leukemia (AML) is associated with changes in the adaptive immune compartment. Such changes include the suppression of memory T-cell function¹ and the suppression of Th1 cytokine (TNF α , IL-2 and IFN γ)-producing cells.² A suppression of immune response in AML is associated with a worse patient outcome and increased risk of relapse,³ as well as increased risk of infection impairing patient recovery.⁴ The over-expression of the immunosuppressive ligand CD200 is also associated with an increased risk of relapse in AML (hazard ratio 1.7); an observation consistent with a hypothesis in which CD200 inhibits clearance of residual disease.^{5,6} As memory T-cell responses are central for tumor immunosurveillance and contribute to prolonged molecular remission,⁷ we carried out this study to establish how these responses were affected in AML patients over-expressing CD200 (Supplemental Table S1). We initially investigated whether CD200 expression on AML blasts influenced CD8⁺ T-cell cytotoxic potential and the frequency of TNFa-, IL-2- and IFNy-producing $CD4^+/CD8^+$ memory T-cells (Supplementary Materials and Methods and Supplemental Figure S1 for gating strategy). Using CD107a as a marker of cytotoxic function, AML cells were activated with PMA/ionomycin. We show that the frequency of CD107a⁺CD8⁺ memory T-cells was significantly reduced by \sim 50% for CD200^{hi} patients when compared with CD200^{lo} AML, demonstrating that cytotoxic memory T-cell activity was compromised in CD200^{hi} patients (Figure 1a). Furthermore, the frequencies of TNF α -, IL-2- and IFN γ -producing CD4⁺ memory cells were also reduced by ~50% for CD200^{hi} patients when compared with CD200^{lo} AML (Figure 1b), significantly so in the case of IL-2 and IFN γ . Interestingly, CD200^{lo} patients displayed a higher IFN γ response, not only with respect to CD200^{hi} patients but also in comparison to healthy donors, suggesting a role for this cytokine in AML, which is attenuated by CD200. No difference was observed for TNF α -, IL-2- and IFN γ -producing CD8⁺ memory cells between CD200^{hi}, CD200^{lo} and healthy donors (data not shown). CD200 has also been reported to mediate suppression of the Th1 response in chronic lymphocytic leukemia as well as solid tumors,^{8,9} suggesting that CD200-mediated Th1 suppression is a central mechanism in cancer immunomodulation.

The ability to simultaneously produce TNF α , IL-2 and IFN γ is an important indicator of 'T-cell quality' in anti-tumor/viral responses.¹⁰ We therefore simultaneously measured the production of all these cytokines in CD200^{bi} and CD200^{lo} patients after PMA/ionomycin stimulation (Supplemental Figure S1). A significant reduction (30%) in CD4⁺ memory T-cells capable of simultaneously producing TNF α , IL-2 and IFN γ was observed in CD200^{bi} compared with CD200^{lo} AML patients (Figure 1c). Although a similar reduction was observed within the CD8⁺ memory cells, the changes in this subpopulation were less consistent and were not statistically significant (Supplemental Figure S2). To assess if CD200 expression on AML blasts influences the memory Th1 response through an antigen-specific mechanism, we compared T-cell responses with common microbial recall

Accepted article preview online 20 March 2012; advance online publication, 13 April 2012

npg 2148