

Osteocyte Remodeling of the Perilacunar and Pericanalicular Matrix

Hai Qing^{1,2}, Lynda F. Bonewald^{1*}

¹Department of Oral Biology, University of Missouri-Kansas City, Kansas City, USA

²Department of Prosthodontics, West China College of Stomatology, Sichuan University, Chengdu, China

Abstract

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With additional functions of osteocytes being identified, the concept that osteocytes are just “static lacunar-dwelling cells” is no longer accepted. We reviewed most of the relevant literature on osteocyte’s function in the direct remodeling of the perilacunar matrix, discussing the advantages and disadvantages. Special attention was paid to how the negative researchers argue about the “osteocytic osteolysis” principle, and how the positive side addressed

the arguments. We also discussed the newly found data of osteocytic remodeling function from our group. With more biotechnology in hand, there is increased excitement in the prospect of now being able to answer the two important questions: do osteocytes have the capability to remove mineral from the perilacunar matrix and if so what are the molecular and cellular mechanisms? do osteocytes have the capability to deposit new mineral on the perilacunar matrix and if so what are the cellular and molecular mechanisms?

Keywords osteocyte, osteocytic resorption, remodeling, osteocytic osteolysis

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Introduction

Osteocytes are the most abundant and longest-living bone cells in the adult skeleton, being 10 times more abundant than osteoblasts with the potential to live as long as the host’s life time (Parfitt, 1977). Osteocytes are regularly spaced throughout the mineralized matrix encased in cave-like structures called lacunae of 15–20 μm with numerous dendritic processes within small “tunnels” called canaliculi, approximately 250–300 nm in diameter (Donahue, 2000). In this manner, osteocyte “housing” forms an extensive lacunocanalicular network in the bone, allowing the osteocytes to maintain contact through their dendritic processes and communicate with each other and with cells on the bone surface such as lining cells, osteoclasts and osteoblasts (Figure 1).

The last decade has witnessed with greater fre-

quency a rapid increase of interest in osteocytes, most likely because of new methods and state-of-the-art technology responsible for new discoveries that have led to breakthroughs in the study of osteocytes. Establishment of a cell line, MLO-Y4, captured the imagination of many investigators who now had access to a osteocyte-like cell (Kato et al., 1997) to examine osteocytic response to mechanical loading in the form of shear stress, osteocyte apoptosis, osteocyte signaling, and osteocyte communication through gap junctions and hemichannels (Bonewald, 2007). Breakthroughs were made in the identification of important markers for osteocyte differentiation which included E11/gp38, an early marker for embedding cells, Phex and dentin matrix protein 1, DMP1, important in metabolism, and *Sost*/sclerostin, a late marker for mature osteocytes (Bonewald, 2007).

Transgenic technology has also allowed advances

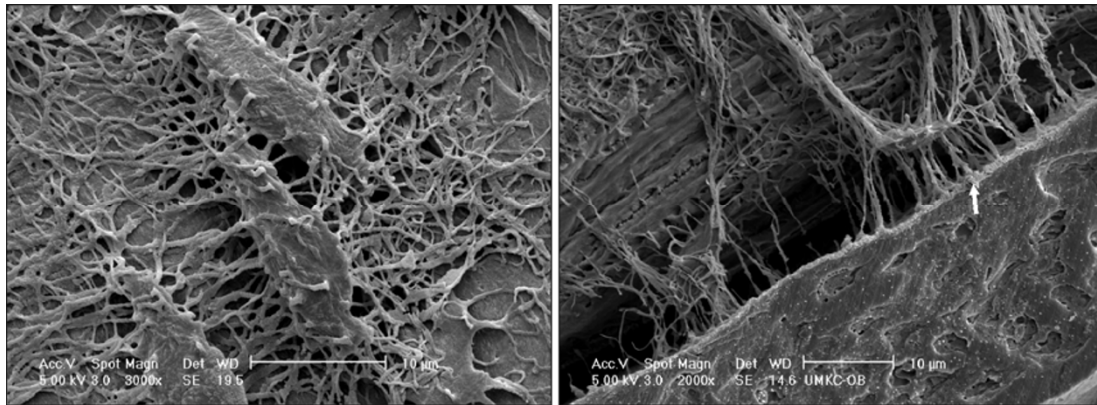


Figure 1 Osteocytes make contact with each other and cells on the surface

The images are of acid-etched resin embedded murine bone visualized by scanning electron microscopy showing the high interconnectivity of the osteocyte lacuno-canalicular system. The left panel shows the complexity of the osteocyte lacuno-canalicular network and the right panel shows osteocyte canaliculi in contact with the surface of the bone (arrow).

in the study of osteocyte functions. The promoter for *Dmp1* has been used to generate animals with green fluorescent protein, GFP, reporter labeled osteocytes (Kalajzic *et al.*, 2004) and to generate a *Dmp1-Cre* mouse to perform targeted deletion of genes in osteocytes (Feng *et al.*, 2006). The use of this promoter has shown the importance of the PTH receptor in osteocytes to maintain and regulate bone mass (O'Brien *et al.*, 2008; Divieti *et al.*, 2005), and the importance of the Wnt/ β -catenin signaling pathway in osteocyte viability and maintenance of bone mass (kramer *et al.*, 2008). However, it was the discovery of a marker for the late mature osteocyte (the gene is *Sost* and the protein is sclerostin), that began to interest the pharmaceutical industry. This osteocyte product appears to target the osteoblast to negatively regulate bone formation. Neutralizing antibody to sclerostin appears to increase bone mass and decrease bone loss, highlighting the osteocyte as a therapeutic target, something previously applied to the osteoclast and osteoblast (Li *et al.*, 2008).

While new functions of osteocytes have been discovered such as their role in phosphate metabolism (Liu *et al.*, 2006), previous functions proposed for osteocytes have been rediscovered. This review will focus on rediscovering an earlier function of osteocytes, that of being able to remove and replace their perilacunar matrix, referred to by Belanger as “osteocytic osteolysis” (Belanger *et al.*, 1967). Again, discoveries of osteocyte function regarding removal of their perilacunar matrix, is

attributable to new technology, in this case the use of new imaging technologies such as Raman spectroscopy, atomic force microscopy, and synchrotron technology.

A “local bone remodeling” function for osteocytes

The surface area of the osteocyte lacuno-canalicular system within bone is several orders of magnitude greater than the bone surface area that is directly remodeled by osteoblasts and osteoclasts (Marotti *et al.*, 1995). Therefore osteocytes have access to an extremely large area and the removal of only a few angstroms of mineral per osteocyte would have significant effects on circulating, systemic ion levels.

This feature of the lacuno-canalicular system inspired bone researchers to hypothesize as early as 100 years ago that osteocytes might have the ability to directly mobilize the bone mineral from the inner lacunar surface. In 1910, von Recklinghausen described enlarged lacunae in patients with rickets or osteomalacia (Recklinghausen, 1910), which suggested to him that pericellular “digestion” was occurring around the osteocytes (Hellersteinberg, 1951). Interest in this area increased in the 1970s as evidenced by the number of publications, however, this was followed by an obvious decrease from the 1980s to the present (Figure 2). We will discuss the rise and fall and the new rise in interest

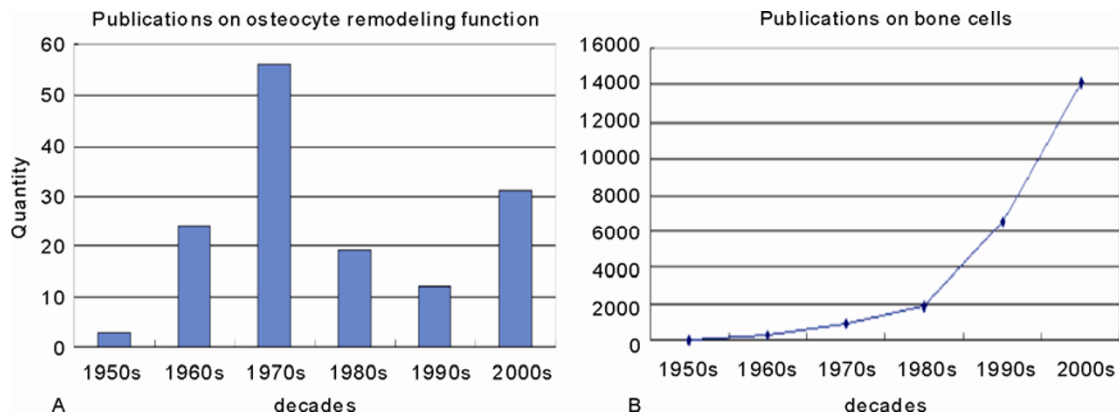


Figure 2 Summary of number of publications obtained from Pubmed

A: Using the keywords “osteocytic osteolysis”, “osteocytic resorption”, “periosteocytic osteolysis” or “osteoplastic”. B: Using the keywords “osteoblast”, “osteoclast” or “osteocyte”.

in this area. One must keep in mind that these early pioneers in osteocyte biology mainly had the tools of histology and histomorphometry. Their interpretations of their histology were insightful, but limitations left few means to prove a hypothesis. With our new technology, scientists are posed to test the hypotheses of these early pioneers.

Enlargement of osteocyte lacunae

In 1951, Heller-Steinberg showed that the areas located around lacunae and sometimes around canaliculi were positive using periodic acid-Schiff (PAS) stain in rat bone treated with parathyroid extract (Heller-Steinberg, 1951). PAS stain can detect polysaccharides and glycoproteins which are normally detected in unmineralized bone matrix or osteoid. Ruth and colleagues found basophilic matrix around osteocytes in bone from lactating rats that had been fed a calcium-free diet (Ruth, 1961). A basophilic matrix suggests what was referred to as “acidic ground material” in the matrix was mainly composed of glycoproteins. These early findings suggested that mineral removal might be taking place around the osteocytes in these animals. However, these investigators were unable to show a direct correlation between matrix staining (PAS, basophilic or toluidine blue) and mineral removal or bone resorption. It was indicated that if the cell forms bone matrix first before incorporation of mineral, this technique will give

false results regarding mineral removal. Conversely, if the osteocyte removes both the mineral and the matrix, this form of matrix staining cannot detect this type of matrix removal.

Baud was credited in 1962 to be the first person to describe rough borders of lacunar walls of osteocytes using electron microscopy again suggesting the osteocytic removal of perilacunar matrix (Baud, 1962). Later, he used microradiographs to show that parathyroid hormone can induce “osteocytic resorption” based on enlarged lacunae (Baud, 1968a; Baud, 1968b). Belanger, around this same time was also investigating this phenomena and coined the term “osteocytic osteolysis” and published a series of papers suggesting that either parathyroid hormone or low-calcium diet can induce this function in osteocytes (Belanger and Robichon, 1964; Belanger and Drouin, 1966; Belanger, 1969). In renal osteodystrophy, a histologic investigation found a significant increase in the number of enlarged and irregular lacunae in uremic subjects (Bonucci and Gherardi, 1975; Bonucci *et al.*, 1976). In 1977, Iagodovskii *et al.* sent rats into space for a 22-day space flight. They found by light and electron microscopy “wide osteocyte lacunae that could be associated with perilacunar osteolysis” (Iagodovskii *et al.*, 1977). This work found a relationship between “osteocytic osteolysis” and microgravity. Additionally, “osteocytic osteolysis” was found in the alveolar bone of hibernating ground squirrels (Haller and Zimny, 1977). In an attempt to uncover or determine the

mechanism whereby osteocytes might remove their matrix Belanger in 1963 reported the presence of protease “mainly over mature osteocytes” (Belanger and Migicovsky, 1963). Later, he reported the presence of lysosomes in the large, mature osteocytes and the stimulating effects of parathyroid hormone on these vesicles (Belanger, 1969).

What happened to “osteocytic osteolysis”?

Even though the aforementioned early investigations tried to establish the function of osteocytes in bone removal in terms of “resorption”, there was not a general acceptance of the osteocytic osteolysis concept. The reasons were ①combination of this concept “osteocytic osteolysis” with a second concept “bone flow” that proved incorrect; ②assuming that the removal of bone matrix by osteocytes would occur similarly to osteoclastic bone resorption; ③problems with sample preparation; and ④lack of additional methods to test the concept. When Krook and Belanger developed their “bone flow—osteocytic osteolysis” theory in 1970 (Krook *et al.*, 1970), considerable controversy was generated. In 1977, Parfitt published a paper disproving the “bone-flow theory” which stated that osteocytes rather than osteoclasts resorb the bone internally and then bone flowed into the osteocyte-resorbed space. This “bone-flow theory” is now known to be incorrect, but Parfitt also mentioned osteocytic osteolysis as an artifact by saying that the procedure might artificially produce the enlargement of lacunae (Parfitt, 1977). This publication, from a highly respected bone biologist, was partially responsible for the decrease of interest in “osteocytic osteolysis” after the 1970s (Figure 2). In addition, van der Plas and co-workers assumed that osteocytes would form resorption lacunae or pits similar to those formed by osteoclasts. When isolated avian osteocytes did not form resorption lacunae when cultured on sperm whale dentin, the concept of osteocyte resorption was discredited (van der Plas *et al.*, 1994). The irregular morphology, variability in spacing, and lack of consistent orientation especially in trabecular bone of osteocyte lacunae created technical challenges when using two dimensional measurements. These inconsistencies appeared to confound

two dimensional measurements that were used to predict three dimensional volumes. No consensus on a criterion for the morphological evaluation has been agreed upon. Also similar changes in lacunae can also be found in the younger osteocytes, therefore these enlarged lacunae could be due to defective mineralization of the periosteocytic matrix by the embedding osteoid osteocyte (Boyde and Jones, 1979; Parfitt, 1977).

However, three papers need to be noted in this “cold period” which used different approaches to provide more supportive evidence for osteocytic resorption. Alcobendas used microradiographs to show that osteocyte lacunae of breeding female or hibernating snakes are significantly enlarged compared to corresponding controls (virgin and non-hibernating snakes). In the snake model, the remodeling of the bone tissue does not occur throughout the life of the animal. Therefore the enlarged lacunae should come from the embedded mature osteocytes and not young, newly forming osteocytes (Alcobendas *et al.*, 1991). Nakano and co-workers used *in situ* hybridization for gene expression and immunostaining to show tartrate resistant acid phosphatase (TRAP) thought to be an osteoclast specific marker, in osteocytes. *In situ* hybridization eliminated the possibility that TRAP is diffused from the nearby osteoclast (Nakano *et al.*, 2004), a criticism of the Baylink work in 1969 (Wergedal and Baylink, 1969). Lane and colleagues compared changes in lacunar size in 6-month-old male glucocorticoid-treated and female ovariectomized mice to their respective controls and showed that lacunar size was increased in those two treated groups, significantly in the first group (Lane *et al.*, 2006). They used three dimensional reconstruction methods to obtain lacunar volume, which is a theoretically more reliable measurement to eliminate the confounding osteocyte-orientation factor.

Osteocytic replacement of perilacunar matrix/mineral

Whereas before the 1970s there was a considerable number of manuscripts reporting “osteocytic osteolysis”, there were very few reporting osteocytic replacement of this previously removed matrix. In 1971, Baylink used tetracycline labeling

to show tetracycline binding to the perilacunar matrix, which led him to suggest that osteocytes have the ability to form bone. He also reported that compared with osteoblasts which have a higher bone-forming rate, osteocytes do not play a major role in regulating serum calcium (Baylink and Wergedal, 1971). Later, Zallone used the egg-laying hen and several methods including autoradiography and tetracycline labeling to show that at least 20% of the osteocytes are active in bone formation, with relatively higher values in the metaphysis than in the diaphysis (Zamboni Zallone *et al.*, 1982; Zamboni Zallone *et al.*, 1983). Currently, the molecular mechanisms responsible for perilacunar matrix replacement are not known but are speculated to be similar to the osteoblast. It would make sense that the osteocyte could revive its previous memory as a matrix producing osteoblast, but this remains to be proven.

Implications for osteocyte remodeling of their local environment

Our group has found that lacunae are significantly enlarged during lactation in both cortical and trabecular bone in tibiae and lumbar vertebrae compared with virgin and post-weaning group, and this change is correlated with TRAP expression (Qing *et al.*, 2008). This study shows that healthy osteocytes can remove and replace their perilacunar matrix and potentially play a role in mineral homeostasis during a calcium-demanding condition such as lactation. However, in 1985, Mercer *et al.* reported that rat osteocytes do not resorb bone during lactation based on toluidine blue staining (Mercer *et al.*, 1985). However, it should be noted that toluidine blue stains proteoglycans and glycosaminoglycans (Shepard and Mitchell, 1976; Zamboni Zallone *et al.*, 1983), therefore enlargement of the lacunae might not be detected. Secondly, the metachromatically toluidine blue stained lacunae may undergo osteocytic resorption (Zamboni Zallone *et al.*, 1983). Lastly, the resolution of this measurement may not be sufficient to reveal significant differences.

Nicolella reported that the osteocyte lacuna acts as a strain concentrator effectively amplifying the macroscopic strain applied to the whole bone and

this amplification factor is a function of the local perilacuna bone tissue material properties (Nicolella *et al.*, 2006; Nicolella *et al.*, 2008). If mechanical loading is a major regulator of osteocyte function, what does this mean for the osteocyte that is hormonally regulated to remove and replace its matrix? Are the effects of mechanical loading ignored or overridden in the case of enlarged lacunae with lactation? Are the effects of mechanical loading diminished with the enlargement of lacunae with hyperparathyroidism? Do changes in lacunar size with aging affect response to mechanical load? These are questions that remain to be answered.

Additionally, the canalicular surface area is 10 times larger than lacunar surface area, which means that potentially osteocyte dendrites could also be involved in removal and replacement of bone matrix and mineral. However, to date, there is no reliable method to investigate this tiny (about 250–300 nm) and irregular structure of the canaliculi. There is the potential for synchrotron technology to be useful in this determination.

Summary

Since additional functions of osteocytes are being identified, the concept that osteocytes are just “static lacunar-dwelling cells” is no longer accepted. More and more bone biologists would speculate that osteocytes can be involved in the local bone matrix remodeling to mobilize mineral ions, but sufficient convincing evidence has still not been presented. Regarding the osteocyte local remodeling function, there are two questions needed to be addressed; Do osteocytes have the capability to remove mineral from the perilacunar matrix and if so what is the molecular and cellular mechanisms? Do osteocytes have the capability to deposit new mineral on the perilacunar matrix and if so what is the cellular and molecular mechanisms? There is increased excitement with the prospect of now being able to answer these questions.

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*Corresponding author: Lynda F. Bonewald

Address: Department of Oral Biology, University of Missouri-Kansas City, Kansas City, MO 64108, USA

Tel: 01 816 2356296 E-mail: Bonewaldl@umkc.edu
