

SHORT COMMUNICATION

Evidence for the direct oxidation of organic nitrogen to N₂ gas in the Arabian Sea

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We performed a suite of ¹⁵N incubations (¹⁵NO₂⁻, ¹⁵NO₃⁻ and ¹⁵NH₄⁺) with and without the organic-nitrogen (N) compound allylthiourea (ATU), in the suboxic waters of the Arabian Sea. Production of ²⁹N₂ in control (-ATU) incubations with either ¹⁵NH₄⁺ + ¹⁴NO₂⁻, or their analogues, ¹⁵NO₂⁻ + ¹⁴NH₄⁺, though small, confirmed the presence of anammox. In contrast, when we added ATU, along with ¹⁵NO₂⁻ and ¹⁴NH₄⁺, there was a much greater production of ²⁹N₂, with 92% of the ¹⁵N-label being recovered as ²⁹N₂ on average. Such stimulated production of ²⁹N₂ could not be due to anammox, as the addition of ATU, along with ¹⁵NH₄⁺ + ¹⁴NO₂⁻, only produced ²⁹N₂ equivalent to that in the controls. The ratios of ²⁹N₂ to ³⁰N₂ produced also precluded stimulation of denitrification. We present this as evidence for a hitherto uncharacterised metabolism potentially capable of oxidising organic-N (e.g. NH₂ groups) directly to N₂ gas at the expense of NO₂⁻.

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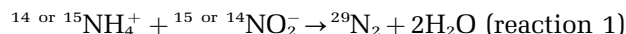
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Much of the surface water across the ocean is near air-saturation with respect to oxygen. In parts of the tropics, however, such as the eastern Pacific (North and South), Arabian Sea and Bay of Bengal, a core of permanently and severely oxygen-deplete ($1 \leq O_2 < 20 \mu M$) water (extending over > 300 m on average) can be found between 160 and 1080 m depth (Paulmier and Ruiz-Pino, 2009). Although the cores of these oxygen-minimum zones occupy only $\sim 0.8\%$ of the total ocean volume, they are responsible for a large amount of the N₂ produced (i.e., loss of fixed nitrogen (N)) on Earth (Seitzinger *et al.*, 2006; Codispoti, 2007). Given that the balance between N fixation and its removal through N₂ production is key to carbon assimilation by primary production and CO₂ modulation in the atmosphere (Falkowski, 1997), the oxygen-minimum zones are vital to the functioning of the biosphere. Evidence supports both denitrification and anammox as being responsible for this N₂ production, though discrepancies in the N-budget indicate that this two-path model is incomplete (Dalsgaard *et al.*, 2003; Kuypers *et al.*, 2005; Thamdrup *et al.*, 2006; Ward *et al.*, 2009).

Primarily we visited the oxygen-minimum zones in the central Arabian Sea to measure nitrous oxide

(Law and Owens, 1990; Nicholls *et al.*, 2007) and collected water samples from 125 to 295 m at two sites, targeting the characteristic peaks and troughs in nitrite concentration (Codispoti and Christensen, 1985; Morrison *et al.*, 1999; Nicholls *et al.*, 2007). We carried out a suite of ¹⁵N incubations with and without the addition of allylthiourea (ATU), an organic-N compound known to inhibit nitrification (Figure 1a and Supplementary Material). Although the role of nitrification in the production of nitrous oxide was minor (Nicholls *et al.*, 2007), the inhibitor measurements themselves provided us with a fortuitous and potentially important finding.

The production of N₂ via anammox has a unique 1:1 stoichiometry, which provides a very sensitive ¹⁵N ‘tool’ to detect anammox in ecosystems, with the stoichiometry proceeding according to (van de Graaf *et al.*, 1995):



The presence of anammox was confirmed at our sites by the production of ²⁹N₂ in our control incubations with either inorganic ¹⁵NH₄⁺ + ¹⁴NO₂⁻ or its analogue ¹⁵NO₂⁻ + ¹⁴NH₄⁺ (reaction 1). Although the production of ²⁹N₂ was low, at less than 70 and 20 nmol ²⁹N₂ per litre with either ¹⁵NO₂⁻ or ¹⁵NH₄⁺, respectively, after 96 h (Figures 1b and c), it was roughly equivalent to that previously reported for the Arabian Sea (e.g., ~ 18 versus $\sim 10 \text{ nmol l}^{-1}$ per day; Ward *et al.*, 2009). In contrast, when we added organic ATU along with ¹⁵NO₂⁻ and ¹⁴NH₄⁺, there was a much greater production of ²⁹N₂ at all depths at both sites, peaking at $1.42 \mu \text{mol } ^{29} \text{N}_2$ per litre over 96 h, on average

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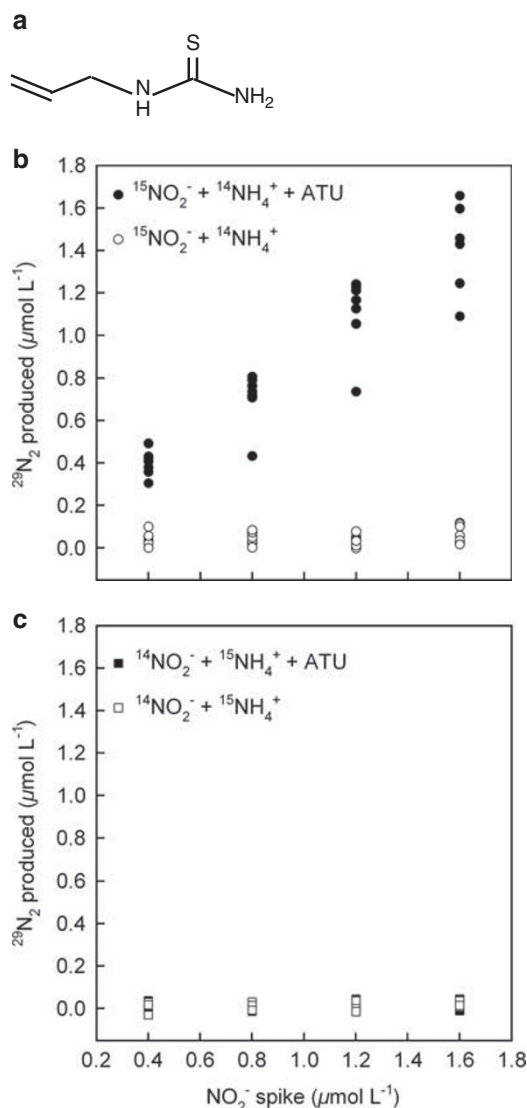


Figure 1 Organic-N and production of ²⁹N₂. (a) Molecular formula of ATU showing the primary and secondary amine groups. (b) Production of ²⁹N₂ in the presence of ¹⁵NO₂⁻ plus ¹⁴NH₄⁺ with and without ATU (filled and open circles, respectively), and (c) the same as (b), but in this case the ¹⁵N-label is on the NH₄⁺ molecule instead of the NO₂⁻ (filled and open squares, respectively). We present all of the data from each depth at both sites to show the reproducibility of the response.

(Figure 1b). Indeed, with ¹⁵NO₂⁻ the turnover of ¹⁵N was almost complete and, across all of the incubations, 92% (± 3, s.e.m., n = 28) of the ¹⁵N-label was recovered as ²⁹N₂. This production of ²⁹N₂ indicated a 1:1 pairing of ¹⁵NO₂⁻ and ¹⁴N, as in anammox (reaction 1). However, here the formation of ²⁹N₂ could not be due to the oxidation of NH₄⁺ present in the water, as the addition of ATU, along with labelled ¹⁵NH₄⁺ (and ¹⁴NO₂⁻), produced ²⁹N₂ only in small amounts equivalent to that in the controls (Figure 1c). It could be argued that the measured production of ²⁹N₂ was due to a simple organic stimulation of denitrification, for example:

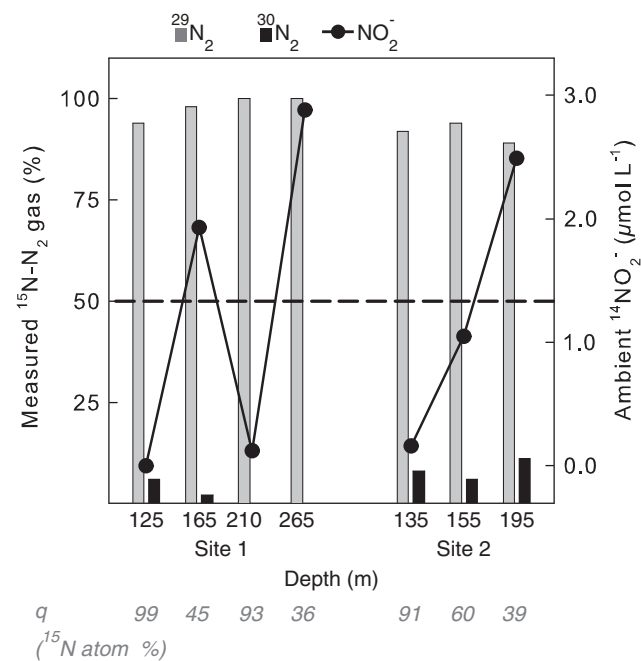
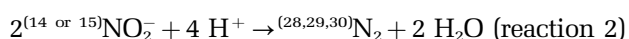


Figure 2 Predicted and measured ²⁹N₂ and ³⁰N₂ production. Measured proportions of ²⁹N₂ and ³⁰N₂ (grey and black bars, respectively) at the maximum enrichment for ¹⁵NO₂⁻ (~1.6 μmol l⁻¹) with allylthiourea. The ‘*q*’ term for each site is the ¹⁵N atom (%) of the NO₂⁻ pool (below the x axis) at each depth, which gives the ‘chances’ of denitrification producing ³⁰N₂ and reflects the mix of ambient ¹⁴NO₂⁻ (filled circles) and ¹⁵NO₂⁻ spike. Dashed line indicates the predicted maximum 50% ²⁹N₂ contribution from denitrification.

The relative amounts of ²⁸N₂, ²⁹N₂ and ³⁰N₂ in denitrification are governed by a binomial distribution reflecting the frequencies of ¹⁵N and ¹⁴N in the NO₂⁻ pool being reduced, according to:

$$[p + q]^2 = p^2 + 2pq + q^2 \quad (1)$$

where *p* and *q* are the proportions of ¹⁴N and ¹⁵N, respectively, in the NO₂⁻ pool (*p* + *q* is equal to unity) and *p*², 2*pq*, and *q*² are the predicted proportions of ²⁸N₂, ²⁹N₂, and ³⁰N₂ produced through denitrification. Accordingly, not more than 50% of the ¹⁵N added can ever be recovered as ²⁹N₂, the remainder being ³⁰N₂. Quite clearly that was not the case as, at all depths at both sites, ²⁹N₂ dominated in the ATU-amended samples, with ³⁰N₂ contributing far less than that predicted for denitrification (Figure 2).

The excess of ²⁹N₂ in the ATU-amended samples was not due to denitrification or anammox. Our observations were fortuitous and we did not fully test the nature of the reaction in the field. We have, however, since tested whether or not ATU ‘spontaneously’ breaks down in the presence of the oxidising agents nitrite and ZnCl₂ in both oxic and anoxic filter-sterilised seawater in the laboratory. After 2 weeks the ATU remained unchanged (~79 μM, Supplementary Table S1) and there was no significant production of ²⁹N₂ from ¹⁵NO₂⁻ either

with or without ATU (Supplementary Table S2). Thus, ATU appears to be stable in sterile seawater. One reasonable explanation for our observations is, therefore, that the ATU was partially metabolised and one of its amine groups provided the ¹⁴N for the production of ²⁹N₂ in the incubations with ¹⁵NO₂⁻, where the ATU acted as an analogue of urea. Urea is a main component of the complex pool of dissolved organic-N cycling in the ocean (Painter *et al.*, 2008). Some may argue that definitive proof could only be provided by the production of ²⁹N₂ after the addition of ¹⁵N-labelled dissolved organic nitrogen (DON). However, as ²⁹N₂ can only be produced through the one-to-one pairing of ¹⁴N and ¹⁵N, the production of ²⁹N₂ from ¹⁴N-DON and ¹⁵NO₂⁻ is just as equally valid.

Recently, denitrification was reported to dominate the production of N₂ gas in the central Arabian Sea, whereas we had previously argued for 'multiple pathways of N₂ production' that could not be ascribed categorically to either anammox or denitrification (Nicholls *et al.*, 2007; Ward *et al.*, 2009). Here we present evidence for an uncharacterised metabolism, potentially capable of oxidising DON (e.g. NH₂ groups) directly to N₂ gas with NO₂⁻.

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Supplementary Information accompanies the paper on The ISME Journal website (<http://www.nature.com/ismej>)