COMMUNICATION TO THE EDITOR

In vitro and in vivo anti-Trypanosoma brucei activities of phenazinomycin and related compounds

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During the course of our screening program to discover new antitrypanosomal compounds, we have evaluated isolates from soil microorganisms as well as compounds from the antibiotic libraries of the Kitasato Institute for Life Sciences and Bioscience Associates. We have previously reported on various microbial metabolites exhibiting potent anti-Trypanosoma brucei properties, which are defined as antitrypanosomal properties.^{1,2} We have recently evaluated the known phenazine antibiotics, phenazinomycin, griseoluteic acid, griseolutein B and related compounds, some of which have shown significant antitrypanosomal activities, both in vitro and in vivo. Here, we report the antitrypanosomal profiles of phenazinomycin and some related compounds (Figure 1) in comparison with those of some clinically used antitrypanosomal drugs, such as suramin and eflornithine. We also present our findings with respect to structure-activity relationships.

Griseoluteic acid, griseolutein B, phenazinomycin and 1,6-dimethoxyphenazine were obtained from the antibiotic libraries of the Kitasato Institute for Life Sciences and Bioscience Associates. Related compounds, namely clinically used clofazimine and phenazine, were purchased from Sigma (Sigma-Aldrich, St Louis, MO, USA), and phenazine 5-oxide was donated by Meiji Seika Kaisha (Yokohama, Japan).

In vitro antitrypanosomal activities against *Trypanosoma brucei brucei* strain GUTat 3.1 (low virulence strain against mice) and cyto-toxicity against human diploid embryonic cell line MRC-5 were measured, as described previously.^{1,3} *In vivo* antitrypanosomal activity for *T. b. brucei* strain S427 (high virulence strain against mice) was measured as described previously.² Test compounds were solubilized in an aqueous mixture of 10% DMSO-Tween 80 and EtOH (7:3) and admi-

nistered i.p. to mice on the next day (day 1) following infection with parasites (day 0). Subsequently, the compounds were successively administered (i.p.) to the infected mice once a day for 3 days (days 2–4). Efficacies of the compounds were determined by parasitaemia levels and the mean of survival days (MSD), and compared with those of the untreated control mice.

Table 1 shows the in vitro antitrypanosomal activities of phenazine compounds and selected standard antitrypanosomal drugs. Griseolutein B showed the highest antitrypanosomal activity, with an IC₅₀ value of 1.4 ng ml^{-1} . The compound was 1100–1600fold more potent than effornithine and suramin. Phenazinomycin was 164-fold less active than griseolutein B, showing an IC₅₀ value of 230 ng ml⁻¹. Griseoluteic acid and clofazimine showed moderate antitrypanosomal activity, with an IC₅₀ value of approximately $2 \mu g m l^{-1}$, which is comparable to those of both eflornithine and suramin, whereas 1,6-demethoxyphenazine, phenazine and phenazine 5-oxide showed no antitrypanosomal activity whatsoever.

The *in vitro* cytotoxicities of phenazine compounds and the antitrypanosomal drugs tested are also presented in Table 1. Griseo-lutein B showed the highest cytotoxicity against MRC-5 cells, with an IC₅₀ value of 9 ng ml⁻¹. Griseoluteic acid and phenazino-mycin were revealed to be slightly cytotoxic, demonstrating IC₅₀ values of $5-6\,\mu$ g ml⁻¹, while clofazimine had an IC₅₀ value of > 100 μ g ml⁻¹ and was shown to be not cytotoxic.

For appropriate evaluation of the combined antitrypanosomal activities and cytotoxicities of test compounds, we introduced selectivity indexes (SI; cytotoxicity (IC₅₀ for the MRC-5 cells)/antitrypanosomal activity (IC₅₀ for the GUTat 3.1 strain)) as presented in Table 1. Among the tested compounds, phenazinomycin and clofazimine showed a medium SI, with ratios of around 23 -> 52, less than or similar to those of effornithine and suramin. Griseoluteic acid and griseolutein B showed a low SI, with ratios of around 3–6. Therefore, we selected phenazinomycin and clofazimine for *in vivo* efficacy tests.

The preliminary in vivo antitrypanosomal activities of phenazinomycin and clofazimine were measured in the T. b. brucei S427 acute mouse model. At a dose of $50 \,\mathrm{mg \, kg^{-1}}$, phenazinomycin did not achieve cure but did extend the MSD to 12.3 days, representing a 2.7-fold increase over control MSD (4.5 days). The same dose of clofazimine did not achieve cure but also extended the MSD to 6.0 days, similar to the control MSD (6.0 days). Under the same conditions, suramin showed a curative effect (MSD: >30 days) at a much lower dose of 1 mg/kg. The phenazinomycin data suggest that it might be a candidate compound for discovering new antitrypanosomal drugs. These efficacy tests, including in vivo antitrypanosomal activity, are ongoing.

1,6-Dimethoxyphenazine, phenazine and phenazine 5-oxide showed no antitrypanosomal activity. The most potent antitrypanosomal activity of griseolutein B and the moderate antitrypanosomal activity of griseoluteic acid and clofamizine, in comparison with phenazinomycin, provide very interesting information about structure-activity relationships. Griseoluteic acid lacks the substituent at N-5 and is 1450- and 9-fold less bioactive than griseolutein B and phenazinomycin, respectively. Furthermore, phenazinomycin possesses a substituent containing a sesquiterpene moiety at N-5, and has a medium SI compared with griseoluteic acid and griseolutein B. Our data therefore suggest

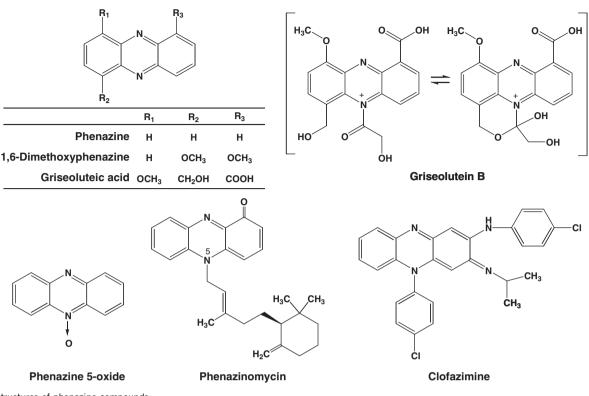


Figure 1 Structures of phenazine compounds.

that the substituent at N-5 present in phenazinomycin bestows significant antitrypanosomal activity and cytotoxicity.

Further studies are necessary for an extensive structure–*in vitro* antitrypanosomal activity evaluation of phenazinomycin-related compounds.

Griseoluteic acid (LL-14I352beta) is known to have antitumor and weak antibacterial activity, through its mode of action in inhibition of DNA synthesis.⁴ Griseolutein B represents an equilibrium between the ketone form and the cyclic hemiacetal form (Figure 1),⁵ and is reported to have antibacterial and antitumor activity.^{6,7} We previously reported that phenazinomycin has antibacterial and antitumor activity,8 although the mode of action is not clearly understood. However, a related antibiotic, lavanducyanin (WS-9659 A), containing a cyclohexenylmethyl moiety at N-5, is reported to have antitumor activity and to inhibit testosterone 5\alpha-reductase in rat, dog and human prostates.9,10 The mode of action of lavanducyanin is believed to be via disruption of signal transduction in cell growth.¹¹ Clofazimine, an anti-inflammatory and antileprosy drug, is a riminophenazine compound and demonstrates antituberculosis and anti-Mycobacterium avium complex activities, while also being used to treat several diseases, including Table 1 *In vitro* antitrypanosomal activity against *Trypanosoma brucei brucei* GUTat 3.1 and cytotoxicity in MRC-5 cells of phenazine compounds and some commonly used antitrypanosomal drugs

$IC_{50} (ng m l^{-1})$		
Antitrypanosomal activity GUTat 3.1	Cytotoxicity MRC-5	Selectivity index (SI) MRC-5/GUTat 3.1
2030	6110	3
1.4	9	6
230	5190	23
> 12 500	ND ^a	_
1920	>100000	>52
> 12 500	ND ^a	_
> 12 500	ND ^a	_
1580	>100000	>63
2270	>100000	>44
	Antitrypanosomal activity GUTat 3.1 2030 1.4 230 > 12 500 1920 > 12 500 > 12 500 > 12 500 1 2 500 1 2 500 1 580	Antitrypanosomal activity Cytotoxicity GUTat 3.1 MRC-5 2030 6110 1.4 9 230 5190 >12 500 ND ^a 1920 >100 000 >12 500 ND ^a 12 500 ND ^a >12 500 ND ^a 212 500 ND ^a >12 500 ND ^a 1580 >100 000

^aNot determined

leishmaniasis.¹² The modes of action of clofazimine are via inhibition of DNA synthesis and stimulation of phospholipase A_2 .¹² The reported mechanisms of action of gliseoluteic acid, griseolutein B and clofazimine may be replicated against *Trypanosoma* too.

Recently, phenazine compounds have been reviewed extensively, as they are producers of reactive oxygen species, and also with a view to identifying their biochemical communication systems in soil or in mammals with bacterial infections.¹³ However, discovery of the antitrypanosomal activities of griseoluteic acid, griseolutein B, phenazinomycin and clofazimine is novel and our data constitute the first report of such properties.

The above results reveal that phenazinomycin is a candidate compound for development of novel antitrypanosomal drugs. Further study of the antitrypanosomal and other biological activities of phenazinomycin is in progress. This work was supported, in part, by funds from the Drugs for Neglected Diseases initiative (DND*i*), Quality Assurance Framework of Higher Education from the Ministry of Education, Culture, Sports, Science and Technology, Japan (MEXT), and the All Kitasato Project Study (AKPS). We are grateful to Ms H Sekiguchi and Mr T Furusawa for their technical assistance.

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