ORIGINAL ARTICLE

Pentacecilides, new inhibitors of lipid droplet formation in mouse macrophages produced by *Penicillium cecidicola* FKI-3765-1: II. Structure elucidation

Hiroyuki Yamazaki¹, Satoshi Ōmura² and Hiroshi Tomoda¹

The structures of pentacecilides, new inhibitors of lipid droplet formation in mouse macrophages produced by *Penicillium cecidicola* FKI-3765-1, were elucidated by spectroscopic studies, including various NMR experiments. Pentacecilides have a common pentacyclic meroterpene core, which contains an aromatic ring and a δ -lactone ring. *The Journal of Antibiotics* (2009) **62**, 207–211; doi:10.1038/ja.2009.19; published online 20 March 2009

Keywords: pentacecilides; structure elucidation; fungal metabolites; lipid droplet formation

INTRODUCTION

Three new compounds, designated pentacecilides A to C (Figure 1) were isolated as inhibitors of lipid droplet formation in mouse macrophages from the culture broth of *P. cecidicola* FKI-3765-1.¹ The taxonomy of the producing strain, fermentation, isolation and biological properties of pentacecilides were described in an earlier paper.¹ In this study, the physicochemical properties and structure elucidation of pentacecilides are described.

RESULTS

Physicochemical properties

The physicochemical properties of pentacecilides A to C are summarized in Table 1. They have a similar pattern with absorption maxima at 214–219 nm, 273–274 nm and 309–310 nm in UV spectra. IR absorption at 1619–1745 cm⁻¹ and 3401–3434 cm⁻¹ suggested the presence of carbonyl and hydroxy groups in their structures. These data indicated that they share a similar skeleton.

Structure elucidation of pentacecilide C

The molecular formula of pentacecilide C was determined to be $C_{27}H_{34}O_8$ on the basis of HRESI-TOF-MS measurement (Table 1). The ¹³C NMR spectrum (in CDCl₃) showed 27 resolved signals, which were classified into six methyl carbons, four methylene carbons, two sp^3 methine carbons, one sp^2 methine carbon, three oxygenated sp^3 methine carbons, two sp^3 quaternary carbons, one oxygenated sp^3 quaternary carbons, two oxygenated sp^2 quaternary carbons by analysis of the DEPT and heteronuclear single quantum coherence (HSQC) spectra.

The ¹H NMR spectrum (in CDCl₃) displayed 33 proton signals, one of which was suggested to be a hydroxyl proton (δ 11.08), as reported in thailandolides.² Taking the molecular formula into consideration, the presence of another hydroxy proton was suggested. The connectivity of proton and carbon atoms was established by the ¹³C-¹H HSQC spectrum (Table 2). Analyses of ¹H-¹H COSY revealed the presence of partial structures I to IV, as shown in Figure 2. Furthermore, ¹³C-¹H long-range couplings of ²J and ³J observed in the ¹³C-¹H HMBC spectrum gave the following linkages (Figure 3a): (1) Cross-peaks from H₂-7' (\$ 2.70, 2.85) to C-1' (\$ 110.7), C-2' (\$ 139.3) and C-3' (δ 102.2), from OH-4' (δ 11.08) to C-3', C-4' (δ 162.3) and C-5' (δ 103.5) and from H-5' (δ 6.30) to C-1', C-3', C-4' and C-6' (δ 159.1) indicated that a phenol skeleton connects the partial structure I at C-2'. Furthermore, the findings that the chemical shift of C-8' (δ 74.7) corresponds to an oxygenated carbon and the OH-4' proton (δ 11.08) shifted to a lower field because of a hydrogen bonding indicated that C-3' and C-8' are connected through an ester bond, which form δ-lactone. This was also supported by the IR absorption (1619-1666 cm⁻¹). Although observation of a cross-peak from H-8' to C-10' (δ 169.9) was important and simple to show the presence of δ -lactone, the cross-peak was not observed because the dihedral angle between H-8' and C-10' is 90°. Therefore, the coupling constant in HMBC experiment was changed from J_{C-H} =8.0 Hz to J_{C-H} =3.0 Hz. As a result, the long-range coupling of ⁴J from H-5' to C-10' was observed, supporting the presence of δ -lactone. (2) Cross-peaks from H₂-11 (δ 2.52) to C-8 (δ 79.0), from H₃-12 (δ 1.23) to C-7 (δ 71.8), C-8 and C-9 (δ 43.0), from H-7 (δ 4.11) to C-8 and C-9, from H-9 (δ 2.23) to C-8, C-10 (δ 36.2) and C-15 (δ 24.6), from H₂-6

¹Graduate School of Pharmaceutical Sciences, Kitasato University, Shirokane, Minato-ku, Tokyo, Japan and ²Kitasato Institute for Life Sciences, Kitasato University, Shirokane, Minato-ku, Tokyo, Japan

Correspondence: Professor H Tomoda, Graduate School of Pharmaceutical Sciences, Kitasato University, 5-9-1 Shirokane, Minato-ku, Tokyo 108-8641, Japan. E-mail: tomodah@pharm.kitasato-u.ac.jp

Received 15 January 2009; revised 6 February 2009; accepted 19 February 2009; published online 20 March 2009

H3C / , ,







Figure 1 Structures of pentacecilides A to C and thailandolides A and B.

	Pentacecilide A	Pentacecilide B	Pentacecilide C	
Appearance White crystalline solid		White crystalline solid	White crystalline solid	
Molecular weight	412	470	486	
Molecular formula HRESI-TOF-MS (m/z)	$C_{25}H_{32}O_5$	C ₂₇ H ₃₄ O ₇	C ₂₇ H ₃₄ O ₈	
Calcd:	435.2147 (M+Na)+	493.2225 (M+Na)+	509.2151 (M+Na)+	
Found:	435.2141 (M+Na)+	493.2202 (M+Na)+	501.2162 (M+Na)+	
UV (MeOH) λ_{\max} nm (ɛ)	219 (18647), 274 (11846), 309 (4194)	219 (33055), 274 (18226), 309 (4784)	214 (54432), 273 (32736), 310 (4947)	
[α] ²⁶	-4.38° (c=0.38, CHCl ₃)	-32.6° (c=0.68, CHCl ₃)	-32.3° (c=0.48, CHCl ₃)	
IR (KBr) $v_{\rm max}$ (cm ⁻¹)	3401, 1697. 1662, 1465, 1380	3440, 1747, 1727, 1666, 1475	3434, 1745, 1666, 1619, 1473	

(δ 1.86, 2.18) to C-10, from H₃-15 (δ 1.54) to C-9, C-10 and C-1 (δ 41.1), from H-5 (δ 1.82) to C-4 (δ 48.3), C-10 and C-15, from H₂-1 (δ 1.88, 2.20) to C-3 (δ 207.8), C-5, C-10 and C-15, from H₃-13 (δ 1.14) to C-3, C-4, C-5 and C-14 (δ 25.7), from H₃-14 (δ 1.20) to C-3, C-4, C-5 and C-13 (δ 20.8) and from H-2 (δ 5.63) to C-3 showed the presence of a 3-oxo-decalin skeleton containing the partial structures II to IV. (3) Cross-peaks from H-2 and H₃-17 (δ 2.17) to C-16 (δ 170.3) showed that an acetoxy group is connected to C-2. The chemical shift of C-7 (δ 71.8) and the molecular formula showed the presence of a hydroxy group. (4) The finding that cross-peaks were

observed from H-11 to C-1', C-2' and C-6' and that the chemical shifts of C-8 (δ 79.0) and C-6' (δ 159.1) correspond to an oxygenated carbon indicated that a phenol and a decalin ring are connected by a pyran ring. The pentacyclic structure was found to consist of a sixmembered lactone, a phenol, a pyran and a decalin ring. Thus, the structure of pentacecilide C was elucidated as shown in Figure 1. The structure satisfied the degree of unsaturation and the molecular formula. Furthermore, all chemical shifts, except for C-2 in pentacecilide C, were comparable with those reported for thailandolide A.²

209

Table 2 ¹H and ¹³C NMR chemical shift of pentacecilides A, B and C

No.		Pentacecilide A		Pentacecilide B		Pentacecilide C	
	δ_{C}	δ _H (J in Hz)	$\delta_{\mathcal{C}}$	δ_H (J in Hz)	$\delta_{\mathcal{C}}$	δ _H (J in Hz)	
1	31.7	1.67 m 2.06 m	40.7	1.85 m 2.18 m	41.1	1.88 m 2.20 m	
2	33.7	2.44 m 2.68 m	72.6	5.59 (12.0, 7.0)	72.2	5.63 (12.0, 7.0)	
3	219.6	_	208.6	_	207.8	_	
4	47.2	_	48.6	_	48.3	_	
5	48.1	1.88 m	48.0	1.77 m	45.6	1.82 m	
6	18.5	1.58 m	17.7	1.68 m	26.7	1.86 m	
				1.75 m		2.18 m	
7	33.7	2.08 m	36.6	2.10 m	71.8	4.11 dd (10.0, 3.0)	
8	78 3	<u> </u>	78 1	_	79.0	_	
9	70.5 44.8	1.86 m	48.5	1 88 m	43.0	2 23 m	
10	35.4		36.6		36.2		
11	20.3	2 44 m	21.2	2 44 dd (10 0 2 5)	20.7	2 52 m	
12	25.5	1.35 s	24.0	1 26s	21.2	1.235	
13	20.0	1.09 s	21.3	1.18s	20.8	1.20s	
14	29.3	1.12 s	26.3	1.16s	25.7	1.14s	
15	23.0	0.96 s	24.9	1.39s	24.6	1.54 s	
16			170.3	_	170.3	_	
17			21.0	2.17s	21.0	2.17s	
1'	110.5	_	110.9		110.7	_	
2'	139.2	_	139.3	_	139.3	_	
3′	101.7	_	101.9	_	102.2	_	
4′	162.5	_	162.6	_	162.3	_	
5′	103.3	6.27 s	103.5	6.27 s	103.5	6.30 s	
6′	160.4	=	160.2	_	159.1	_	
7′	32.0	2.72 dd (17.0, 11.0) 2.84 dd (17.0, 3.5)	32.1	2.70 dd (17.0, 11.0) 2.85 dd (17.0, 3.5)	31.8	2.70 dd (17.0, 11.0) 2.85 dd (17.0, 3.5)	
8′	75.0	4.64 m	74.9	4.62 m	74.7	4.62 m	
9′	21.2	1.54 d (7.0)	21.2	1.55 d (7.0)	20.9	1.55 d (7.0)	
10′	170.5	_	170.3	_	169.9	_	
4'-0H		11.08s		11.07 s		11.08s	



Structure elucidation of pentacecilide B

The molecular formula $C_{27}H_{34}O_7$ of pentacecilide B is smaller by one oxygen atom than that of pentacecilide C. Comparison of the ¹H NMR spectra between pentacecilides B and C indicated that the oxygenated sp^3 methine proton (H-7) in pentacecilide C is replaced by methylene protons (δ 2.10) in pentacecilide B. In fact, analyses of the ¹H–¹H COSY revealed the presence of the partial structure V containing the replaced part (Figure 3b). The partial structure V was also confirmed by observing cross-peaks from H₃-12 (δ 1.26) to C-7 (δ 36.6), C-8 (δ 78.1) and C-9 (δ 48.5), from H₂-7 (δ 2.10) to C-8, from H-9 (δ 1.88) to C-8, C-10 (δ 36.6) and C-15 (δ 24.9), from H₂-6 (δ 1.68, 1.75) to C-10, from H₃-15 (δ 1.39) to C-9 and C-10 and from H-5 (δ 1.77) to C-10 in ¹³C–¹H HMBC experiments (Figure 3b). Taken together, the structure of pentacecilide B was elucidated as 7-dehydroxy pentacecilide C (Figure 1).

Structure elucidation of pentacecilide A

The molecular formula $C_{25}H_{32}O_5$ of pentacecilide A is smaller by $C_2H_2O_3$ than that of pentacecilide C. Comparison of the ¹H NMR spectra of pentacecilides A and B showed that the methyl protons (H₃-17) disappear and the oxygenated *sp*³ methine proton (H-2) is



Figure 3 ¹H–¹H COSY and ¹³C–¹H HMBC experiments of pentacecilides A (a), B (b) and C (c).

replaced by methylene protons (δ 2.44, 2.68). Analyses of the ¹H–¹H COSY revealed that the partial structure VI contains the replaced part (Figure 3c). The partial structure VI was also confirmed by observing cross-peaks from H₃-15 (δ 0.96) to C-1 (δ 31.7) and C-10 (δ 35.4), from H-5 (δ 1.88) to C-4 (δ 47.2) and C-10, from H₂-1 (δ 1.67, 2.06) to C-3 (δ 219.6), C-5 (δ 48.1), C-10 and C-15 (δ 23.0), from H₃-13 (δ 1.09) to C-3, C-4, C-5 and C-14 (δ 29.3), from H₃-14 (δ 1.12) to C-3, C-4, C-5 and C-13 (20.0) and from H₂-2 to C-3 in ¹³C–¹H HMBC experiments (Figure 3c). Taken together, the structure of pentacecilide A was elucidated as 2-deacetoxy pentacecilide B (Figure 1).

Stereochemistry of pentacecilides

Pentacecilide C has seven chiral carbons in its structure. The relative stereochemistry at C-2, C-5, C-7, C-8, C-9 and C-10 of the tricyclic skeleton (A–B–C) was investigated by NOESY experiments. As shown in Figure 4, cross-peaks were observed between H-2 (δ 5.63) and H₃-13 (δ 1.20)/H₃-15 (δ 1.54), between H₃-15 and H_{ax}-6 (δ 2.18)/H₃-13, between H-5 (δ 1.82) and H-7 (δ 4.11)/H-9 (δ 2.23) and H-7 and H-9, indicating that they are oriented in a 1,3-diaxial conformation. Accordingly, rings A and B are oriented in a chair-chair form. Secondly, cross-peaks were observed between H₃-12 (δ 1.23) and H-7/H₂-11 (δ 2.52) and between H₃-15 and H₂-11, suggesting that rings B and C are oriented in a chair-boat form. Thirdly, cross-peaks were observed between H_{ar}-7' (δ 2.85) and H-11/H-8' (δ 4.62) and



ax; axial *eq*; equatorial



The Journal of Antibiotics

between H_{ax} -7' and H_3 -9 (δ 1.55), indicating that the conformation of H_3 -9' is equatorial, which was also supported by a large coupling constant (J=11 Hz) between H_{ax} -7' and H-8'. Taken together, the relative stereochemistry of pentacecilide C was determined as shown in Figure 1. These results were consistent with those of thailandolide A except for the stereochemistry at C-8', although chemical shifts at C-8' of pentacecilide C and thailandolide A were almost the same.²

The relative stereochemistry of pentacecilides A and B was deduced to be the same as that of pentacecilide C by the similarity of NOESY experiments and the coupling constants in ¹H NMR.

DISCUSSION

Pentacecilides A to C, structurally related to thailandolides, were isolated from the culture broth of *P. cecidicola* FKI-3765-1, and were found to have a common pentacyclic core containing an aromatic ring and a δ -lactone ring. The core seemed to be a meroterpene, consisting of a sesquiterpene and a pentaketide. Thailandolides were reported to be produced by *Talaromyces thailandiasis*,² whereas pentacecilides were produced by a different genus *Penicillium*. Thailandolides A and B were not detected in the culture broth of *P. cecidicola* FKI-3765-1.

From the structure elucidation, the planar structures of pentacecilides were seen to be similar to those of thailandolides. The relative stereochemistry of the compounds is almost the same, but the C-8' stereochemistry is different; the C-8' methyl group of thailandolides is oriented in the axial conformation,² whereas that of pentacecilides is in the equatorial conformation.

METHODS

General experimental procedures

UV spectra were recorded on a spectrophotometer (8453 UV-Visible spectrophotometer, Agilent Technologies Inc., Santa Clara, CA, USA). IR spectra were recorded on a Fourier transform infrared spectrometer (FT-710, Horiba Ltd, Kyoto, Japan). Optical rotations were measured with a digital polarimeter (DIP-1000, JASCO Corporation, Tokyo, Japan). ESI-TOF-MS and HRESI-TOF-MS spectra were recorded on a mass spectrometer (JMS-T100LP, JEOL Ltd, Tokyo, Japan). Various NMR spectra were measured with a spectrometer (XL-400, Varian Inc., Palo Alto, CA, USA).

ACKNOWLEDGEMENTS

This study was supported by the Program for the Promotion of Fundamental Studies in Health Sciences (to HT) from the National Institute of Biomedical Innovation (NIBIO). We express our thanks to Ms N Sato for measuring NMR experiments, and Mr K Nagai and Ms A Nakagawa for measuring mass spectra. We also thank Mr N Ugaki for excellent technical assistance.

 Yamazaki, H et al. Pentacecilides, new inhibitors for lipid droplet formation in mouse macrophages produced by *Penicillium cecidicola* FKI-3765-1.
I. Taxonomy, fermentation, isolation and biological properties. J. Antibiot. 62, 195–200 (2009).

² Dethoup, T et al. Merodrimanes and other constituents from Talaromyces thailandiasis. J. Nat. Prod. 70, 1200–1202 (2007).