

ORIGINAL ARTICLE

Whole-exome sequencing reveals a novel frameshift mutation in the *FAM161A* gene causing autosomal recessive retinitis pigmentosa in the Indian population

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Retinitis pigmentosa (RP) is a heterogenous group of inherited retinal degenerations caused by mutations in at least 50 genes. To identify genetic mutations underlying autosomal recessive RP (arRP), we performed whole-exome sequencing study on two consanguineous marriage Indian families (RP-252 and RP-182) and 100 sporadic RP patients. Here we reported novel mutation in *FAM161A* in RP-252 and RP-182 with two patients affected with RP in each family. The *FAM161A* gene was identified as the causative gene for RP28, an autosomal recessive form of RP. By whole-exome sequencing we identified several homozygous genomic regions, one of which included the recently identified *FAM161A* gene mutated in RP28-linked arRP. Sequencing analysis revealed the presence of a novel homozygous frameshift mutation p.R592FsX2 in both patients of family RP-252 and family RP-182. In 100 sporadic Indian RP patients, this novel homozygous frameshift mutation p.R592FsX2 was identified in one sporadic patient ARRP-S-I-46 by whole-exome sequencing and validated by Sanger sequencing. Meanwhile, this homozygous frameshift mutation was absent in 1000 ethnicity-matched control samples screened by direct Sanger sequencing. In conclusion, we identified a novel homozygous frameshift mutations of RP28-linked RP gene *FAM161A* in Indian population. *Journal of Human Genetics* (2015) 60, 625–630; doi:10.1038/jhg.2015.92; published online 6 August 2015

INTRODUCTION

Retinitis pigmentosa (RP) is a group of hereditary degenerative diseases of the retina displaying a significant phenotypic and genotypic variability and characterized by the progressive decrease of rod and cone photoreceptor function.¹ The disease is highly heterogeneous with a prevalence of 1:4000 in the world² and it can be inherited as autosomal recessive (50–60%), autosomal dominant (30–40%) or X-linked (5–15%).³ Initial symptoms include night blindness and a gradual constriction of peripheral vision caused by rod photoreceptor death. Secondary loss of cone photoreceptors may subsequently lead to impairment of central vision and ultimately legal blindness.⁴ Characteristic clinical features are elevated final dark adaptation thresholds and reduced and delayed full-field electroretinogram signals, which measures the electrical response of the retina to flashes

of light.⁵ In recent years enormous efforts have been made in research and an early recognition of retinal degenerations has become possible.

At present, according to the RetNet database (<https://sph.uth.edu/retnet/>), over 60 genes have been associated with non-syndromic RP.⁶ The functions of these genes include components of the photo-transduction cascade and the visual cycle, the regulation of retinal gene expression and general cellular processes like splicing, protein modification and degradation.⁷ In the recent years, numbers of RP genes encode proteins localized at the base of and within the connecting cilium as well as the outer segment axoneme of photoreceptor cells are growing.⁸ Nevertheless, these mutations account for only ~50–60% of the existing disease alleles.³

Enormous efforts have been made in identifying genetic mutations contributive to inherited retinal diseases since 1980s. However, most

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of these techniques are relatively inefficient, expensive and labor intensive. These days, the whole-exome sequencing has been proven to be the most efficient tool for identifying novel autosomal recessive RP (arRP) genes and mutations.^{9,10} On account of its remarkable characteristic, whole-exome sequencing can contribute to novel disease gene identification and more efficient and accurate molecular diagnosis.

In Indian population, family-based disease gene analysis is highly effective, because of relatively high levels of consanguinity, large number of siblings in families and presence of isolated subpopulations. As a part of an international cooperation, we used exome sequencing to identify the disease causing genes in families from Indian population with RP. Here we report the identification of a novel homozygous frameshift mutation p.R592FsX2 in the *FAM161A* gene in two consanguineous Indian families and one sporadic Indian RP patient.

MATERIALS AND METHODS

Patients and controls

Two patients and three unaffected family members from two consanguineous marriage families in South India were recruited from Aravind Eye Hospital of India. 100 sporadic Indian RP patients and 1000 control individuals with no history of retinal degeneration were also from Aravind Eye Hospital of India. This research was carried out in accordance with the tenets of the Declaration of Helsinki and was approved by the Aravind Medical Research Foundation and Aravind Eye Hospital of India and Hospital of the University of Electronic Science and Technology of China and Sichuan Provincial People's Hospital. Written informed consent was obtained from the patients who participated in this study or from the legal guardians in case of minors.

DNA samples and exome sequencing

All genomic DNA samples were extracted from peripheral blood leukocytes of relative RP patients and control individuals using a blood DNA extraction kit according to the protocol provided by the manufacturer (TianGen, Beijing, China). Exome sequencing was performed on DNA samples of the index patients by Macrogen Inc, Seoul, Korea. DNA Illumina (San Diego, CA USA) TruSeq Exome Capture System (62 Mb) was used to collect the protein coding regions of human genome DNA. It covered 20 794 genes and 20 1121 exons in the Consensus Coding Sequence Region database (<http://www.illumina.com/applications/sequencing/targetedresequencing.ilmn>).

Data analysis

High-quality sequencing reads were aligned to the human reference genome (NCBI build 37.1/hg19) with SOAPaligner (soap2.21). Based on the SOAP

alignment results, SOAPsnp v1.05 was used to assemble the consensus sequence and call genotypes in target regions. Data were provided as lists of sequence variants (single-nucleotide polymorphisms (SNPs) and short indels). We filtered SOAPsnp results following several steps as described previously.¹¹ Briefly, for SNP quality control, we filtered SOAPsnp results as follows: (i) base quality is more than 20; (ii) depth is between 4 and 200; (iii) estimate copy number is ≤ 2 ; and (iv) the distance between two SNPs must be longer than 4. Small indel detection was performed using the Unified-Genotyper tool from GATK (version v1.0.4705) after all high-quality reads were aligned to the human reference genome using BWA (version 0.5.9-r16). SNP and Indel detection were performed only on the targeted exome regions and flanking regions within 200 bp. The detected variants were annotated and filtered based on four databases, that is, NCBI CCDS (<http://www.ncbi.nlm.nih.gov/CCDS/CcidsBrowse.cgi>), RefSeq (<http://www.ncbi.nlm.nih.gov/RefSeq/>), Ensembl (<http://www.ensembl.org>) and Encode (<http://genome.ucsc.edu/ENCODE>). Four major steps were taken to prioritize all the high-quality variants: (i) variants within intergenic, intronic and UTR regions and synonymous mutations were excluded from downstream analysis; (ii) variants in dbSNP137 (<http://www.ncbi.nlm.nih.gov/projects/SNP/>), 1000 Genome project (<ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp>), YH Database (<http://yh.genomics.org.cn/>), HapMap Project (<ftp://ftp.ncbi.nlm.nih.gov/hapmap>) and our in-house database, which was generated by our laboratory using 1600 whole-exome sequencing data, were excluded; (iii) possible damaging impacts of each variant on protein structure/function were predicted by SIFT (<http://sift.bii.a-star.edu.sg/>) and Polyphen2 (<http://genetics.bwh.harvard.edu/pph2/>); and (iv) gene ontology (<http://www.geneontology.org>) and KEGG pathway annotations (<http://www.ebi.ac.uk/cluster>) were used to predict the biological function of each putative gene.

Mutations validation

After reads calling, mapping and filtering against multiple databases, genomic variants were identified. Validation of the mutations in patients and their relatives was performed by Sanger sequencing on an ABI 3730XL Genetic Analyzer using the following primers: FAM161-p.R592FsX2-F, 5'-GAG GTAAACTGTGAAGTGTGCT-3'; FAM161-p.R592FsX2-R, 5'-CACCTCAGC CTCCAAGTAG-3'. Sequencing data were used to determine whether the remaining mutations co-segregated with the disease in these families and whether the frameshift mutations were confirmed in the 100 sporadic RP patients.

RESULTS

Patients and clinical information

Two patients affected with RP from two consanguineous marriage Indian families RP-252 and RP-182 were recruited (Figure 1). These two RP patients presented with an early-onset increasing difficulty to adapt in the dark and gradually decreased vision acuity in both eyes

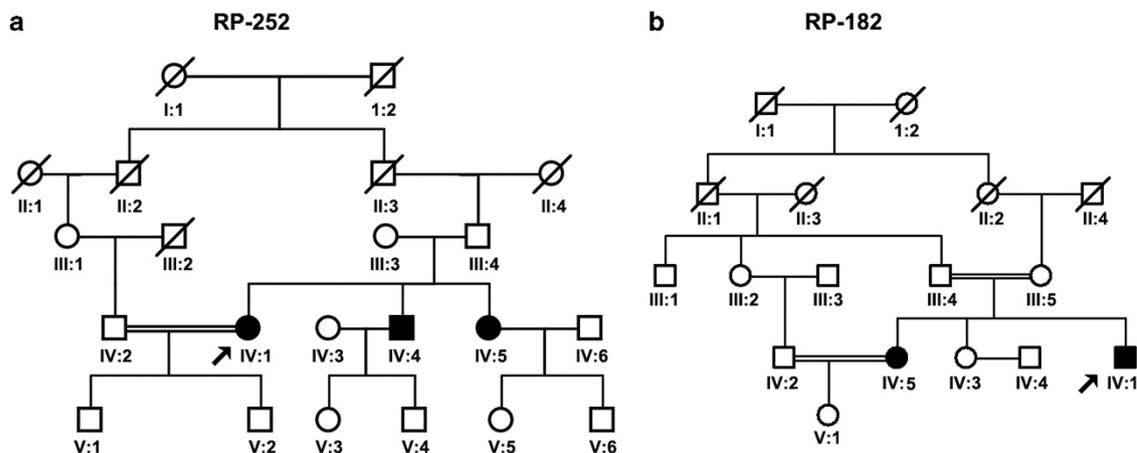


Figure 1 Pedigrees of Indian families RP-252 and RP-182 with arRP. (a) Arrow indicates the proband IV:1 in family RP-252. (b) Arrow indicates the proband IV:1 in family RP-182. Solid symbol indicate affected individual and open symbols indicate unaffected individuals.

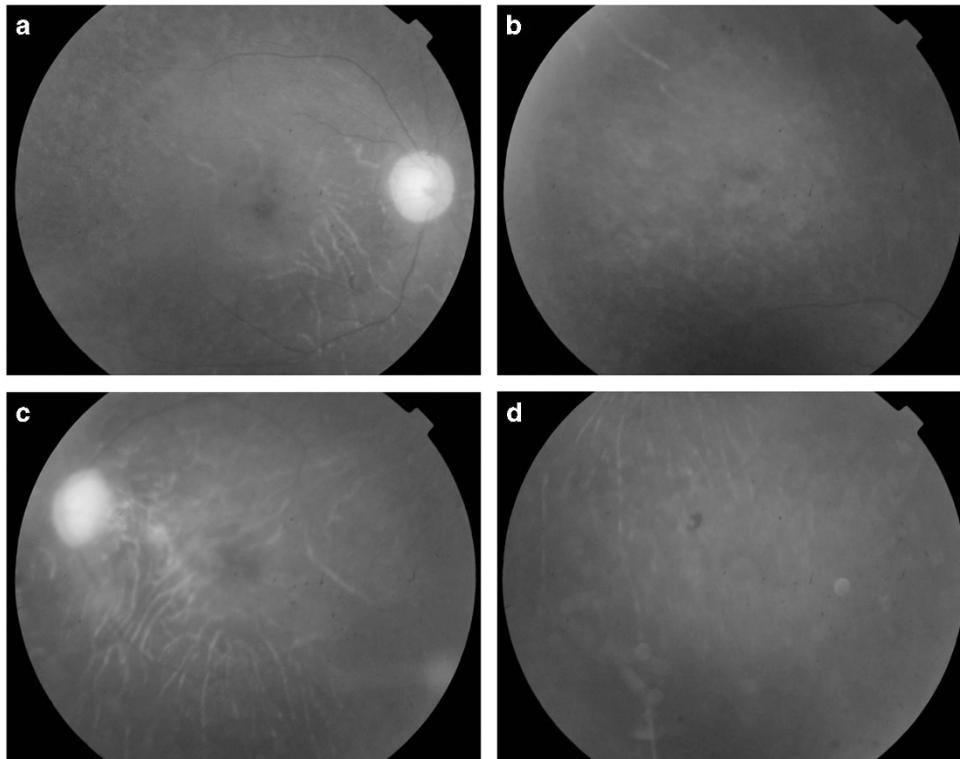


Figure 2 Fundus examination of patient IV:1 in family RP-252. (a and b) Fundus examination for right eye. (c and d) Fundus examination for the left eye. Both retina showed typical signs of RP: pigment formation. A full color version of this figure is available at the *Journal of Human Genetics* journal online.

Table 1 Number of candidate SNPs/Indels filtered against several public variation databases and the in-house data

	Feature—SNPs and indels in patient IV:1 of family RP-252	Feature—SNPs and indels in patient IV:1 of family RP-182
	Total SNPs	64 744
Total Indels	4755	5609
Functional SNP/indels	9385/399	9281/434
Filtered known gene	47	37
Filtered DBsnp135common/indel	43	32
Filtered DBsnp/indel 1000 genome (2011)	42	29
Filtered in-house data	3	4
Homozygous	1	1
Filtered synonymous mutation	1	1

Abbreviation: SNP, single-nucleotide polymorphism.

and typical RP features were observed. The fundus examination of patient IV:4 in RP-252 family was shown in Figure 2. Other examinations, including main complaints, history, ocular examination and electroretinogram examination were shown in Supplementary Data 1. All sporadic Indian RP patients recruited in this study were under normal ophthalmological examinations.

Exome sequencing analysis

By exome sequencing of patient IV:1 in family RP-252 and patient IV:1 in family RP-182, we generated 5.2 billion and 5.3 billion bases of

sequence with average throughput depth of target regions $103.2\times$ and $104.3\times$, respectively. 64 744 SNPs and 4755 indels that may affect amino acid sequence were identified in patient IV:1 of family RP-252 and 67 494 SNPs and 5609 indels that may affect amino acid sequence were identified in patient IV:1 of family RP-182 (Table 1).

In this study, we firstly compared these total variants with reported retina genes (<https://sph.uth.edu/Retnet/>). Screening through the RP gene database, we identified 47, 37 variants of several known RP genes in patients of family RP-252 and family RP-182, respectively. We then compared these variants with the dbSNP135, 1000 Genome Project, HapMap project, YH database and our in-house database, described previously.¹¹ Therefore, we can exclude the variants with high frequency in normal population. By excluding the heterozygous variants and synonymous mutations, we found one variant in each patient. The variants screened by several database were show in Table 1 (candidate mutations filtered with 1000 genome database for RP-252 and RP-182 were listed in Supplementary Table 1).

After these steps, we identified a homozygous frameshift mutation p.R592FsX2 in the known RP gene *FAM161A* in both patients of family RP-252 and family RP-182. We checked the p.R592FsX2 mutation in the human gene mutation database (<http://www.hgmd.org/>) and found that the mutation is novel. We also checked the mutation in the newly available ExAC database of 63 000 control exomes (<http://exac.broadinstitute.org/>) and no variants were reported in this locus of *FAM161A* gene.

Mutation validation and analysis

Sanger sequencing confirmed that the homozygous frameshift mutation p.R592FsX2 was in the affected wife and demonstrated that her son was an unaffected carrier, while her husband carried no

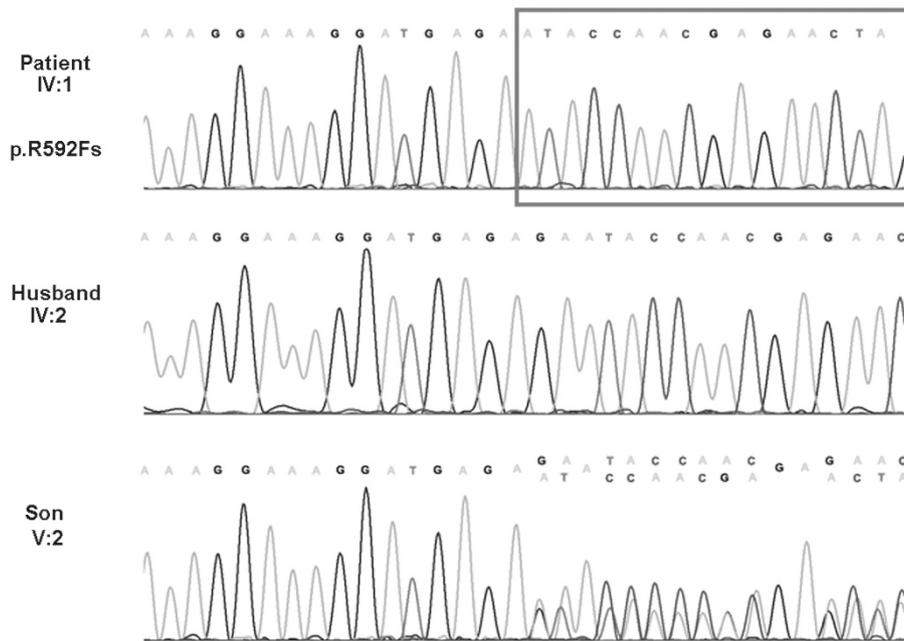
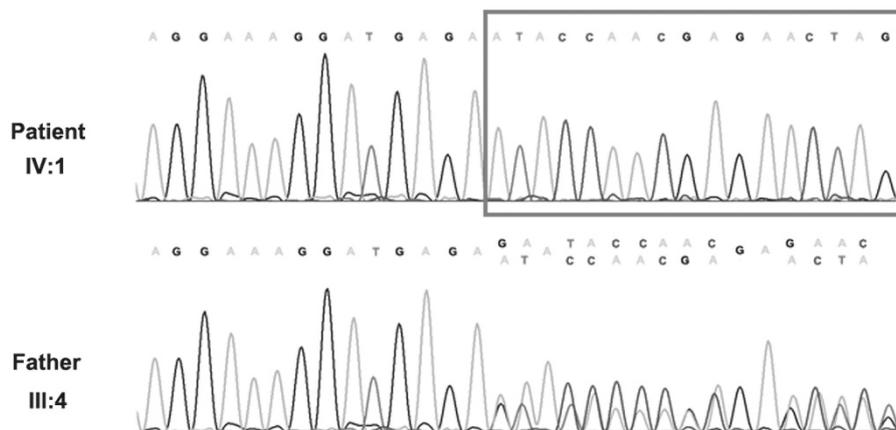
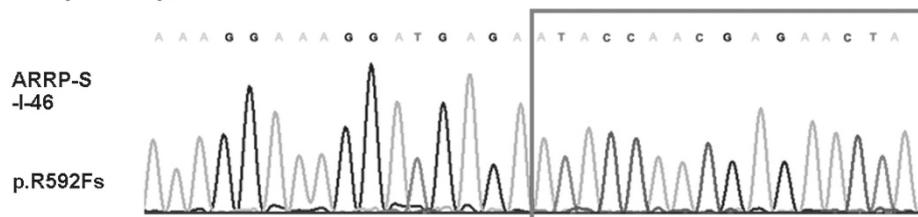
a Family RP-252**b Family RP-182****c Sporadic patients**

Figure 3 Mutation identification of *FAM161A* gene in families RP-252 and RP-182. (a) Sanger sequencing analysis of *FAM161A* in family RP-252 showing the homozygous frameshift mutation p.R592FsX2 co-segregated with the phenotype. Patient IV:1 harbored homozygous frameshift mutation p.R592FsX2 of the *FAM161A* gene. The heterozygous mutation p.R592FsX2 was carried by her son V:2, while her husband IV:2 did not carry this mutation. (b) Sanger sequencing analysis of *FAM161A* in family RP-182 showed patient IV:1 harboring the homozygous p.R592FsX2 mutation while the father harboring the heterozygous mutation. (c) Sanger sequencing analysis of sporadic RP patient ARR-P-S-I-46 showed the homozygous frameshift mutation p.R592FsX2. A full color version of this figure is available at the *Journal of Human Genetics* journal online.

Table 2 Genotype of the family members

Family	Family Member	Gender	Relationship	Clinical information			Mutation(s)	
				Disease onset (age)	Visual acuity (OD, OS)	Systemic disease	Nucleotide change	Genotype
RP-252	IV:1	F	Mother	29	13/20, 3/400	No	p.R592Fs2X	Homozygous
	IV:2	M	Husband	No eye disease	20/20, 20/20	No	p.R592Fs2X	Heterozygous
	V:2	M	Son	No eye disease	20/20, 20/20	No	p.R592Fs2X	Heterozygous
RP-182	IV:1	M	Son	Since birth	7/20, 7/20	No	p.R592Fs2X	Homozygous
	III:4	M	Father	No eye disease	20/20, 20/20	No	p.R592Fs2X	Heterozygous

Abbreviations: F, female; M, male; OD, oculus dexter; OS, oculus sinister.

Organism				
H.sapiens	572	LAQISKSRVKCLRKSEKERMREYQRELEEREKLRPLLFERVAQKNAR		621
P.troglodytes	572	LAQISKSRVKCLRKSEKERMREYQRELEEREKLRPLLFERVAQKNAR		621
M.mulatta	528	-----RKSEKERMREYQRELEEREKLRPLLFERVAQKNAR		565
C.lupus	574	LAQMSKSRVKSRLRKSEKERMREYQRELEEREKLRPLLFERVAQKNAR		623
B.taurus	579	LAQMSKSRIKALRKSEKERMREYRRELEEREKLRPLLFERVAQKNAR		628

p.R592Fs

Figure 4 Protein alignment showed conservation of residues of *FAM161A* p.R592FsX2 across five species. This mutation abolished evolutionarily conserved amino acids. A full color version of this figure is available at the *Journal of Human Genetics* journal online.

mutations in family RP-252 (Figure 3a). In family RP-182, the frameshift mutation p.R592FsX2 was homozygous in the affected son and heterozygous in his unaffected father (Figure 3b). The genotypes of the family members were shown in Table 2.

Sanger sequencing result showed complete co-segregation of the mutations with the disease phenotype in the two consanguineous Indian families. We then screened these variants in 100 sporadic Indian RP patients by whole-exome sequencing. The homozygous frameshift mutation p.R592FsX2 was identified in one sporadic patient ARRP-S-I-46 (figure 3c). This mutation described above was absent in 1000 ethnicity-matched control samples screened by direct Sanger sequencing. In summary, we totally analyzed 14 India families with arRP and 100 sporadic Indian ARRP patients and we found two pedigrees with the *FAM161A* p.R592FsX2 homozygous mutation and one sporadic patient with this mutation. Thus, the frequency of *FAM161A* mutations in Indian RP patients from our result is similar to that of Israel and the Palestinian territories. These data, together with the clinical information, demonstrated that homozygous frameshift mutation p.R592FsX2 in the gene *FAM161A* was responsible for these three arRP patients from Indian.

The mutation p.R592FsX2 in the gene *FAM161A* introduced a frameshift, according to GenBank accession number NM_001201543.1. As shown in Figure 4, the amino acid changes affected highly conserved residues in five species. Therefore, the novel frameshift mutation was likely to affect the *FAM161A* protein function.

DISCUSSION

RP is an inherited disease of the retina, which leads to vision impairment due to progressive photoreceptor cell death. There are more than 50 known disease genes responsible for RP and there are still many new RP disease genes yet to be identified. For the mapping of new loci and subsequent identification of new genes for all inherited forms of RP, the whole-exome sequencing has been shown to be a powerful method. In this study, we found a novel homozygous frameshift mutation p.R592FsX2 in the known RP gene *FAM161A*

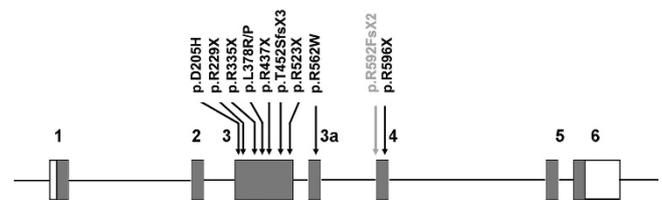


Figure 5 Localization of currently known RP-associated mutations in *FAM161A* gene. The novel p.R592FsX2 mutation found in this study is shown in red. Blue bars indicate coding exons or exon segments whereas white bars represent the 5' and 3' untranslated region (UTR), respectively. A full color version of this figure is available at the *Journal of Human Genetics* journal online.

in the Indian population with arRP using the whole-exome sequencing technology.

The novel mutation p.R592FsX2 in *FAM161A* identified in this study introduces a frameshift from exon 4, which truncates 124 amino acids from the *FAM161A* protein (716 amino acids). Therefore, the novel mutation is likely to affect the *FAM161A* protein function, thus resulting in the occurrence of arRP.

FAM161A gene mutation was initially identified in a study of RP patients in 2010.¹² The previous study showed *FAM161A* mutations a major cause of arRP in Israel and the Palestinian territories, probably 12%, meanwhile they appear to be less frequent in other parts of the world, for example, Germany and the USA, about 1.5–3%.^{12,13} In our result, the frequency of *FAM161A* mutations in Indian RP patients from our result is close to that of Israel and the Palestinian territories. Quite a few known mutations cluster in exon 3 of the *FAM161A* gene (Figure 5), which is by far the largest coding exon of the whole gene.^{12,14} These mutations are predicted to result in a truncated polypeptide that lacks the carboxyl-terminus of the genuine protein, therefore only little of the truncated protein is expected to be synthesized.¹⁵ *FAM161A*-associated retinal disease displays typical

features of arRP but shows a wide phenotypic variability in terms of age of onset and the severity of clinical symptoms.¹⁴

The human *FAM161A* gene consists of seven exons and spans ~30 kb of genomic DNA on chromosome 2p15 with two proteins coding transcript variants, 660 amino acids (76 kDa) and 716 amino acids (83 kDa), respectively. Recent studies have shown that *FAM161A* localizes to the connecting cilium, the basal body and the adjacent centriole in mammalian photoreceptors. *FAM161A* was also present in synaptic layers and ganglion cells of the retina,^{12,14} suggesting that this molecule participates in the maintenance of microtubule tracks. In addition, *FAM161A* directly binds to microtubules and increases the acetylation of α -tubulin and execution of microtubule-dependent functions in the non-photoreceptors.¹⁶ Moreover, *FAM161A* was found post-synaptic to the ribbon synapses in retinal second-order neurons similar to some IFT complex B proteins and the KIF17 motor.^{17,18} Taken together, loss of *FAM161A* function results in mislocalization and accumulation of cargo vesicles in the inner segment as well as disturbed synaptic transmission, eventually causing retinal cell degeneration.

In this report, we identified a novel homozygous frameshift mutation p.R592FsX2 in the Indian population affected by RP disease. This study enriched the mutation spectrum of *FAM161A* gene for RP disease and confirmed that a loss of *FAM161A* function leads to RP disease in India population.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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Supplementary Information accompanies the paper on Journal of Human Genetics website (<http://www.nature.com/jhg>)