

ORIGINAL ARTICLE

An acid-tolerant ammonia-oxidizing γ -proteobacterium from soil

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Nitrification, the microbial oxidation of ammonia to nitrate via nitrite, occurs in a wide range of acidic soils. However, the ammonia-oxidizing bacteria (AOB) that have been isolated from soil to date are acid-sensitive. Here we report the isolation and characterization of an acid-adapted AOB from an acidic agricultural soil. The isolated AOB, strain TAO100, is classified within the *Gammaproteobacteria* based on phylogenetic characteristics. TAO100 can grow in the pH range of 5–7.5 and survive in highly acidic conditions until pH 2 by forming cell aggregates. Whereas all known gammaproteobacterial AOB (γ -AOB) species, which have been isolated from marine and saline aquatic environments, are halophiles, TAO100 is not phenotypically halophilic. Thus, TAO100 represents the first soil-originated and non-halophilic γ -AOB. The TAO100 genome is considerably smaller than those of other γ -AOB and lacks several genes associated with salt tolerance which are unnecessary for survival in soil. The ammonia monooxygenase subunit A gene of TAO100 and its transcript are higher in abundance than those of ammonia-oxidizing archaea and betaproteobacterial AOB in the strongly acidic soil. These results indicate that TAO100 plays an important role in the nitrification of acidic soils. Based on these results, we propose TAO100 as a novel species of a new genus, *Candidatus Nitrosoglobus terrae*.

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Introduction

Extensive applications of nitrogen fertilizers have enhanced nitrification activity, resulting in the pollution of surface and ground waters and emission of the greenhouse gas nitrous oxide (Schlesinger, 2009; Wrage *et al.*, 2001). Ammonia oxidation, the first and rate-limiting step of nitrification, is carried out in soil by ammonia oxidizing bacteria (AOB) (Kowalchuk and Stephen, 2001), ammonia-oxidizing archaea (AOA) (Leininger *et al.*, 2006) and complete ammonia oxidizing (comammox) bacteria (Daims *et al.*, 2015; van Kessel *et al.*, 2015). AOB, of which sequences have been found suggesting that they may be involved in ammonia oxidation in soil (Di *et al.*, 2009; Jia and Conrad, 2009), have been shown to be

acid-sensitive and do not grow below pH 6.0 in pure culture (Jiang and Bakken, 1999). However, high-nitrification rates have been found in a wide variety of acidic soils with pH below 5.5 (Booth *et al.*, 2005). Thus, the microbiological basis of ammonia oxidation in acidic soil remained unclear until the recent discovery of AOA (Konneke *et al.*, 2005; Leininger *et al.*, 2006; Tourna *et al.*, 2011). Indeed, AOA are more abundant than AOB in many acidic soils (Zhang *et al.*, 2012), and acidophilic AOA, *Nitrosotalea devanattera* and *Nitrosotalea* species, have been isolated from acidic soil and characterized (Lehtovirta-Morley *et al.*, 2011, 2014). These observations strongly support the hypothesis that AOA are more important than AOB in nitrification in acidic soil.

AOB have, however, been identified as the dominant ammonia oxidizers in several acidic soils (Long *et al.*, 2012; Petersen *et al.*, 2012; Wertz *et al.*, 2012). A few AOB have been isolated from acidic soils with a pH around 4 (Jiang and Bakken, 1999) although they have been found to be acid sensitive in

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pure culture. Ammonia oxidation by these acid-sensitive AOB in acidic soils might be explained by several hypothetical mechanisms (De Boer and Kowalchuk, 2001) including pH-neutral micro-sites, urea hydrolysis (Burton and Prosser, 2001; De Boer *et al.*, 1989), and acid resistant biofilm and aggregate formation (Allison and Prosser, 1993; De Boer *et al.*, 1991). Conversely, we previously reported the potential presence of acid-tolerant AOB in acidic soil from tea fields, where nitrification occurs at high levels despite the soil having a pH of about 3 (Hayatsu, 1993; Hayatsu and Kosuge, 1993). These studies suggested the existence of acid adapted AOB and their contribution to nitrification in acidic soil. Therefore, AOB cannot be excluded from the organisms that mediate nitrification in acidic soil, and the discovery of new acid-tolerant AOB is anticipated.

The aim of the current study was to isolate and characterize acid-adapted AOB from the acidic soil of tea fields and to demonstrate the contribution of the isolated AOB to ammonia oxidation activity in the tea field soil. In addition, we aimed to provide genomic level evidence for the taxonomic and evolutionary relationships between isolated acid-adapted AOB and known related AOB.

Materials and methods

Soil samples

Soil samples, classified as Andosol, were obtained in July 2012 from tea field plots at Kanaya Tea Research Station, Institute of Fruit Tree and Tea Science, NARO, in Japan. These plots are fertilized every year to study the effects of N, P, K and Ca on tea plants (Table 1). N, P, K and Ca were supplied as ammonium sulfate, superphosphate, potassium sulfate and dolomite ($\text{CaCO}_3 \cdot \text{MgCO}_3$), respectively. Soil samples were collected as cores from 10-cm deep and immediately stored at -80°C until molecular analyses or at 4°C until potential nitrification rate analyses.

Potential ammonia oxidation rate (PAR)

Potential rates of ammonia oxidation were measured as rates of nitrite accumulation in the presence of chlorate, which inhibits nitrite oxidation (Belser and Mays, 1980).

Slurries were obtained by mixing 2.5 g soil sample with 10 ml of 1 mM potassium phosphate buffer (pH 7) containing 10 mM $(\text{NH}_4)_2\text{SO}_4$ and 10 mM KClO_3 . Subsequently, the slurries were incubated for several hours at 25°C with shaking at 150 r.p.m. Aliquots of 1 ml were drawn at 0 and 4 h and centrifuged at $10000g$ for 10 min at 4°C . The nitrite concentration in the supernatant was determined calorimetrically via diazotization (Keeney and Nelson, 1982). PAR were calculated as nmol nitrite per hour per g of dried soil. The obtained potential rates, which were measured at pH 7, may not reflect actual nitrification rates in acidic soil at *in situ* pH. However, potential nitrification rate assessment at this pH value has been utilized in many studies, including those on nitrification in acidic soil (for example, Peterson *et al.*, 2012; Wertz *et al.*, 2012). We used the potential rate to facilitate comparisons with other studies.

Enrichment of ammonia oxidizing bacteria

Soil samples with pH 2.9 and 3.1 were collected from no Ca and high N treatment plots, respectively (Table 1) and used to obtain enrichment cultures. The initial cultures were obtained by suspending 10 g soil in 100 ml Schmidt AOB media (Schmidt and Belser, 1994) containing 100 mM ammonium sulfate at pH 5.5. These cultures were grown at 25°C with shaking at 110 r.p.m. until high-rates of ammonia oxidation were observed. The cultures were then diluted 10-fold and grown further. Multiple cycles of dilution-to-extinction were performed until no culturable heterotrophic microorganisms were detected. Contaminating microorganisms were assessed by 10-fold diluted nutrient agar. We designed 16S rRNA-specific fluorescent probes for fluorescence *in situ* hybridization (FISH) analysis and a polymerase chain reaction (PCR) primer set (Supplementary Table S1) for enriched ammonia oxidizing bacteria using retrieved 16S rRNA gene sequences of DNA fragments amplified from the enriched culture using the Bacteria-specific primers 27F and 1492r (Lane, 1991). The final culture purity was evaluated by FISH (Supplementary Methods) and quantitative PCR (qPCR) for the 16S rRNA gene using the designed FISH probe and PCR primer set,

Table 1 Characteristics of soils used in the study

Treatment ^a	Soil pH	Total C (%)	Total N (%)	PAR ^b ($\text{nmole}^{-1}\text{h}^{-1}\text{g}^{-1}$)	Fertilization ($\text{kg ha}^{-1}\text{y}^{-1}$) ^c	
					N input	Ca (dolomite)
Standard	3.36 (0.05) c	16.86 (1.32) b	1.23 (0.13) b	45.91 (5.74) a	400	1 000
High Ca	4.74 (0.10) d	11.93 (0.13) a	0.84 (0.04) a	253.66 (5.10) c	400	3 000
No Ca	2.89 (0.05) a	19.47 (0.59) c	1.54 (0.06) d	55.84 (4.80) a	400	0
High N	3.09 (0.04) b	17.82 (0.37) b	1.52 (0.06) c	180.18 (17.90) b	1200	1 000

s.d. are given in parentheses. Values within the same column followed by the same letter are not significantly different at $P < 0.05$.

^aTreatments: standard fertilization (Standard), high-rate Ca fertilizer input (high Ca), no Ca fertilizer input (no Ca) and high-rate N fertilizer input (high N).

^bPAR, potential ammonia oxidation rate ($\text{NO}_2^{-1}\text{nmole}^{-1}\text{h}^{-1}\text{g}^{-1}$ dry soil).

^cDolomite and ammonium sulfate were added to the fields as sources of Ca and N, respectively.

respectively. Total bacterial 16S rRNA gene copy number was quantified by qPCR using a Bacteria-specific primer set. AOA ammonia monooxygenase sub-unit A (*amoA*) and betaproteobacterial AOB (β AOB)-*amoA* were checked by PCR using their specific primers. The primers and probes used in this study are listed in Supplementary Table S1.

Quantification of *amoA* genes

Total DNA was extracted from the enriched cultures of ammonia oxidizing bacteria designated as TAO100 by using the Fast DNA spin kit for bacteria (Qbiogene, Inc., Irvine, CA, USA), and the number of copies of the TAO100 *amoA* gene was quantified by real-time PCR using the SYBR Premix Ex Taq kit (TaKaRa Bio, Inc., Shiga, Japan). We designed a PCR primer set, TAO-amoAF and TAO-amoAR (Supplementary Table S1). TAO100 *amoA* was amplified using the following cycling conditions: 94 °C for 2 min; 40 cycles of 30 s at 94 °C, 30 s at 58 °C, 45 s at 72 °C; and assessment at 72 °C. A standard curve was constructed using the TAO100 *amoA* DNA fragment amplified by PCR that had been cloned into pGEM. The detection limit was $\sim 2.5 \times 10^5$ copies ml⁻¹ of culture.

Phylogenetic analysis

Genomic DNA was extracted from TAO100 cells using a Fast DNA spin kit for bacteria. Bacterial 16S rRNA genes were PCR-amplified using the bacterial 27F and 1492r primer set. The TAO100 *amoA* gene was amplified using TAO-amoAF and TAO-amoAR. The PCR products were sequenced using ABI Prism BigDye Terminator cycle sequencing ready reaction kits and an ABI Prism 3730xl DNA analyzer (Applied Biosystems, Foster City, CA, USA). The phylogenetic analyses were performed using the MEGA 6 software program (Tamura *et al.*, 2013). Average nucleotide identity (ANI) values for 3 whole-genome sequences of strains of the genera *Nitrosococcus* including *N. oceani* ATCC 19707, *N. halophilus* NC4 and *N. watosoni* C-113, were determined using a web-based service (<http://enve-omics.ce.gatech.edu/ani/>).

Growth experiments

Ten milliliters of cultures enriched in TAO100 were inoculated into 100 ml Schmidt AOB media modified as described below. Cultures were incubated at 25 °C with rotary shaking at 110 r.p.m. and 1 ml samples were drawn periodically. Subsequently, nitrite and/or ammonium in the supernatant were determined colorimetrically using diazotization and indophenol blue (Keeney and Nelson, 1982), respectively. To assess the effect of pH on growth, media were buffered with 80 mM 2-morpholinoethanesulfonic acid (MES) to pH 5.0, 5.5, 6.0, 6.5 and 7.0 or with 80 mM 4-(2-hydroxyethyl)-1-piperazine-ethanesulphonic acid (HEPES) to pH 7.0, 7.5 and 8.0. To assess the effects of NaCl, media were buffered with 80 mM MES to pH 6.0 and supplemented

with 50, 100, 200, 300, 400 and 500 mM NaCl. The effects of NH₄⁺ were assessed in the media with 80 mM MES to pH 6.0 and supplemented with 50, 100, 200, 300, 400 and 500 mM NH₄⁺.

Kinetics of ammonia oxidation

The half-saturation constant (K_m) values of ammonia oxidation were determined by estimating the O₂ consumption at 25 °C of a suspension of TAO100 cells in a stirred vessel with a Clark-type oxygen electrode. The vessel contained TAO100 cells in 1 ml buffer containing 50 mM MES (pH 6.0) and various concentrations of ammonium sulfate. K_m values were calculated using a double-reciprocal plot using Sigma Plot 12.5 (Systat Software, San Jose, CA, USA).

Analysis of conserved chromosome structure and genome-wide phylogenetic analysis

Whole-genome sequencing and annotation of TAO100 were performed as described in Supplementary Methods. The results were used for further analysis. Orthologous groups among the TAO100 and three γ -AOB were constructed using DomClust (Uchiyama, 2006) on the Microbial Genome Database (MBGD) server (Uchiyama *et al.*, 2013). An alignment of syntenically conserved regions, designated the 'core genome structure,' was subsequently constructed using CoreAligner (Uchiyama, 2008) based on a consensus chromosomal arrangement conserved among these orthologs. To infer the phylogenetic relationships among TAO100, the three γ -AOB, and nine related strains belonging to order Chromatiales (the specific strains are listed in Supplementary Methods), we first constructed the core genome structure among these genomes using CoreAligner. Among the resulting core genes, 245 were found to be conserved in all 13 strains with one-to-one correspondence. CLUSTALW (Larkin *et al.*, 2007) was used to generate a multiple sequence alignment of these genes, from which blocks suitable for phylogenetic analyses were extracted using Gblocks program (Talavera and Castresana, 2007). The resulting sequence blocks were concatenated to construct an approximately maximum likelihood phylogenetic tree using FastTree (Price *et al.*, 2010).

Evaluation of metabolic and physiological potential

Metabolic and physiological potential harbored in the TAO100 genome were investigated by the Metabolic And Physiological potential Evaluator (MAPLE) (<http://www.genome.jp/tools/maple/>) to calculate the completion ratio of the functional modules defined by KEGG (Takami *et al.*, 2012, 2016). The module completion ratio (MCR) was calculated based on a Boolean algebra-like equation according to a previously described procedure (Takami *et al.*, 2012). The MCR pattern of the TAO100 was compared with those of other γ -AOB to characterize differences in the physiological potential.

Quantification of *amoA* genes and their transcripts in soil samples

DNA was extracted from 0.4 g of triplicate soil samples using Fast DNA Spin Kits for soil, as described in previous studies (Morimoto *et al.*, 2011; Takada-Hoshino and Matsumoto, 2004). The extracted DNA was then purified using a Sepharose gel filtration kit (Roche Applied Science, Indianapolis, IN, USA) and an UltraClean 15 DNA Purification Kit (MOBIO Labs, Solana Beach, CA, USA). The abundances of *amoA* of AOA, β -AOB, and TAO100 were quantified by real-time PCR using the SYBR Premix Ex Taq kit with the primer sets of *amoA*19F (Leininger *et al.*, 2006) and *CrenamoA*616r (Tourna *et al.*, 2008), *amoA*1F (Rotthauwe *et al.*, 1977) and *amoA*2R (Nicolaisen and Ramsing, 2002), and TAO-*amoA*F and TAO-*amoA*R (details in Supplementary Methods).

For transcript analysis, RNA was extracted from the soil as described in a previous report (Wang *et al.*, 2012). A one-step reverse transcription (RT)-qPCR was performed using a QuantiTect SYBR Green RT-PCR Kit (Qiagen, Inc., Valencia, CA, USA) and the StepOnePlus Real-Time PCR System (Applied Biosystems) (details in Supplementary Methods).

Availability of sequences

Sequences obtained in this study were deposited in the DNA Data Bank of Japan (DDBJ) under the accession numbers AP014836, AP014837 and DRA003611.

Results and discussion

Enrichment of acid-tolerant AOB

We used soil samples with pH values of 2.9 and 3.1 (Table 1), which were obtained from experimental plots of tea fields treated with heavy nitrogen fertilization, to isolate acid-tolerant AOB. The *amoA*

gene abundance of AOA and β -AOB was below the detection limit of qPCR although the soil exhibited high nitrification activities. We therefore speculated that unknown acid-tolerant or acidophilic ammonia oxidizers might be involved in nitrification of these tea field soils.

The culture media used for isolation was adjusted to pH 5.5 and supplemented with ammonium at concentrations higher than those used to isolate AOA or AOB in previous studies (Jiang and Bakken, 1999; Tourna *et al.*, 2011; Bollmann *et al.*, 2011). Higher levels of ammonium were used because acidification reduces the concentration of NH_3 , which is believed to be the preferred substrate (instead of NH_4^+) for ammonia monooxygenase (Suzuki *et al.*, 1974). Cultures highly enriched in ammonia-oxidizing microorganisms were successfully obtained from the soil samples via serial dilution-to-extinction techniques. The final enrichment culture contained single morphotype large coccus cells. AOA *amoA* and β -AOB *amoA* genes were not detected in the culture by PCR with specific primers for these genes. Clones amplified with the bacterial 16S rRNA gene primers 27F and 1492r were of a single sequence type. The 16S rRNA gene sequence showed the highest similarity to those of *Nitrosococcus* genus. We evaluated the culture purity by FISH and qPCR analysis using fluorescent probes and a PCR primer set specific to the 16S rRNA gene of the enriched AOB designed strain TAO100. FISH analysis showed that the enriched cells comprised >99% of all 4,6-diamidino-2-phenylindole (DAPI) stained cells (Supplementary Figure S1). The qPCR analysis also showed that the 16S rRNA gene copy number of the enriched AOB was equal to that obtained for the total bacteria. Ammonia consumption, nitrite production and the *amoA* gene abundance were determined to confirm

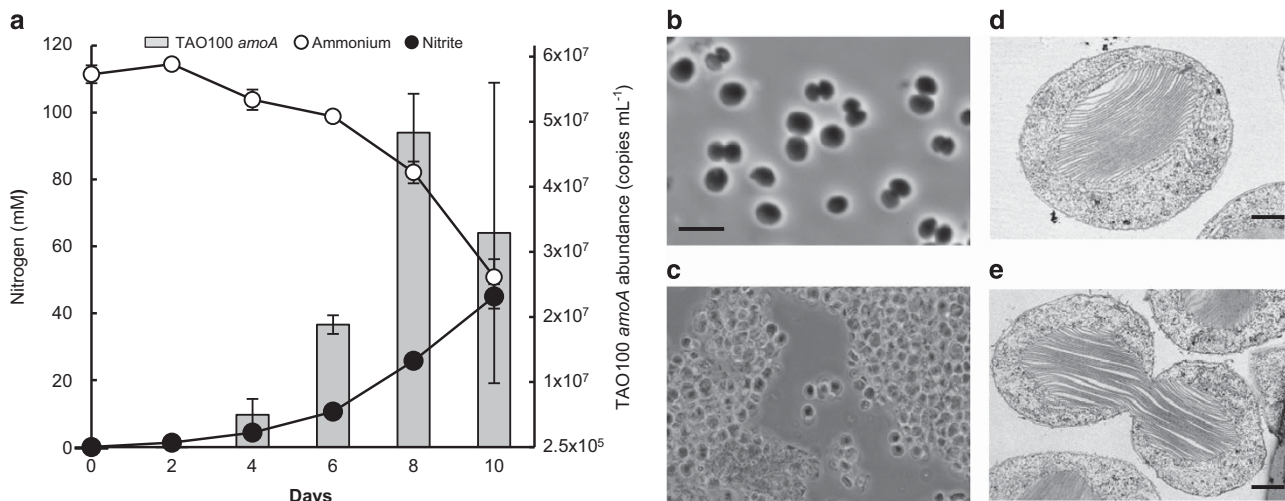


Figure 1 Growth and morphology of TAO100. (a) Correlation of ammonia oxidation with growth of TAO100. TAO100 was grown in media at pH 6 and containing 100 mM ammonium. Error bars indicate s.d. and appear smaller than symbols in some cases. (b) Phase contrast photomicrograph. Scale bar, 5 μm . (c) Aggregates of TAO100 cells. Scale bar, 10 μm . (d, e) Thin-section electron micrographs of a cell (d) and paired cells (e). Scale bar, 0.5 μm . The method of electron microscopy was as described in Supplementary information.

the autotrophic ammonia oxidation and growth of TAO100. Growth of TAO100 was accompanied by a near-stoichiometric conversion of ammonia to nitrite (Figure 1a). To confirm the expression of essential genes required for autotrophic bacterial ammonia oxidation, total mRNA was extracted and analyzed by pyrosequencing (Supplementary Methods). Supplementary Table S2 lists the 23 transcripts with the highest copy numbers in the enrichment culture of TAO100. Genes encoding ammonia monooxygenase, hydroxylamine dehydrogenase and ribulose 1, 5-bisphosphate carboxylase oxygenase were abundantly expressed (Supplementary Table S2).

Phylogenetic analysis and morphological and physiological characteristics

Phylogenetic analysis based on the 16S rRNA gene and *AmoA* amino acid sequences (Supplementary Figure S2) revealed that TAO100 clusters with γ -AOB of the genus *Nitrosococcus*, which includes three known species, *N. oceani*, *N. halophilus* and *N. watonii*. However, the 16S rRNA and *amoA* gene sequences of TAO100 exhibited 94 and 70% (79% by amino acid sequence) similarity to those of the most closely related species of *Nitrosococcus halophilus*, respectively. The ANI value between TAO100 and these three species was 75%, which is far below the threshold for species demarcation (95–96%). These low percentages of 16S rRNA gene and *amoA* gene sequence similarities combined with the ANI value strongly suggest that TAO100 is phylogenetically distant from the species belonging to genus *Nitrosococcus*. AOB generally falls within the classes *Beta-proteobacteria* or *Gammaproteobacteria*. Established β -AOB genera, such as *Nitrosomonas* and *Nitrospira*,

have been found in various environments such as soils, oceans, lakes and activated sludge (Prosser and Embley, 2002). The *Nitrospira* genus has been shown to be the dominant functional AOB in agricultural and forest soils (Kowalchuk and Stephen, 2001). In contrast, γ -AOB of the *Nitrosococcus* genus thrive only in saline or hyper-saline aquatic environments such as oceans and salt lakes (Campbell *et al.*, 2011). Thus, TAO100 is the first classified *Gammaproteobacteria* AOB to be isolated from soil.

TAO100 cells in liquid culture were found to be non-motile, spherical or ellipsoidal in shape, and occurred singly or in pairs with diameters ranging from 2 to 3 μm (Figure 1b), and aggregated cells were occasionally observed (Figure 1c). Each cell contained a centrally located extensive intracellular membrane system composed of a stack of flattened membrane vesicles (Figures 1d and e). These cell features are extremely similar to the morphological characteristics of the *Nitrosococcus* genus. TAO100 grew optimally at 25 °C and also grew at 15 °C, but no growth was observed at 5 or 40 °C.

Previously isolated γ -AOB of the *Nitrosococcus* genus were reported to be obligatory halophiles and to require 500–700 mM NaCl for optimum growth (Campbell *et al.*, 2011; Wang *et al.*, 2016). Although TAO100 grew better at 50 mM NaCl than at 0 mM, TAO100 did not grow in NaCl concentrations greater than 400 mM (Figure 2), indicating that the strain is not halophilic.

Growth of TAO100 was observed in the pH range of 5–7.5 in medium containing 100 mM ammonium. The optimal pH for growth was 6–6.5 and the specific growth rate was 0.89 day^{-1} at pH 6 (Figures 3a and b), which is equivalent to the generation time of 27.0 h. The pH of a vigorously

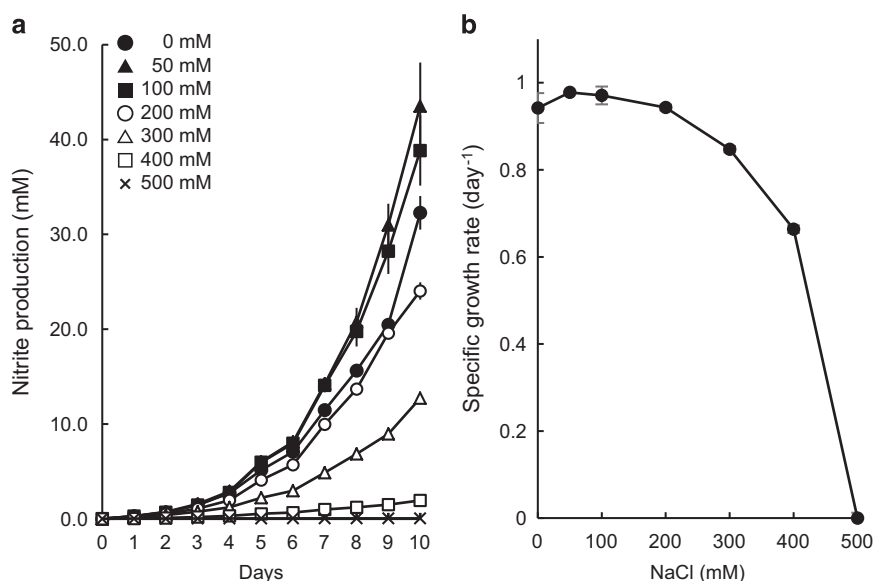


Figure 2 Effect of NaCl on nitrite production. (a) Nitrite production of TAO100 in media at pH 6 supplemented with 0–500 mM NaCl. Growth was estimated from nitrite production. (b) Specific growth rate from log-linear plots of nitrite during exponential growth. Error bars indicate s.d. from triplicate experiments and appear smaller than the symbols in some cases.

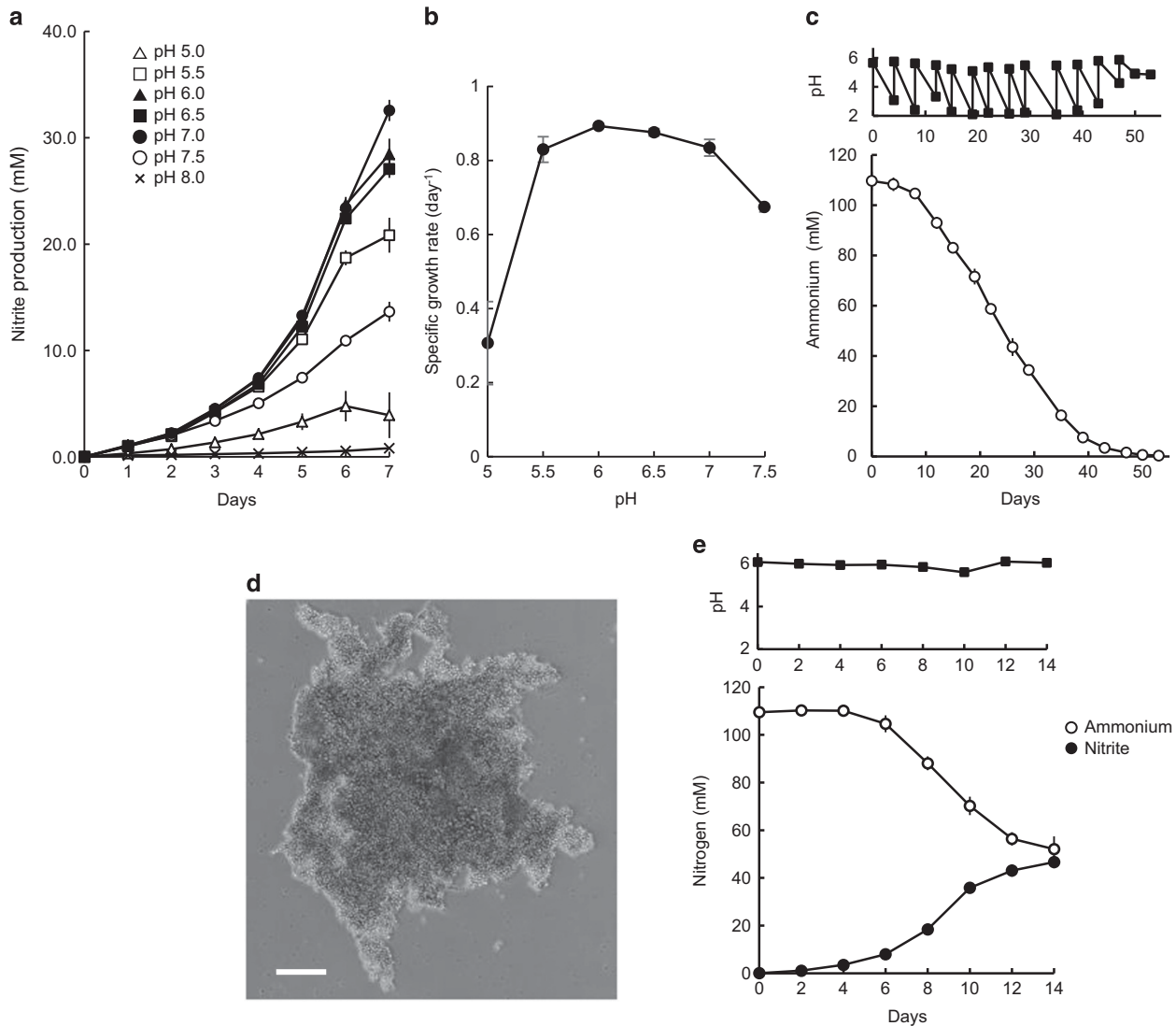


Figure 3 Effect of pH on ammonia oxidation of TAO100. (a) Ammonia oxidation of TAO100 in media containing 100 mM ammonium at pH 5–8. (b) Specific growth rate from log-linear plots of nitrite during exponential growth. (c) Ammonia oxidation of TAO100 in medium without MES buffer in which pH was periodically adjusted to ~6. Growth was estimated by measuring ammonium oxidation to nitrite. Nitrite levels were not determined because nitric acid produced from ammonium was lost via evaporation under acidic conditions. (d) Visible cell aggregates formed in the acidified culture. Scale bar, 100 μ m. (e) Growth of TAO100 in a medium with MES buffer in which pH was maintained continuously at ~6 after inoculation of the cell aggregates. Data were obtained from triplicate experiments. Error bars are s.d. and appear smaller than the symbols in some cases.

growing culture decreased from 6 to ~2 when pH was not controlled (Figure 3c). After the acidified pH of the culture was adjusted to ~6, the cells continued to utilize ammonia. When this process was repeated several times, TAO100 formed visible, large cell aggregates (Figure 3d). When the culture containing these aggregates was inoculated into fresh medium at pH 6 (Figure 3e), TAO100 proliferated as a single cell, indicating that the aggregates themselves are not resistant, but the formation of aggregates might represent a mechanism enabling cells to cope with low pH. Similarly, aggregates formed in acidic conditions by *Nitrosospira*-like bacteria and *Nitrosospira* sp. AHB1 isolated from Dutch acid soils were able to nitrify at approximately pH 4 (De Boer et al., 1991). The lowest pH limit for the growth of TAO100

aggregates indicated that TAO100 is tolerant to more highly acidic conditions than β -AOB.

At pH 6, TAO100 grew optimally in ammonium concentrations between 100 and 200 mM (Figure 4a) but could tolerate concentrations up to 500 mM. Thus, TAO100 appears to require an unusually high concentration of ammonium, presumably because NH_3 rather than NH_4^+ is the actual substrate for its ammonia monooxygenase, the concentration of which decreases with acidification. The K_m for ammonia ($\text{NH}_3 + \text{NH}_4^+$) oxidation was 58.5 mM at pH 6, at which the NH_3 concentration was theoretically estimated to be 33.3 μ M (Figure 4b). The estimated K_m for NH_3 was similar to that of *Nitrosomonas europaea* but higher than those of several *Nitrosospira* strains (Koper et al., 2010). Overall, TAO100 appeared to have adapted to the soil,

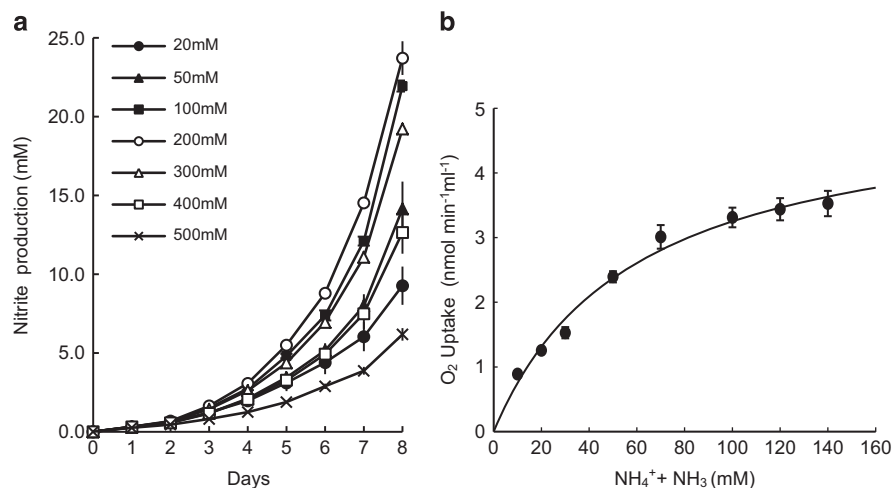


Figure 4 Effect of ammonium concentration on ammonium oxidation to nitrite by TAO100. (a) Nitrite production by TAO100 in media at pH 6 supplemented in various concentrations of ammonium. Data are from triplicate experiments. Error bars indicate s.d. and are smaller than the symbols in some cases. (b) Michaelis–Menten plot of ammonium-dependent oxygen uptake. The Michaelis–Menten equation was used to calculate kinetic parameters. K_m for ammonia ($\text{NH}_3 + \text{NH}_4^+$) = 58.5 mM at pH 6, at which the NH_3 concentration was theoretically estimated as 33.3 μM .

which is highly acidic and contains high amounts of ammonium.

Genome characteristics

The TAO100 genome consists of a single circular chromosome (2011873 bp) and a plasmid (6441 bp) with mean G+C contents of 42.1% and 40.9%, respectively (Supplementary Figure S3). This chromosome is considerably smaller than the typical *Nitrosococcus* genus γ -AOB chromosome, which ranges from 3.33 to 4.08 Mb (Klotz and Stein, 2011). The G+C content of the TAO100 chromosome was lower than those of *Nitrosococcus*, which range from 50.1 to 51.6%. Of the 1839 protein-coding sequences (CDSs) identified in the chromosome, 946 genes (55%) were most similar to those of *Nitrosococcus halophilus*. In addition, 260 (15%) and 189 genes (11%) were most similar to CDSs in *Nitrosococcus oceani* and *Nitrosococcus watsonii*, respectively. We performed ortholog analyses among TAO100 and three other γ -AOB using the MGD (Uchiyama et al., 2013) and identified 1,381 orthologous genes that were shared among the four strains (Supplementary Figure S4). The proportion of these shared orthologous genes in the TAO100 genome is considerably higher than that in other γ -AOB. Furthermore, we investigated the synteny ('core genome structure') conserved in at least two of the four species (Figure 5a; Supplementary Figure S5 and Table S3). The results indicated that TAO100 appears to have lost a number of genes from a common ancestor of γ -AOB through the evolutionary process. Finally, based on a genome-wide phylogenetic analysis using the alignment of concatenated sequences of 245 orthologs shared among TAO100 and 12 other related strains, we found that TAO100 is distinct from the *Nitrosococcus* genus despite the fact that TAO100 belongs to family *Chromatiaceae* (Figure 5b).

An analysis of the metabolic and physiological potential in the TAO100 genome using MAPLE indicated that the TAO100 genome possesses the elements necessary for establishment of complete pathways responsible for a chemolithoautotrophic lifestyle such as ammonia oxidation and the Calvin cycle (Figure 6a). The organization of the *amo* operon in TAO100 is identical to that of *N. halophilus*. On the other hand the gene for nitrite reductase (*nirK*) is missing from the TAO100 genome. The enzyme required for denitrification is found in all known AOB except *Nitrosomonas communis* Nm2 (Kozłowski et al., 2016). Kozłowski et al. (2014) proposed that the *nirK* of AOB might be primarily involved in an ammonia oxidation process rather than nitrite reduction during denitrification. Thus, a comparison of TAO100 with AOB strains having the *nirK* gene and with *N. communis* Nm2 might provide useful information to investigate the role of *nirK* in the ammonia oxidation process. Additionally, although all known AOB utilize polar flagella for motility, flagella-related genes were not identified in the TAO100 genome. Furthermore, ammonia transporters of the Rhesus clade are encoded in many AOB genomes (Offre et al., 2014); however, an ammonia transporter gene was not identified in the TAO100 genome. Similarly, *N. oceani* and *Nitrosomonas eutropha* genomes also do not encode known ammonia transporters (Klotz et al., 2006; Stein et al., 2007). TAO100 and these AOB might therefore primarily depend on a passive ammonia diffusion process across the membrane.

In contrast to the halophilic γ -AOB genome, the TAO100 genome does not encode a gene set for ectoine synthesis or for osmoprotectant transport (Figure 6a), both of which confer salt tolerance (Ventosa et al., 1998). In addition, fewer genes were found to be associated with sodium transport, which

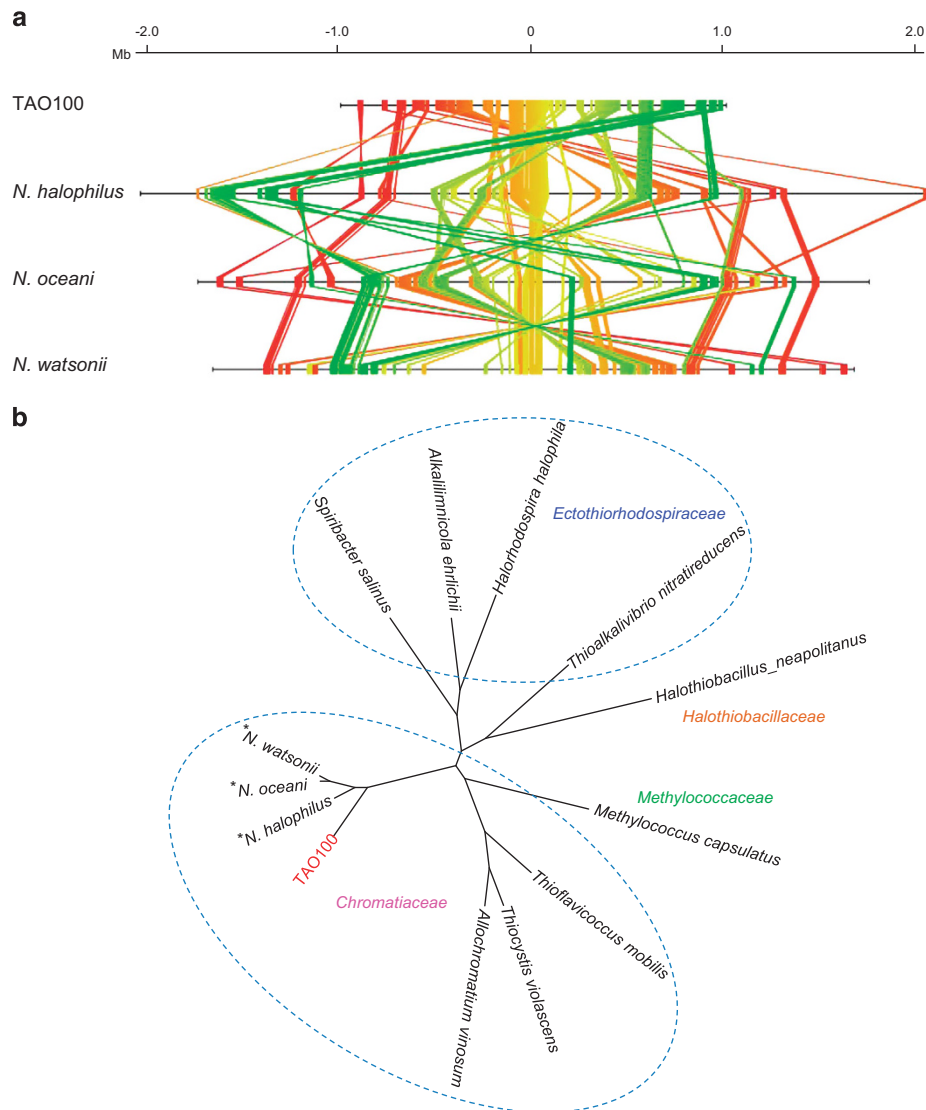


Figure 5 Comparison of orthologous genes in TAO100 and related strains. **(a)** Location of the genes in the core genome structure conserved among TAO100 and three halophilic γ -AOB on each chromosome. The conserved chromosomal structure ('core genome structure') was constructed based on consensus arrangement of conserved orthologs. In the core genome structure, an ortholog group is denoted as a colored line. For simplicity, only universally conserved ortholog groups with one-to-one correspondence are presented. To visualize chromosomal rearrangement, a color gradation scale from red to yellow to green is assigned according to location on the TAO100 chromosome. Replication origins are located at the center. **(b)** Phylogenetic relationships of orthologs common to TAO100, three γ -AOB, and nine related strains from class *Gammaproteobacteria*. The tree was generated using the concatenated amino acid sequence of 245 orthologous genes conserved in all genomes. Asterisk indicates three halophilic γ -AOB belonging to the genus *Nitrosococcus*.

would also likely confer halophilicity in the TAO100 genome. These genomic features are consistent with a nonhalophilic TAO100 classification, as indicated by the cultivation experiments. TAO100 therefore appears to have lost a number of genes critical for survival in oceans or salt lakes after diverging away from a common ancestor of halophilic γ -AOB. In contrast, the gene cluster encoding urea hydrolase is present in the TAO100 genome. This finding is consistent with the importance of ureolysis for survival in acidic soil (Burton and Prosser, 2001; De Boer *et al.*, 1989). Cation transporter and glycosyltransferase genes, both potentially associated with acid tolerance (Krulwich *et al.*, 2011; Stack *et al.*, 2010), were also identified in the

TAO100 genome (Figure 6b). Furthermore, glycosyltransferase has been implicated in exopolysaccharide synthesis and the formation of cell aggregates and biofilms, which confer stress tolerance including acid tolerance (Krulwich *et al.*, 2011). However, because these genes have also been identified in the acid-sensitive, neutrophilic γ -AOB genome, further study is required to clarify the molecular mechanism of acid tolerance in TAO100.

Contribution of TAO100 to nitrification in tea field soil
To evaluate the contribution of TAO100 to nitrification in the used soil samples (Table 1), the abundance of *amoA* genes and their transcript levels were measured

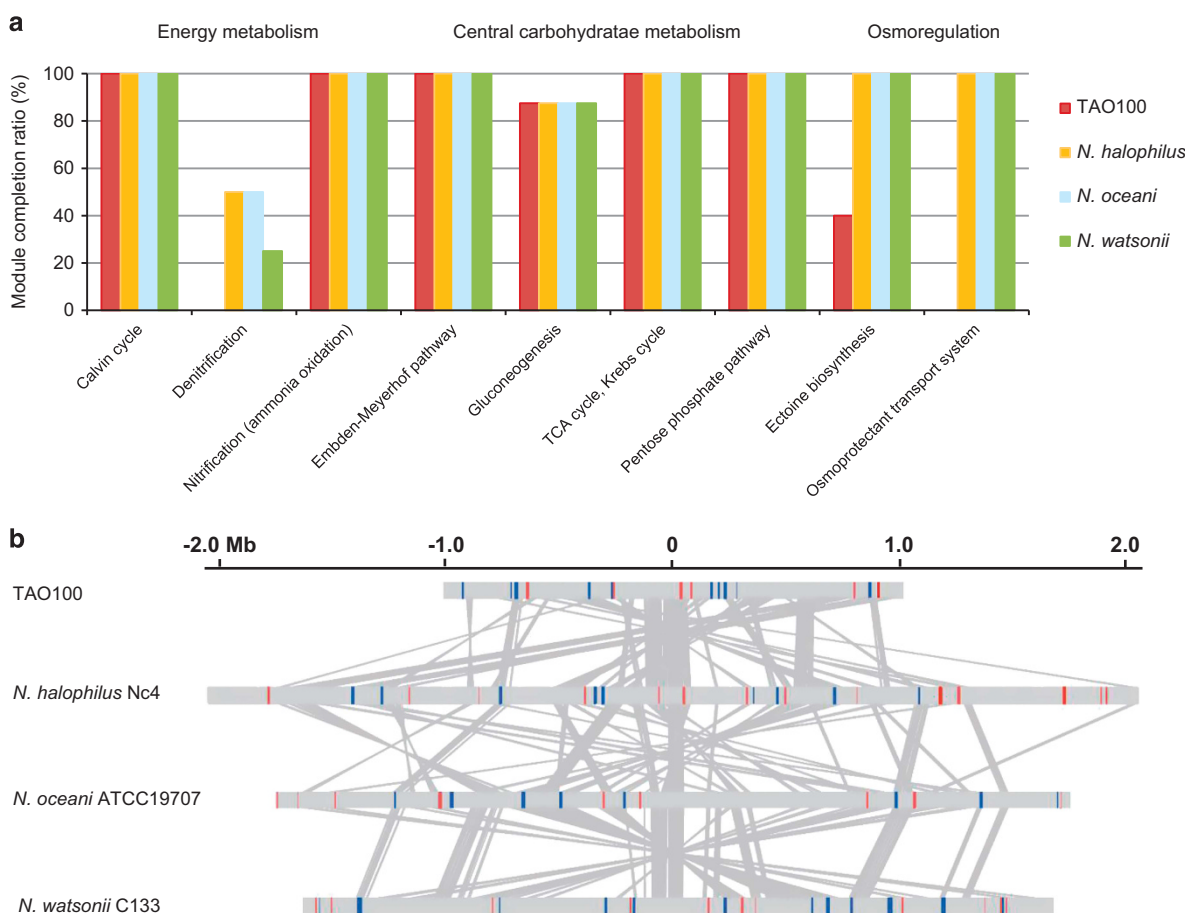


Figure 6 Metabolic and physiological potential encoded in the TAO100 genome. **(a)** Comparison of MCR patterns in major pathway modules between TAO100 and other γ -AOB. Completion ratio patterns of the modules categorized into energy metabolism, central carbohydrate metabolism and osmoregulation were compared between TAO100 and other major γ -AOB. MCR of each module was calculated by using the MAPLE system. **(b)** Distribution of glycosyltransferase and cation transporter genes in TAO100 and three γ -AOB. Glycosyltransferase and cation transporter genes are denoted in blue and red, respectively. Orthologous genes found in all four strains are connected with gray lines.

in the soils by qPCR using primers specific for TAO100, β -AOB and AOA (Figure 7). In the pH 2.9 and 3.1 soil samples, the TAO100 *amoA* gene and its transcript were more abundant compared with those of AOA and β -AOB. In contrast, AOA and β -AOB *amoA* genes and their transcripts were more abundant in pH 4.7 soil. The results indicated that TAO100 may be the primary group of ammonia oxidizers in very acidic soil. High-nitrogen application and the strongly acidic conditions of tea field soils might therefore have resulted in preferential growth of TAO100. On the other hand, the soil pH values of high N and no Ca were lower than that of the limit of growth of TAO100 in pure culture. Further experimentation such as monitoring the growth of TAO100 and measuring the actual nitrification rates using the acidic soil microcosms would be required to elucidate the role and validate contribution of TAO100 to nitrification in acidic soils.

We next screened 2212 whole genome shotgun sequencing data sets from soil, sediments and wastewater treatment plants available on the Metagenomics Rapid Annotation using Subsystem

Technology (MG-RAST) server (Meyer *et al.*, 2008) (Supplementary Methods). However no sequence read was affiliated with TAO100-*amoA* indicating that TAO100-*amoA* related γ -AOB *amoA* appeared to be absent or extremely rare in the soils used for the metagenomic analysis. Ultradeep sequencing is required to detect rare species in soil samples necessitating an estimated 50 Tbp for an individual gram of soil (Gans *et al.*, 2005). As few metagenome data sets of acidic and high-ammonium content soils such as that of the studied tea field are available, further metagenomic surveys of acidic soil are required to evaluate the distribution of TAO-related γ -AOB in soil and other environments.

In summary, evidence from genomic, phenotypic and also ecological analyses indicate that strain TAO100 of γ -AOB is uniquely adapted to acidic conditions and could contribute significantly to nitrification in acidic soil. To date, no strains of γ -AOB except TAO100 have been isolated and characterized from terrestrial ecosystems. This study suggests that γ -AOB are more phylogenetically and physiologically diverse than previously observed.

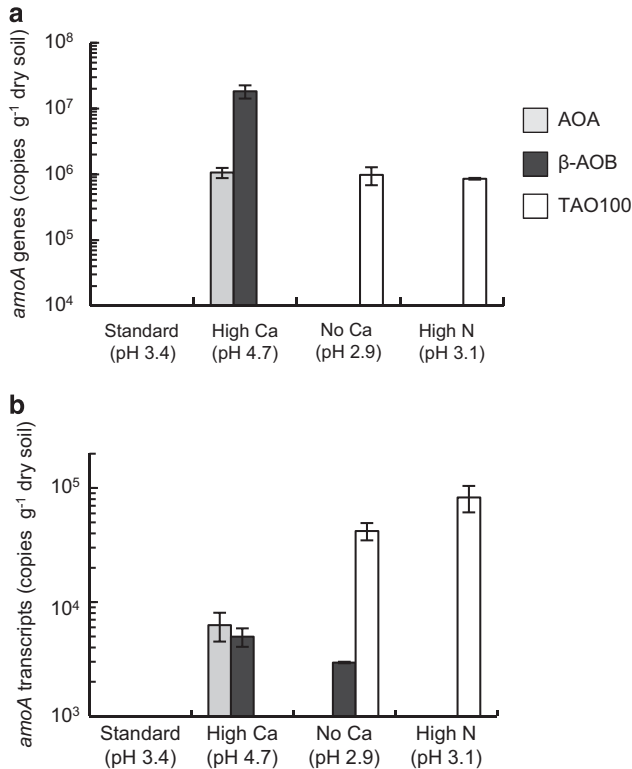


Figure 7 Abundances of *amoA* genes and *amoA* transcripts in the used soil samples. (a) *amoA* gene abundances of AOA, β -AOB and TAO100 in the soil samples. (b) *amoA* transcript abundances of AOA, β -AOB and TAO100 in the soil samples. Abundance is expressed as gene and transcript copy numbers per g of dry soil. No column indicates that the copy numbers of *amoA* gene and its transcript were below the detection limit of the qPCR assays. Error bars represent standard errors for triplicate samples. Soil pH is indicated in parentheses. Standard, High N, High Ca and No Ca indicate the fertilization conditions of the sampling fields (Table 1).

In support of this, recently, 16S rRNA gene sequences of *Nitrosococcus* genera were detected in moderately acidic pristine soil (Wang *et al.*, 2015), indicating the existence and distribution of γ -AOB in soil ecosystems. As such, the isolation and characterization of γ -AOB from soil can provide useful information to further our understanding of nitrification in acidic soil.

The differentiation of TAO100 from its closely related genus *Nitrosococcus* was confirmed by phylogenetic analysis based on 16S rRNA gene and *amoA* gene sequences, genome-wide analyses, and physiological features. These results support the classification of TAO100 into a separate genus and species from *Nitrosococcus*. We propose the new name *Nitrosoglobus terrae* gen. nov., sp. nov., and the following candidate status.

Description of Candidatus *Nitrosoglobus terrae*

The candidate status of strain TAO100 comprises *Nitrosoglobus terrae*, (Ni.tro.so.glo'bus. N.L. adj.), with the following etymology: nitrosus, full of

natron; here intended to mean nitrous; L. masc. n. globus, a ball; N.L. masc. n. Nitrosoglobus, the globus producing nitrite: ter'rae. L. gen. n. terrae, soil, where the type strain was isolated.

Cells are large cocci or very short rods with 2.0–3.0 μ m diameters. Cells contained a centrally located and well-developed intracytoplasmic membrane system composed of a stack of flattened membrane vesicles. This species is an obligate chemolithotroph and oxidizes ammonia to nitrite. Cells are nonmotile and the genes involved in flagellum synthesis have not been identified. The optimum growth temperature is ~25 °C and growth is observed between 15 and 35 °C. The optimum pH for growth is between 6 and 6.5. The growth is confirmed acidic conditions with a pH range between 5 and 6. Aggregated cells are formed in strongly acidic conditions (pH 2–4). Cell aggregates are acid-resistant to pH 2. The optimum ammonium concentration for growth is 100 mM at pH 6. A urease operon was identified in the genome and cells have urease activity. Cell growth is better at 50 mM NaCl than at 0 mM and does not occur at concentrations over 400 mM NaCl. The GC content of the chromosome DNA is 42.1%.

Conflict of Interest

The authors declare no conflict of interest.

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