Total syntheses of codonopsinine and 4-*epi*-codonopsinine via gold-mediated tandem-catalyzed pyrrole synthesis

Minami Yamaguchi, Daichi Itagaki, Hirofumi Ueda and Hidetoshi Tokuyama

The total syntheses of codonopsinine (1) and 4-*epi*-codonopsinine (2) were accomplished. The key substituted pyrrole intermediate was constructed via gold-catalyzed addition-cyclization cascade of an aminoacetaldehyde acetal derivative and a terminal alkyne. After diastereoselective reduction of the pyrrole intermediate to the corresponding 3-pyrroline derivative with zinc dust and sulfonic acid, the total synthesis of 4-*epi*-codonopsinine (2) was achieved via stereoselective construction of the diol by dihydroxylation. In addition, the total synthesis of codonopsinine (1) was completed through stereochemical inversion of the hydroxyl group via epoxide and subsequent ring cleavage under the acidic aqueous condition. *The Journal of Antibiotics* (2016) **69**, 253–258; doi:10.1038/ja.2016.13; published online 17 February 2016

INTRODUCTION

Codonopsinine, isolated from Codonopsis clematidea by Matkhalikova and colleagues in 1969,¹⁻⁵ has a 1,2,3,4,5-pentasubstituted pyrrolidine ring bearing a *p*-anisyl group at the C2 position. Two decades after its isolation, its structure was revised through a total synthesis of 1 by Kibayashi and colleagues.^{6,7} Codonopsinine has garnered considerable attention from organic chemists because of its substantial biological activities, which include antibiotic activity and hypotensive pharmacological activity.8 4-epi-Codonopsinine, by contrast, exhibits inhibitory activity against α-fucosidase.⁹ Furthermore, compound 1 and its synthetic analogs were recently demonstrated to possess anti-methicillin-resistant Staphylococcus aureus activity.¹⁰ Because of these important biological activities, the development of new anti-infective agents through structure-activity relationships of these compounds is intriguing. However, the divergent synthesis of this class of compounds would be difficult because of the lack of an efficient and versatile synthetic route.11-23

Recently, we developed a method for the synthesis of substituted pyrroles via gold-catalyzed addition–cyclization cascade (Scheme 1).^{24,25} This reaction proceeds via gold-mediated auto-tandem catalysis,²⁶ where the initially generated gold acetylide 5^{27} adds to oxonium ion **6** to give intermediate **7**, which cyclizes in 5-*endo-dig* manner by π -activation with the cationic gold catalyst. An advantage of this methodology is its versatility for the synthesis of substituted pyrroles **9**. Thus a variety of substituted pyrroles can be synthesized in a modular manner by changing two fragments. We envisioned that this methodology would be effective for constructing the pyrrolidine core of codonopsinine and its analogs. Herein we describe the total

syntheses of codonopsinine (1) and 4-*epi*-codonopsinine (2) featuring our gold-mediated tandem-catalyzed pyrrole synthesis.

RESULTS AND DISCUSSION

Our retrosynthetic analysis of codonopsinine (1) and 4-*epi*codonopsinine (2) is shown in Scheme 2. Compounds 1 and 2 could be derived from the common 3-pyrroline intermediate 10 via dihydroxylation of the olefin. 3-Pyrroline derivative 10 would be obtained from the corresponding pyrrole 12 by diastereoselective reduction. We planned to synthesize 5-alkyl-2-*p*-anisyl pyrrole 12 via our gold-catalyzed addition–cyclization cascade using 4-methoxyphenylacetylene 13 and *N*-benzoyl 2-aminopropanal acetal 14. The acetal segment 14 bearing a methyl group should be readily prepared from (L)-alanine (15). On the basis of our previous studies²⁴ on the scope of the substituted pyrrole synthesis by this cascade reaction, we selected diisopropyl acetal 14 as a suitable substrate for construction of the methyl-substituted pyrrole 12.

First, we prepared *N*-benzoyl 2-aminopropanal diisopropyl acetal **14** according to the procedure established in our previous study²⁴ (Scheme 3). Our preliminary studies revealed that the benzoyl protected substrate provided better yield than that of the substrate with carbamate group. Therefore, after conversion of *N*-Cbz alaninal **16**²⁸ to the corresponding diisopropyl acetal by heating with pyridinium p-toluenesulfonate (PPTS) in *i*-PrOH, the Cbz group was replaced with a benzoyl group in a two-step sequence to give the requisite acetal segment **14** in good yield. Then we conducted an optimization study of the gold-catalyzed addition–cyclization cascade to maximize the yield of pyrrole **12** (Table 1). The previously established conditions of 10 mol% of RuphosAuCl²⁹ and AgOTf in

Graduate School of Pharmaceutical Sciences, Tohoku University, Sendai, Japan

Correspondence: Professor H Tokuyama, Graduate School of Pharmaceutical Sciences, Tohoku University, 6-3 Aoba, Aramaki, Aoba-ku, Sendai 980-8578, Japan.

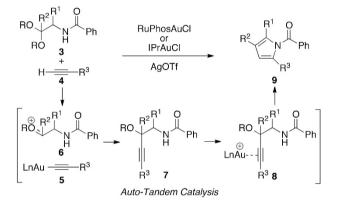
E-mail: tokuyama@m.tohoku.ac.jp

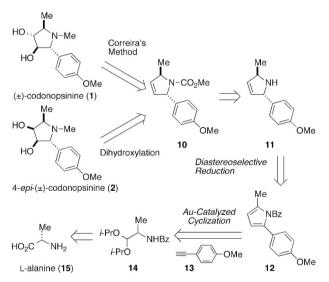
This paper is dedicated to Professor Amos B Smith III on the occasion of his 70th birthday.

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toluene at 110 °C provided the desired pyrrole **12** in 55% yield (entry 1). We observed that the choice of silver salt is important (entries 1–4) and that the yield of **12** was slightly improved when AgBF₄ was used (entry 4). Eventually, the desired pyrrole **12** was obtained in highest yield (84%) by treatment of a mixture of **13** and **14** with 10 mol% of RuphosAuCl and AgBF₄ in xylene (0.25 M) at 140 °C.

Having successfully constructed the 1,2,5-trisubstituted pyrrole intermediate **12**, we then focused on manipulating the pyrrole skeleton to complete the total syntheses of the target natural products. After removing the benzoyl group by heating **12** with aqueous KOH in a mixture of EtOH and CH_2Cl_2 (Scheme 4), we subjected the resulting





Scheme 2 Retrosynthetic analysis of codonopsinine (1) and its C-4 epimer (2).

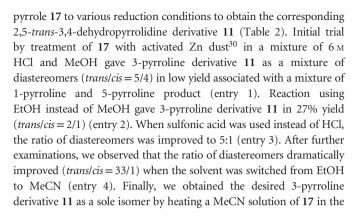
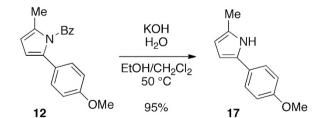


Table 1 Optimization of the pyrrole synthesis by gold-catalyzed addition–cyclization sequence

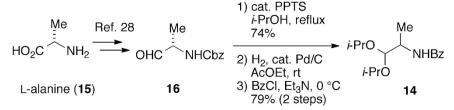
i-PrO i-PrO i-PrO 14		$ \begin{array}{c} & & Me \\ & & & \\ \hline & & \\ 13 \end{array} \begin{array}{c} & OMe \\ (5.0 \text{ eq}) \end{array} \begin{array}{c} & & \\ & & & \\ & & \\ & & \\ & & \\ & & & \\ & & \\ & & \\ & & & \\ & & \\ & & \\ &$				
		RuPhosAuCl ^{a)} (10 mol%) AgX (10 mol%) conditions	12	OMe		
Entry	AgX	Solvent (0.25 м)	Temp (°C)	Yield (%)		
1	AgOTf	Toluene	110	55		
2	AgSbF ₆	Toluene	110	34		
3	AgNTf ₂	Toluene	110	41		
4	AgBF ₄	Toluene	110	62		
5	$AgBF_4$	Xylene	140	84		



RuPhosAuCl

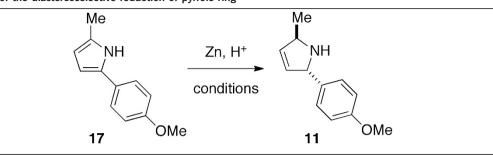


Scheme 4 Hydrolysis of benzoyl pyrrole 12.



Scheme 3 Synthesis of key substrate 14.

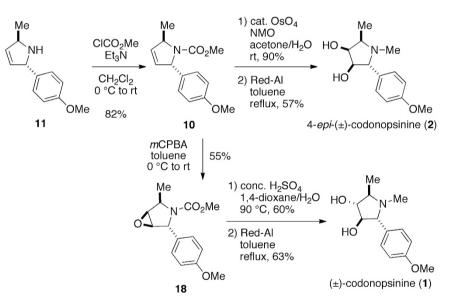
Table 2 Examination of the diastereoselective reduction of pyrrole ring



Entry	Zn (equiv)	Acid	Solvent (0.1 м)	Temp (°C)	Yield (%)	<i>dr^a (</i> trans:cis)
1	8	HCI aq.	MeOH	-6 to RT	22	5:4
2	8	HCI aq.	EtOH	-6 to RT	27	2:1
3	9	Conc. H ₂ SO ₄	EtOH	RT	34	5:1
4	9	Conc. H ₂ SO ₄	MeCN	RT	45	33:1
5	9	Conc. H_2SO_4	MeCN	70	57	Sole isomer

Abbreviation: RT, room temperature.

^aThe ratio of diastereomers was determined by ¹H-NMR.



Scheme 5 Endgame of total syntheses of codonopsinine (1) and 4-epi-codonopsinine (2).

presence of Zn dust and sulfonic acid at 70 °C. The plausible mechanisms would be initiated by protonation of the pyrrole ring at the 5-position to give α , β -unsaturated iminium ion intermediate. Then the intermediate would be reduced by single-electron reduction with zinc dust to generate a stable 2,5-*trans* radical species, which would be further reduced by zinc reagent to provide the *trans* product after protonation.

The endgame sequence of the total synthesis of codonopsinine (1) and 4-*epi*-codonopsinine (2) is depicted in Scheme 5. First, methoxycarbonylation of secondary amine of 11 produced the pivotal intermediate 10. The total synthesis of 4-*epi*-codonopsinine (2) was achieved by diastereoselective dihydroxylation of 10 and reductive transformation of methyl carbamate to the corresponding methyl amine. By contrast, conversion of 10 to codonopsinine (1) was executed by diastereoselective meta-3-chloroperoxybenzoic acid (*mCPBA*) epoxidation of the olefin, ring cleavage of the epoxide under acidic conditions and reduction with Red-Al according to the

procedure¹³ developed by Correira *et al.* All the properties of synthetic 1^{13} and 2^9 were identical to the published data.

In conclusion, we accomplished the total syntheses of codonopsinine (1) and 4-*epi*-codonopsinine (2). For the construction of the pentasubstituted pyrrolidine core, we utilized our original multisubstituted pyrroles synthesis by gold-mediated auto-tandem catalysis and diastereoselective reduction of the pyrrole ring to the 3-pyrroline derivative. Taking advantage of our modular synthesis of substituted pyrroles, the synthetic strategy developed in this work should be applicable to the versatile synthesis of a wide range of codonopsinine congeners.

EXPERIMENTAL PROCEDURE Gereral remarks

Unless otherwise noted, all reactions were performed using oven-dried glassware, sealed with a rubber septum under a slight positive pressure of argon. Anhydrous tetrahydrofuran, MeCN, 1,4-dioxane and dichloromethane

255

256

were purchased from Kanto Chemical Co., Inc. (Tokyo, Japan) Anhydrous toluene, xylene, acetone and dimethylformamide were purchased from Wako Pure Industries (Osaka, Japan). Anhydrous EtOH, MeOH and Et₃N were dried and distilled according to the standard protocols. Unless otherwise mentioned, materials were obtained from commercial suppliers and used without further purification. Microwave irradiation experiments were performed on a CEM Discover Microwave Reactor (Discover-SP W/ACTIVENT, Tokyo, Japan). Reactions and chromatographical fractions were monitored by TLC analysis with precoated silica gel plates 60 F254 (Merck, Frankfurt, Germany). Flash column chromatography was carried out using Kanto silica gel 60N (spherical, neutral, 40-50 µm). Preparative TLC was performed on precoated silica gel plates 60 F254 (Merck). Gel permeation chromatography was carried out using a Japan Analytical Industry Co., Ltd., LC-9201 (Tokyo, Japan). IR spectra were measured on a Shimadzu FTIR-8300 spectrometer (Kyoto, Japan) or a JASCO FT/IR-4100 spectrometer (Tokyo, Japan). NMR spectra were measured on a JNM-AL400 spectrometer (JEOL Resonance Inc., Tokyo, Japan). For ¹H spectra, chemical shifts were expressed in p.p.m. downfield from internal tetramethylsilane (δ 0) or relative internal CHCl₃ (δ 7.26). For ¹³C spectra, chemical shifts were expressed in p.p.m. downfield from relative internal CHCl₃ (δ 77.0). J values were expressed in Hertz. Elemental analyses were performed by a Yanaco CHN corder MT-6. Mass spectra (Kyoto, Japan) were recorded on a JEOL JMS-DX-303, a JMS-700, a JMS-T100GC (respectively, JEOL Ltd., Tokyo, Japan) and a Bruker micrOTOF spectrometer (MicrOTOF II-TH, Bruker Daltonics, Yokohama, Japan).

N-Carboxybenzylalaninal diisopropylacetal

To a solution of aldehyde **16** (1.23 g, 5.94 mmol) in *i*-PrOH (40 ml) was added PPTS (150 mg, 0.597 mmol). After stirring at reflux for 18 h with Dean–Stark trap containing MS4 Å, the reaction was quenched with saturated aqueous NaHCO₃ and the mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexanes/AcOEt=9:1) to afford the titled compund (1.35 g, 4.37 mmol, 74%) as a colorless oil; R_f =0.69 (silica gel, hexanes/AcOEt=7:3); IR (neat) 3344, 2972, 1715, 1504, 1454, 1332, 1229, 1053, 1028 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.41–7.28 (m, 5H), 5.17–5.03 (m, 2H), 5.02–4.88 (m, 1H), 4.74 (s, 1H), 3.91–3.70 (m, 3H), 1.24–1.08 (m, 15H); ¹³C NMR (100 MHz, CDCl₃) δ 156.0, 136.6, 128.5, 128.04, 128.01, 99.6, 69.5, 69.0, 66.5, 49.5, 23.00, 22.98, 22.4, 22.1, 14.6; LRMS (FAB) *m/z* 310.2 ([M+H]⁺); HRMS Calcd for C₁₇H₂₈NO₄ ([M+H]⁺) 310.2019, found 310.2008.

N-Benzoylalaninal diisopropylacetal (14)

A suspension of the above diisopropyl acetal (1.33 g, 4.29 mmol) and 10% Pd/C (457 mg, 0.429 mmol) in EtOH (42 ml) was stirred under a hydrogen atmosphere at room temperature for 1.5 h. The reaction mixture was filtered through a pad of Celite and the filtrate was concentrated under reduced pressure. The residue was dissolved in CH_2Cl_2 (13 ml), and Et_3N (1.69 ml, 12.1 mmol) and benzoyl chloride (510 µl, 4.40 mmol) were added at 0 °C. After stiring for 10 min, the reaction was quenched with saturated aqueous NH₄Cl and the mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na2SO4, filtered and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexanes/AcOEt=8:2) to afford benzamide 14 (877 mg, 3.14 mmol, 73%) as a white solid; $R_f = 0.33$ (silica gel, hexanes/AcOEt = 7:3); Mp 89-90 °C; IR (KBr) 3292, 2970, 2361, 1636, 1558, 1377, 1339, 1159, 1047, 916, 693 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, J=8.0 Hz, 2H), 7.50 (t, J=8.0 Hz, 1H), 7.43 (dd, J=8.0, 8.0 Hz, 2H), 6.34 (br s, 1H), 4.59 (d, J = 2.4 Hz, 1H), 4.36–4.26 (m, 1H), 3.88 (sep, J = 6.0 Hz, 1H), 3.83 (sep, J = 6.0 Hz, 1H), 1.32–1.17 (m, 12H), 1.13 (d, J = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.8, 134.7, 131.3, 128.5, 126.8, 99.5, 69.6, 69.0, 48.0, 23.01, 22.99, 22.5, 22.2, 14.2; Anal Calcd for C16H25NO3: C, 68.79; H, 9.02; N, 5.01. Found: C, 68.74; H, 9.26; N, 5.07.

1-Benzoyl-2-(4-methoxyphenyl)-5-methyl-1H-pyrrole (12)

A two necked round-bottom flask equipped with a magnetic stirring bar was charged with AgBF₄ (18.1 mg, 92.8 µmol), RuPhosAuCl (64.9 mg, 92.8 µmol) and xylenes (3.7 ml, 0.25 M). To the solution were added benzamide 14 (260 mg, 0.928 mmol) and 1-ethynyl-4-methoxybenzene 13 (0.700 ml, 4.64 mmol) at room temperature. After stirring at 140 °C for 10 min, the reaction was quenched with saturated aqueous NaHCO3, and the mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na2SO4, filtered and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexanes/toluene = 1:3) and preparative TLC on silica gel (hexanes/ AcOEt = 9:1) to afford N-benzoyl pyrrole 12 (226 mg, 0.776 mmol, 84%) as a pale yellow solid; $R_f = 0.40$ (silica gel, hexanes/toluene = 1:3); Mp 103–104 °C (hexanes/CH2Cl2); IR (KBr) 1697, 1531, 1493, 1333, 1290, 1242, 1032, 833, 802, 700 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.56 (d, J=7.4 Hz, 2H), 7.40 (t, J=7.2 Hz, 1H), 7.33-7.20 (m, 2H), 7.09 (d, J=8.8 Hz, 2H), 6.66 (d, J=8.8 Hz, 2H), 6.21 (d, J=3.6 Hz, 1H), 6.10 (d, J=3.6 Hz, 1H), 3.71 (s, 3H), 2.34 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.6, 158.1, 135.1, 135.0, 133.2, 132.4, 130.4, 128.8, 128.2, 126.4, 113.6, 110.2, 110.0, 55.1, 14.0; Anal Calcd for C19H17NO2: C, 78.33; H, 5.88; N, 4.81. Found: C, 78.07; H, 5.93; N. 4.78.

2-(4-Methoxyphenyl)-5-methyl-1H-pyrrole (17)

To a solution of *N*-benzoyl pyrrol **12** (225 mg, 0.774 mmol) in a mixture of EtOH (2.6 ml), CH₂Cl₂ (1.3 ml) and H₂O (0.7 ml) was added KOH (195 mg, 3.48 mmol). After stirring for 2 h at 50 °C, the mixture was concentrated under reduced pressure. The residue was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexanes/AcOEt = 8:2) to afford pyrrol **17** (138 mg, 0.737 mmol, 95%) as a light yellow solid; Mp 129–131 °C; IR (neat) 3433, 3398, 2924, 1525, 1254, 824, 787 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.01 (br s, 1H), 7.35 (d, *J* = 8.8 Hz, 2H), 6.89 (d, *J* = 8.8 Hz, 2H), 6.27 (dd, *J* = 3.4, 2.8 Hz, 1H), 5.93 (m, 1H), 3.81 (s, 3H), 2.32 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.8, 130.8, 128.3, 126.1, 124.8, 114.3, 107.6, 105.0, 55.3, 13.1; ESI-MS *m*/*z* Calcd for C₁₂H₁₄NO 188.1060 (M⁺+H), found 188.1070.

(2S*,5R*)-2-(4-Methoxyphenyl)-5-methyl-2,5-dihydro-1*H*-pyrrole (11)

To a suspension of pyrrole 17 (117 mg, 0.628 mmol) and zinc dust (activated, 367 mg, 5.61 mmol) in MeCN (6.3 ml) was added dropwise concentrated H₂SO₄ (0.402 ml). After stirring at 70 °C for 10 min, the mixture was filtered through a pad of Celite. The pH value of the filtrate was adjusted to 10 by addion of 2 M NaOH. The solution was saturated with NaCl and the mixture was extracted with AcOEt five times. The combined organic extracts were dried over Na₂SO₄, filtered and concentrated under reduced pressure to give a crude product as a single isomer. The crude material was purified by flash column chromatography on silica gel (CH₂Cl₂/MeOH = 10:1) to afford 3-pyrroline derivative 11 (67.2 mg, 0.355 mmol, 57%) as a pale yellow oil. $R_f = 0.28$ (Silica gel, MeOH/CH₂Cl₂ = 1:10); IR (neat) 2959, 2835, 1611, 1510, 1244, 1175, 1036 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.21 (d, J = 8.8 Hz, 2H), 6.86 (d, J = 8.8 Hz, 2H), 5.90 (d, J = 6.0 Hz, 1H), 5.80 (d, J = 6.0 Hz, 1H), 5.14 (br s, 1H), 4.37–4.26 (m, 1H), 3.79 (s, 3H), 2.47 (br s, 1H), 1.26 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.8, 136.4, 134.2, 131.3, 127.9, 113.9, 68.2, 60.3, 55.2, 22.7; ESI-MS m/z Calcd for C12H16NO 190.1226 (M++H), found 190.1235.

$(2S^*,5R^*)$ -2-(4-Methoxyphenyl)-5-methyl-2,5-dihydro-1*H*-pyrrole-1-carboxylic acid methyl ester (10)

To a solution of 3-pyrroline derivative **11** (40.2 mg, 0.212 mmol) and Et₃N (89 μ l, 0.64 mmol) in CH₂Cl₂ (0.7 ml) was added methyl chloroformate (20 μ l, 0.26 mmol) at 0 °C. After stirring for 2 h at room temperature, the reaction was quenched with saturated aqueous NH₄Cl, and the mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by preparative TLC on silica gel (hexanes/AcOEt = 3:7) to afford methyl carbamate **10** (43.1 mg, 0.174 mmol, 82%) as a clear oil; its ¹H

NMR spectral data were identical with those reported¹³; ¹H NMR (400 MHz, CDCl₃, a mixture of rotamers) δ 7.18 (d, *J*=8.8 Hz, 0.9H), 7.08 (d, *J*=8.8 Hz, 1.1H), 6.86 (d, *J*=8.8 Hz, 0.9H), 6.82 (d, *J*=8.8 Hz, 1.1H), 5.80–5.73 (m, 1H), 5.64 (ddd, *J*=6.4, 2.0, 0.5 Hz, 0.5H), 5.61 (ddd, *J*=6.4, 2.0, 0.5 Hz, 0.5H), 5.64 (m, 0.5H), 4.78–4.70 (m, 0.5H), 3.79 (s, 1.6H), 3.78 (s, 1.4H), 3.63 (s, 1.4H), 3.42 (s, 1.6H), 1.44 (d, *J*=6.0 Hz, 1.4H).

(2*R**,3*R**,4*S**,5*R**)-3,4-Dihydroxy-2-(4-methoxyphenyl)-5methylpyrrolidine-1-carboxylic acid methyl ester

To a solution of 3-pyrroline derivative $10~(15.2\mbox{ mg},~61.5\mbox{ }\mu mol)$ and Nmethylmorpholine N-oxide (NMO) (10.8 ml, 92.3 µmol) in acetone (0.50 ml) and H₂O (0.12 ml) was added OsO₄ (1.0% in H₂O, 78 µl, 3.1 µmol). After stirring for 5 days at room temperature, the reaction was quenched with saturated aqueous NaHSO3, and the mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by preparative TLC on silica gel (hexanes/AcOEt = 1:1) to afford the titled *cis*-diol (15.6 mg, 55.5 μ mol, 90%) as a colorless oil; $R_f = 0.20$ (silica gel, hexanes/AcOEt = 1:1); IR (neat) 3419, 3303, 2937, 1680, 1612, 1513, 1455, 1386, 1249 cm⁻¹; ¹H NMR (400 MHz, CD₃CN, 60 °C) δ 7.08–7.00 (m, 2H), 6.92-6.80 (m, 2H), 4.72 (br s, 1H), 4.29-4.18 (m, 1H), 4.17-4.09 (m, 1H), 3.86 (br s, 1H), 3.78–3.72 (m, 3H), 3.67–3.29 (m, 4H), 3.14 (br s, 1H), 1.35–1.33 (m, 1H); ¹³C NMR (100 MHz, CD₃CN, 60 °C) δ 160.1, 156.4, 130.3, 127.9, 115.1, 114.1, 70.9, 68.6, 64.3, 57.2, 56.2, 52.5; ESI-MS m/z Calcd for C14H19NNaO5 304.1155 (M++Na), found 304.1157.

$(2R^{*}\!,\!3R^{*}\!,\!4S^{*}\!,\!5R^{*}\!)\text{-}2\text{-}(4\text{-}Methoxyphenyl)\text{-}1,\!5\text{-}dimethylpyrrolidine-3,4\text{-}diol;}$ 4-epi-(\pm)-codonopsinine (2)

To a solution of the above *cis*-diol (12.1 mg, 43.0 µmol) in toluene (0.22 ml) was added Red-Al (65% w/v in toluene, 129 µl) at 0 °C. After heating at reflux for an hour, the reaction was quenched with saturated aqueous Rochell's salt at 0 °C. The resulting mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by preparative TLC on silica gel (CHCl₃/MeOH=3:1) to afford 4-*epi*-(\pm)-codonopsinine (**2**) (5.8 mg, 24.4 µmol, 57%) as a white solid; its ¹H NMR spectral data were identical with those reported⁹; ¹H NMR (400 MHz, CDCl₃) δ 7.21 (d, *J*=8.4 Hz, 2H), 6.88 (d, *J*=8.4 Hz, 2H), 4.28 (dd, *J*=6.4, 6.4 Hz, 1H), 4.06 (dd, *J*=6.4, 5.2 Hz, 1H), 3.81 (s, 3H), 3.66 (d, *J*=5.2 Hz, 1H), 3.53–3.45 (m, 1H), 2.64 (br s, 1H), 1.12 (s, 3H).

$(1S^*, 2S^*, 4S^*, 5R^*)$ -2-(4-Methoxyphenyl)-4-methyl-6-oxa-3-azabicyclo[3.1.0]hexane-3-carboxylic acid methyl ester (18)

To a solution of pyrroline **10** (14.7 mg, 59.4 µmol) in toluene (0.3 ml) was added *m*CPBA (51.0 mg, 0.297 mmol) at 0 °C. After stirring at room temperature for 28 h, the reaction was quenched with saturated aqueous NaHCO₃, and the resulting mixture was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by preparative TLC on silica gel (hexanes/AcOEt=3:7) to afford epoxide **18** (8.6 mg, 33 µmol, 55%) as a clear oil; its ¹H NMR spectral data were identical with those reported¹⁴, ¹H NMR (400 MHz, CDCl₃) δ 7.14 (d, *J*=8.4 Hz, 0.9H), 7.08 (d, *J*=8.8 Hz, 2.1H), 6.92–6.84 (m, 2H), 5.04 (br s, 0.3H), 4.96 (br s, 0.7H), 4.22–4.07 (m, 1H), 3.80 (s, 2.1H), 3.79 (m, 0.9H), 3.76–3.70 (m, 1H), 3.62 (s, 0.9H), 3.52–3.47 (m, 1H), 3.44 (s, 2.1H), 1.58 (d, *J*=6.0 Hz, 2.1H), 1.51 (d, *J*=6.0 Hz, 0.9H).

(2*R**,3*R**,4*R**,5*R**)-3,4-Dihydroxy-2-(4-methoxyphenyl)-5methylpyrrolidine-1-carboxylic acid methyl ester

To a solution of epoxide 18 (8.6 mg, 33 μ mol) in 1,4-dioxane (0.13 ml) and H₂O (90 μ l) was added dropwise concentrated H₂SO₄ (9.0 μ l). After stirring for 9 h at 90 °C, the reaction was quenched with saturated aqueous NaHCO₃. The resulting mixture was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered and

concentrated under reduced pressure. The residue was purified by preparative TLC on silica gel (hexanes/AcOEt = 2:1) to afford the titled *trans*-diol (5.5 mg, 20 µmol, 60%) as a clear oil; its ¹H NMR spectral data were identical with those reported¹³; ¹H NMR (400 MHz, C₅D₅N, a mixture of rotamers) δ 7.65 (d, *J* = 7.6 Hz, 1H), 7.45 (br s, 1H), 6.93 (d, *J* = 8.4 Hz, 2H), 4.66 (br s, 1H), 4.61–4.33 (m, 2H), 3.69 (br s, 1H), 3.60 and 3.59 (s, 3H), 3.48 (s, 3H), 1.93 (d, *J* = 6.4 Hz, 1.8H), 1.75 (d, *J* = 6.4 Hz, 1.8H).

$(2R^*, 3R^*, 4R^*, 5R^*)$ -2-(4-Methoxyphenyl)-1,5-dimethylpyrrolidine-3,4-diol; codonopsinine (1)

To a solution of *trans*-diol compound (8.5 mg, 30.2 µmol) in toluene (0.15 ml) was added Red-Al (65% w/v in toluene, 233 µl) at 0 °C. After heating at reflux for 45 h, the reaction was quenched with saturated aqueous Rochell's salt at 0 °C, and the mixture was extracted with AcOEt three times. The combined organic extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by preparative TLC on silica gel (CHCl₃/MeOH=4:1) to afford (\pm)-codonopsinine (1) (4.5 mg, 19 µmol, 63%) as a white solid; Mp 155–157 °C; IR (neat) 3365, 2919, 2837, 1611, 1514, 1459, 1249, 1180, 1035, 837 cm⁻¹; ¹H NMR (600 MHz, C₅D₅N) δ 7.58 (d, *J*=8.4 Hz, 2H), 6.97 (d, *J*=8.4 Hz, 2H), 4.60 (dd, *J*=6.0, 4.8 Hz, 1H), 4.36 (dd, *J*=6.0, 4.8 Hz, 1H), 4.02 (d, *J*=6.0 Hz, 1H), 3.67 (qd, *J*=6.4, 3.6 Hz, 1H), 3.66 (s, 3H), 2.20 (s, 3H), 1.31 (s, *J*=6.4 Hz, 3H); ¹³C NMR (150 MHz, C₅D₅N) δ 159.0, 134.7, 129.5, 113.8, 86.9, 84.7, 74.0, 64.7, 54.8, 34.4, 13.6; ESI-MS *m*/*z* Calcd for C₁₃H₂₀NO₃ (M⁺+H) 238.1438, found 238.1435.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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