

ORIGINAL ARTICLE

Beneficial effects of a combination of Rho-kinase inhibitor and ACE inhibitor on tubulointerstitial fibrosis induced by unilateral ureteral obstruction

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We and others recently reported that long-term Rho-kinase inhibition has renoprotective effects. This study was designed to compare the effects of an angiotensin-converting enzyme (ACE) inhibitor (imidapril), a Rho-kinase inhibitor (fasudil) and a combination of them both on renal interstitial fibrosis induced by unilateral ureteral obstruction (UUO). We also attempted to elucidate the mechanism involved. Imidapril (50 mg l⁻¹), fasudil (1 g l⁻¹) or a combination of them both was given in drinking water to mice, and their effects were compared on renal interstitial fibrosis induced by UUO. We assessed histological findings, monocyte/macrophage infiltration, myofibroblast differentiation, oxidative stress and the expression of various mRNA in the kidney by UUO. Eleven days after UUO, wild-type kidney was characterized by increased fibrotic area, dihydroethidium (DHE)-positive area, α -smooth muscle actin (SMA)-positive area, F4/80-positive area and the increased expression of various mRNA. Fasudil and imidapril similarly improved fibrotic area (–23%, –15%), DHE-positive area (–13%, –11%), α -SMA-positive area (–22%, –15%), F4/80-positive area (–42%, –34%) and the expression of various mRNA, most of which were significant ($P < 0.05$). The combination of imidapril and fasudil further improved fibrotic area (–52%), DHE-positive area (–26%), α -SMA-positive area (–33%), F4/80-positive area (–62%) and the expression of various mRNA (all $P < 0.05$ vs. monotherapy). Compared with either agent alone, the combination of an ACE inhibitor and a Rho-kinase inhibitor was more effective for the prevention of renal interstitial fibrosis because of the inhibition of transforming growth factor- β /collagen, monocyte/macrophage infiltration, myofibroblast differentiation, inflammation and the oxidative stress pathway.

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INTRODUCTION

Rho, a small guanosine triphosphate-binding protein, functions as a molecular switch in various cellular functions, including formation of stress fibers, focal adhesions, regulation of calcium ion sensitivity, production of cytokines and the regulation of G1 to S phase in cell cycle progression.^{1,2} The cellular function and signal transduction of Rho-kinase have been extensively studied. The discovery of specific inhibitors of Rho-kinase greatly advanced the knowledge of Rho/Rho-kinase pathway *in vitro* and *in vivo*.³ As Rho enhances the Ca²⁺ sensitization of vascular smooth muscle cells by inhibiting myosin phosphatase activity, the effects of Rho/Rho-kinase on the tone of blood vessels and the function of Rho in the pathogenesis of arteriosclerosis have been intensively investigated.⁴

Recent studies have revealed that the Rho/Rho-kinase pathway is involved in the pathophysiology of various cardiovascular diseases, such as hypertension, angina pectoris, myocardial infarction and pulmonary hypertension.⁵ However, few studies have investigated the function of Rho/Rho-kinase in kidney disease.⁶ Earlier studies

revealed that specific Rho-kinase inhibitors, such as Y-27632 or fasudil, significantly attenuate the tubulointerstitial fibrosis in the kidney induced by unilateral ureteral obstruction (UUO).^{7,8} More recently, we and other investigators have shown that the Rho-kinase inhibitor fasudil attenuates glomerulosclerosis in salt-induced hypertensive rats,⁹ subtotal nephrectomized spontaneously hypertensive rats,¹⁰ malignant hypertensive rats,¹¹ severely hypertensive rats,¹² aldosterone-induced glomerular injury in rats,¹³ diabetic renal injury in rats¹⁴ and N(G)-nitro-L-arginine methyl ester-induced hypertensive renal injury rats.¹⁵

Angiotensin-converting enzyme (ACE) inhibitors have been shown to attenuate renal damage including glomerulosclerosis, tubulointerstitial fibrosis and arteriosclerosis and improve kidney function.¹⁶ In the clinical setting, ACE inhibitors are a first line therapy for kidney disease and are widely used for the prevention of renal disease;¹⁷ however, their effects are limited, and a new drug or combination therapy involving ACE inhibitors is desired. Therefore, in this study, we examined the effect of a combination of an ACE inhibitor and

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a Rho-kinase inhibitor on renal fibrosis induced by UUU and compared its effects with the effects of either agent alone. We also attempted to elucidate the mechanism involved.

METHODS

This study was approved by our institutional Animal Care Committee, and all of the procedures were in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Experimental animals and design

Male C57BL/6 mice, aged 3–6 months ($n=40$), were used in this study. A UUU kidney disease model was induced in the mice (20–25 g body weight (BW), 3–6 months of age) by left ureteral ligation as described earlier.^{8,18} Sham-operated mice (the same operation without ureter ligation) were also produced. After the UUU operation, the mice were randomly assigned to four groups: untreated ($n=8$), imidapril treated (50 mg l^{-1}) ($n=8$), fasudil treated (1 g l^{-1}) ($n=8$) or a combination group ($n=8$). Imidapril (50 mg l^{-1}), fasudil (1 g l^{-1}) or a combination of them was given in drinking water to the mice for 11 days, and their effects were compared with untreated mice. After oral administration, fasudil is metabolized to hydroxyfasudil, a major active metabolite of fasudil that specifically inhibits Rho-kinase.¹⁹ We determined the dose of fasudil and imidapril according to the methods of earlier studies, which function as a renoprotective drugs in kidney disease.^{12,16} Kidney tissue samples collected at day 11 after UUU were used for histology, immunohistochemistry, renal superoxide anion detections and a real-time polymerase chain reaction (PCR) analysis study.

Tail-cuff blood pressure analysis

Systolic blood pressure (BP) and pulse rate were measured noninvasively in conscious and restrained mice as described earlier,²⁰ before and at 11 days after the operation.

Assessment of renal interstitial fibrosis

A part of obstructed kidney was excised and immersed in neutralized formalin for histological examination. The area of fibrotic lesions in the interstitium (fibrosis area) was determined on sections stained by Masson's trichrome method to stain collagen fibers (stained blue), using a computer-aided manipulator program as described earlier.¹⁸

Assessment of interstitial myofibroblast differentiation

Immunohistochemical analysis using an antibody against anti- α -smooth muscle actin (SMA), a marker of myofibroblast expression, was performed as reported earlier.¹⁸ Quantification of the α -SMA-positive area was determined using a computer-aided manipulator program as described earlier.¹⁸

Assessment of monocyte/macrophage infiltration

Immunohistochemical analysis using an antibody against F4/80, a monocyte/macrophage marker, was performed as reported earlier.¹⁸ Quantification of the F4/80-positive area was determined using a computer-aided manipulator program as described earlier.¹⁸

In situ detection of renal superoxide anion

Superoxide (O_2^-) anion generation in renal tissue was evaluated using the fluorescent dye dihydroethidium (DHE) as earlier reported.¹⁵ A part of renal tissue embedded in optimal cutting temperature was quickly frozen and cut with a cryostat ($5 \mu\text{m}$, cross-section). The sections were incubated with DHE $1 \mu\text{mol l}^{-1}$ in a light-protected humidified chamber at 37°C for 5 min. After washing the sections with phosphate buffer saline, images were acquired with identical acquisition parameters using a fluorescence microscope (Olympus AX80; Olympus, Tokyo, Japan). DHE specifically reacts with intracellular O_2^- and is converted to the red fluorescent compound ethidium, which then binds irreversibly to double-stranded DNA and appears as punctuate nuclear staining. Fluorescence intensity was quantified using Mac SCOPE software (Mitani Shoji, Fukui, Japan) and recorded as mean gray scale intensity.¹⁵

Quantification of mRNA by real-time PCR

All of the procedures used for mRNA extraction and cDNA synthesis were described in detail in earlier reports.²⁰

The gene expression levels of collagen-I, collagen-III, transforming growth factor (TGF)- β , monocyte chemoattractant protein (MCP)-1, tumor necrosis factor (TNF)- α , nicotinamide adenine dinucleotide phosphate (NADPH) subunit (p22phox, gp91 phox, p40phox, p47phox, p67phox), angiotensinogen (ANG), renin, ACE, angiotensin II type-1 receptor (AT1-R), RhoA, RhoB, RhoC and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) in the renal tissue were determined by real-time quantitative reverse-transcription polymerase chain reaction using Applied Biosystem 7700 (Applied Biosystem, Foster City, CA, USA) and specific primers as reported earlier.²⁰

Statistical analysis

All data are expressed as mean \pm s.d. Statistical comparisons among the five groups were carried out using analysis of variance. If appropriate, the data were compared with the Bonferroni's *post hoc* test for multiple comparisons. $P < 0.05$ was considered to indicate statistical significance.

RESULTS

BW, BP, pulse rate and kidney weight in the five groups

The physiological profiles of the five groups at day 11 after the UUU operation are shown in Table 1. There were no significant differences in BW among the five groups. There were no significant differences in normal side kidney weight/BW among the five groups, either. In disease side, creation of a UUU significantly decreased kidney weight/BW compared with sham-operated mice; however, there were no differences in kidney weight/BW among the four UUU groups. There were no significant differences in systolic BP or pulse rate before the operation among the five groups (data not shown). UUU operation did not change the systolic BP. Fasudil treatment did not change the systolic BP, either. As a result, there were no differences in systolic BP among the sham, untreated-UUU and fasudil-treated UUU groups. In contrast, imidapril significantly reduced the systolic BP. Cotreatment with imidapril and fasudil also significantly reduced BP.

Table 1 BW, SBP, heart rate and Kid W/BW in five groups at 11 days after UUU production

	Sham	Untreated UUU	Fasudil-treated UUU	Imidapril-treated UUU	Imidapril+fasudil-treated UUU
Number	8	8	8	8	8
SBP (mm Hg)	102 \pm 3	102 \pm 3	101 \pm 4	92 \pm 5*†‡	95 \pm 3*†‡
PR (beats per minute)	703 \pm 30	723 \pm 38	713 \pm 27	735 \pm 17	734 \pm 21
BW at 11 days	24.6 \pm 2.4	24.9 \pm 4.7	24.4 \pm 5.0	24.6 \pm 2.4	23.3 \pm 3.1
Kid W/BW (g kg^{-1}) (normal side)	7.12 \pm 0.32	7.18 \pm 0.50	7.39 \pm 1.09	6.95 \pm 0.24	7.27 \pm 0.51
Kid W/BW (g kg^{-1}) (disease side)	7.16 \pm 0.22	4.93 \pm 1.02*	5.01 \pm 0.53*	4.96 \pm 0.63*	5.59 \pm 1.22*

Abbreviations: BW, body weight; Kid W, kidney weight; PR, pulse rate; SBP, systolic blood pressure; UUU, unilateral ureteral obstruction.
* $P < 0.05$ vs. sham; † $P < 0.05$ vs. untreated UUU; ‡ $P < 0.05$ vs. fasudil-treated UUU.

Interstitial fibrosis of the kidneys in the five groups

The representative appearances of Masson's trichrome-stained sections 11 days after ureteral obstruction are shown in Figure 1. Interstitial fibrosis of the obstructed kidney was prominent in the untreated mice compared with that in the normal kidney of the sham-operated mice (Figures 1a and b). Fasudil or imidapril treatment slightly ameliorated the interstitial fibrosis compared with that in the untreated mice (Figures 1c and d). The combination of fasudil and imidapril further ameliorated renal interstitial fibrosis (Figure 1e). The tubular structure was also slightly more preserved in the fasudil- or imidapril-treated mice compared with the untreated mice and was further preserved in the mice treated with both drugs. In the measurement of the fibrous area in obstructed kidneys by computed quantitative analysis, untreated-UUO mice showed a greater fibrous area than the sham-operated mice ($P < 0.01$), and fasudil or imidapril treatment significantly reduced the fibrous area (both $P < 0.05$ vs. untreated), and the combination of fasudil and imidapril further reduced the fibrous area ($P < 0.05$ vs. fasudil- or imidapril-treated group) (Figure 1f).

Interstitial α -SMA expression in the kidneys in the five groups

The induction of interstitial myofibroblasts was assessed by immunohistochemical detection of α -SMA. Positive staining for α -SMA was seen only in vascular smooth muscle cells, but not in the interstitial space in the normal kidney of the sham-operated mice (Figure 2a). Intense immunostaining of α -SMA was observed in the peritubular interstitium in addition to the vascular smooth muscle cells of the arterioles in obstructed kidney in untreated mice 11 days after ureteral ligation (Figure 2b). Fasudil or imidapril treatment reduced the appearance of α -SMA-positive myofibroblasts in the interstitium (Figures 2c and d), and the combination of fasudil and imidapril further ameliorated it (Figure 2e). The results of the quantitative

analysis of α -SMA immunostaining are shown in Figure 2f. The α -SMA-positive area in obstructed kidneys was significantly greater in the untreated-UUO mice than in the sham-operated mice ($P < 0.01$), and fasudil or imidapril treatment significantly reduced the positive area (both $P < 0.05$ vs. untreated), and the combination of fasudil and imidapril further reduced it ($P < 0.05$ vs. fasudil- or imidapril-treated group) (Figure 2f).

Interstitial F4/80 expression in the kidneys in the five groups

Monocyte/macrophage migration was assessed by immunohistochemical detection of F4/80-positive cells. F4/80-positive cells in the interstitium are shown in Figure 3. The number of F4/80-positive cells in the interstitium was obviously increased in the untreated mice compared with the normal kidneys of the sham-operated mice (Figures 3a and b). Fasudil or imidapril treatment similarly reduced the number of F4/80-positive cells in the interstitium (Figures 3c and d), and the combination of fasudil and imidapril further reduced it (Figure 3e). The results of the quantitative analysis of F4/80-positive area are shown in Figure 3f. The F4/80-positive area in the obstructed kidneys was markedly greater in the untreated-UUO mice than in the sham-operated mice ($P < 0.01$), and fasudil or imidapril treatment significantly reduced the F4/80-positive area (both $P < 0.05$ vs. untreated), and the combination of fasudil and imidapril further reduced it ($P < 0.05$ vs. fasudil- or imidapril-treated group) (Figure 3f).

DHE staining in the kidneys in the five groups

Intracellular O_2^- levels, a marker of oxidative stress, in the renal cortex were measured using DHE and fluorescence microscopy. Representative results of the DHE staining in the five groups are shown in Figure 4. The fluorescence intensity in the interstitium was obviously

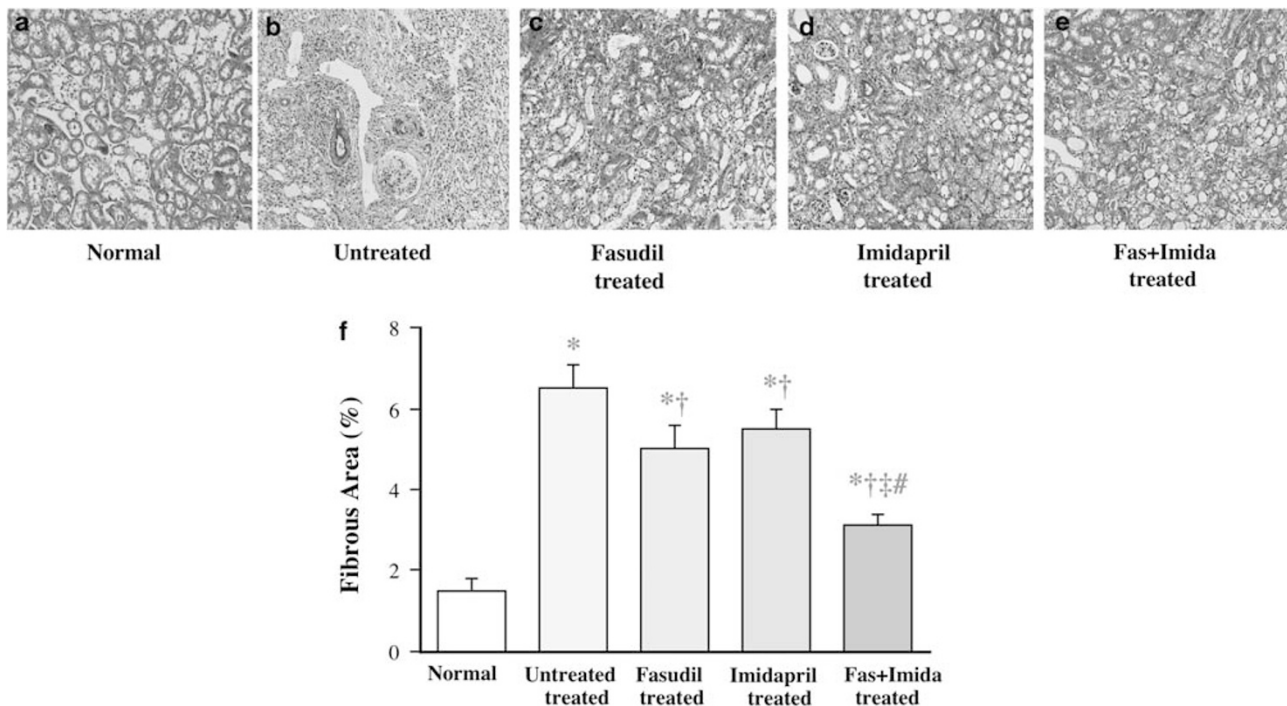


Figure 1 The morphological appearance of the interstitium as shown by Masson's trichrome staining in normal kidney in sham-operated mice (a) and in obstructed kidneys in untreated (b), fasudil-treated (c), imidapril-treated (d) and fasudil- and imidapril-treated mice (e) 11 days after the production of UUO are shown. The areas of the fibrotic lesions of the interstitium (fibrosis area) are shown in (f). Data are expressed as mean \pm s.d. The number of mice was 6–8 in each group. * $P < 0.05$ vs. normal, † $P < 0.05$ vs. untreated, ‡ $P < 0.05$ vs. fasudil-treated, # $P < 0.05$ vs. imidapril-treated mice.

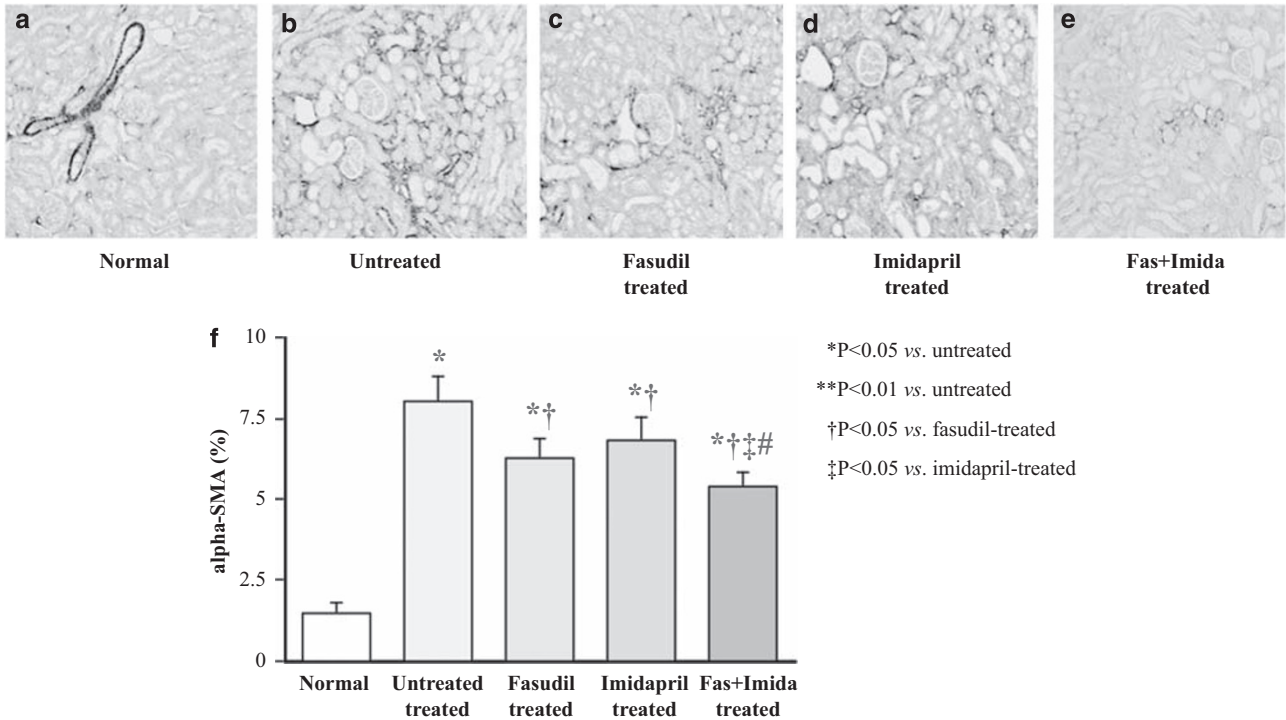


Figure 2 The α -SMA-positive cells in the interstitium in normal kidney in sham-operated mice (a) and in obstructed kidneys in untreated (b), fasudil-treated (c), imidapril-treated (d) and fasudil- and imidapril-treated mice (e) 11 days after the production of UUO are shown. The α -SMA-positive areas in the interstitium are shown in (f). Data are expressed as mean \pm s.d. The number of mice was 6–8 in each group. * P <0.05 vs. normal, † P <0.05 vs. untreated, ‡ P <0.05 vs. fasudil-treated, # P <0.05 vs. imidapril-treated mice.

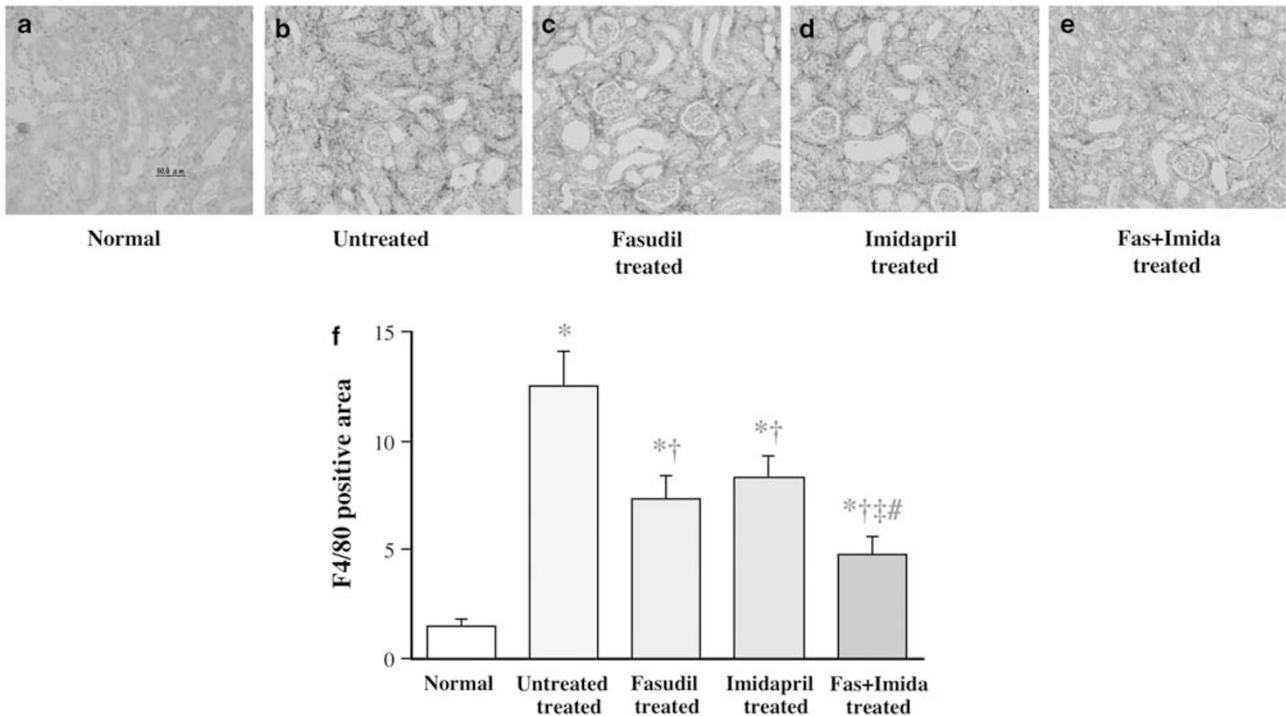


Figure 3 F4/80-positive cells in the interstitium in normal kidney in sham-operated mice (a) and in obstructed kidneys in untreated (b), fasudil-treated (c), imidapril-treated (d) and fasudil- and imidapril-treated mice (e) 11 days after the production of UUO are shown. The F4/80-positive areas in the interstitium are shown in (f). Data are expressed as mean \pm s.d. Number of mice was 6–8 in each group. * P <0.05 vs. normal, † P <0.05 vs. untreated, ‡ P <0.05 vs. fasudil-treated, # P <0.05 vs. imidapril-treated mice.

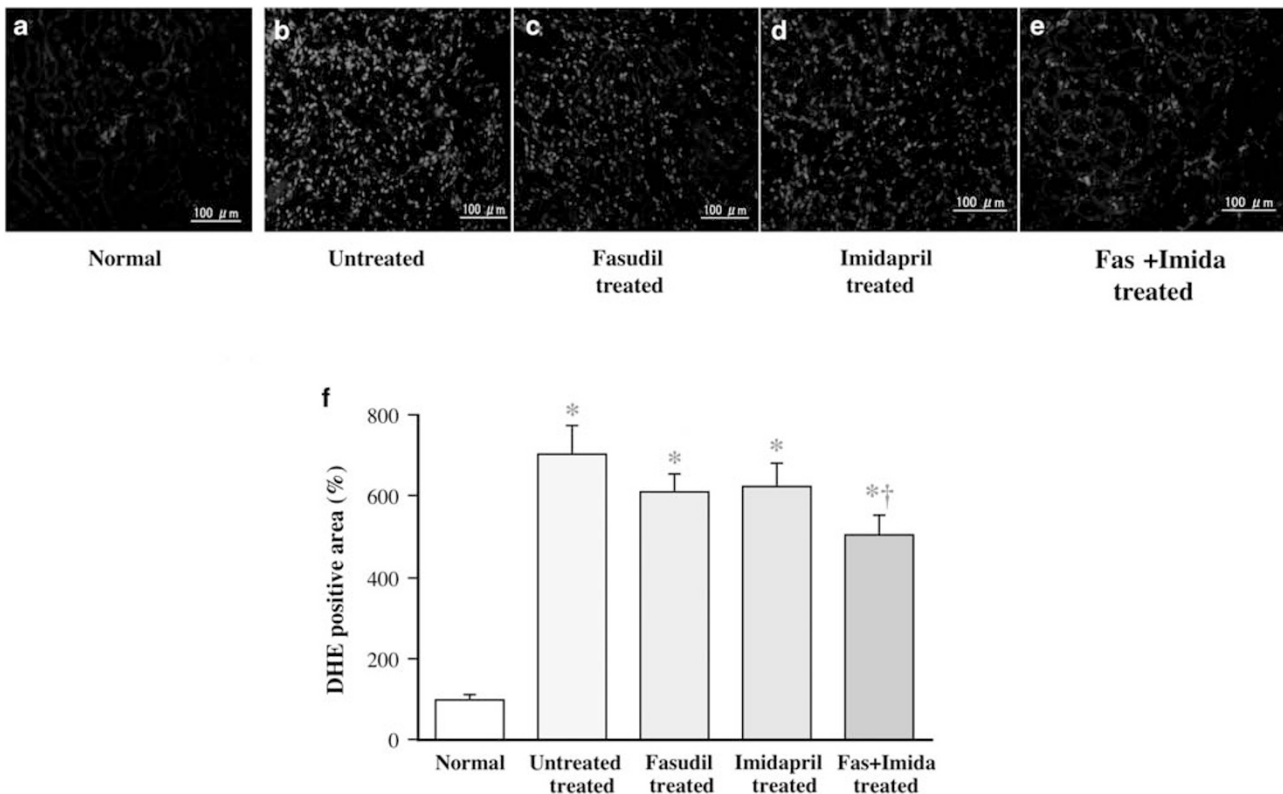


Figure 4 DHE staining was used to estimate superoxide generation. Representative DHE fluorescent photomicrographs in the interstitium in normal kidney in sham-operated mice (a) and in obstructed kidneys in untreated (b), fasudil-treated (c), imidapril-treated (d) and fasudil- and imidapril-treated mice (e) 11 days after the production of UUO are shown. The DHE-positive area in the interstitium is shown in (f) (normal kidney is shown in open bar, normalized as 100%). Data are expressed as mean \pm s.d. The number of rats was 6–8 in every group. * $P < 0.05$ vs. normal, † $P < 0.05$ vs. untreated.

greater in the untreated-UUO mice compared with the sham-operated mice (Figures 4a and b). Fasudil or imidapril treatment slightly reduced the fluorescence intensity in the interstitium (Figures 4c and d), and the combination of fasudil and imidapril further reduced it (Figure 4e). The results of the quantitative analysis of fluorescence intensity in the obstructed kidney are shown in Figure 4f. The mean fluorescence intensity values in the obstructed kidneys were markedly greater in the untreated-UUO mice than in the sham-operated mice ($P < 0.01$). Fasudil or imidapril treatment did not reduce the mean fluorescence intensity value; however, the reduction caused by the combination of fasudil and imidapril was significant ($P < 0.05$ vs. untreated group) (Figure 4f).

Renal gene expression levels in the five groups

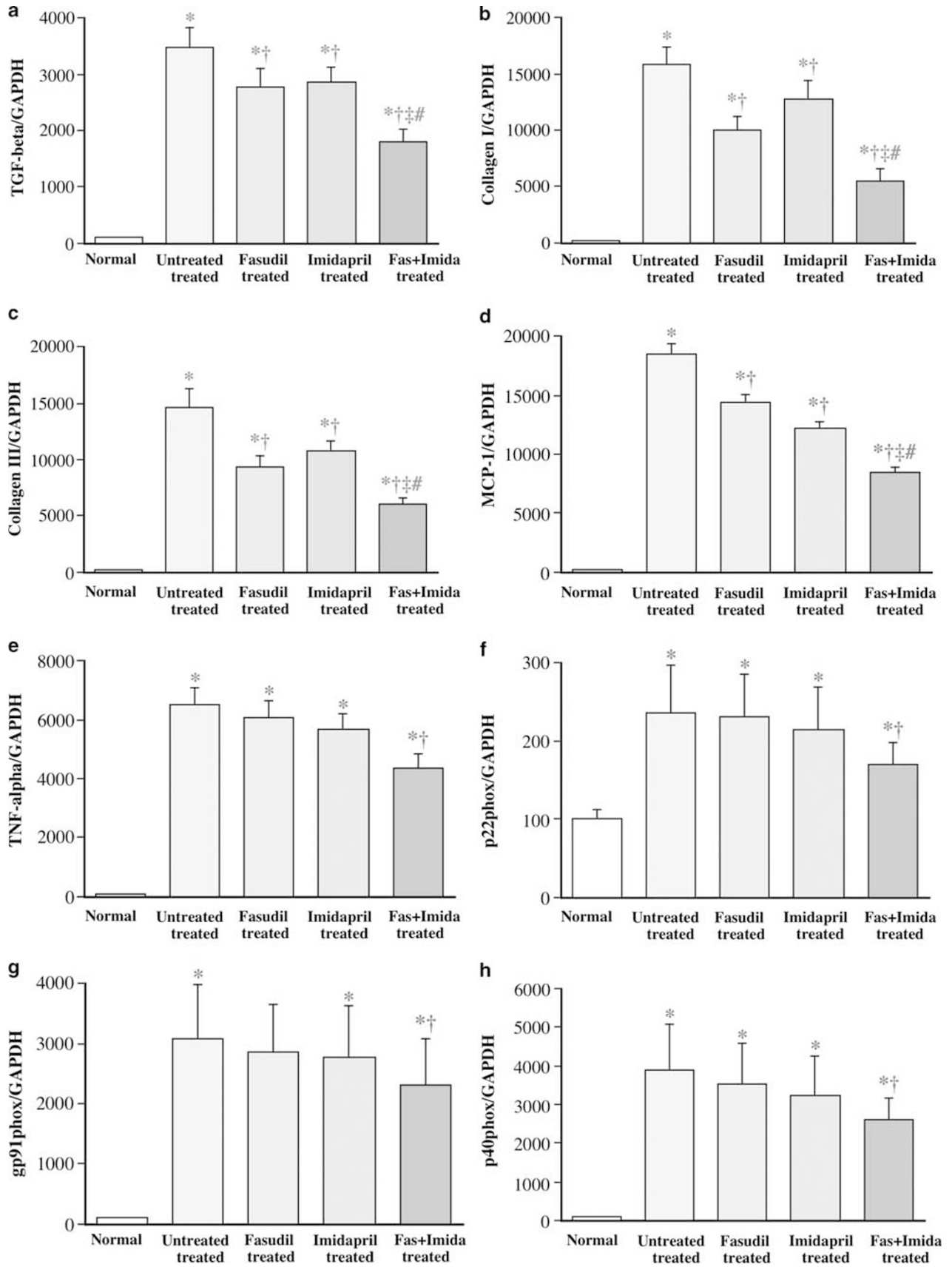
The mRNA levels of RhoA, RhoB and RhoC relative to the GAPDH mRNA levels in the renal cortex were significantly increased in obstructed kidney in UUO mice (about 1.5–2.5-fold) compared with those in the normal kidney. Long-term fasudil treatment did not change the mRNA levels of RhoA, RhoB or RhoC relative to the GAPDH mRNA levels (data not shown).

The mRNA expression levels of TGF- β , collagen-I and collagen-III, as assessed by real-time PCR at 11 days after the operation, are shown in Figures 5a–c. The mRNA expression levels of TGF- β and collagen-I and the collagen-III/GAPDH ratio were markedly increased in the obstructed kidney in UUO mice (about 30–150-fold) compared with those in the normal kidney in the sham-operated mice ($P < 0.01$) (Figures 5a–c). Fasudil or imidapril monotreatment significantly

reduced these increased mRNA expression levels compared with those in the untreated mice (both $P < 0.05$ vs. untreated), and the combination of fasudil and imidapril further reduced them ($P < 0.05$ vs. fasudil- or imidapril-treated group) (Figures 5a–c).

The mRNA expression levels of MCP-1 and TNF- α , as assessed by real-time PCR at 11 days after the operation, are shown in Figures 5d and e. The mRNA expression level of MCP-1 and the TNF- α /GAPDH ratio were markedly increased in the obstructed kidney in the UUO mice (about 60–150-fold) compared with those in the normal kidney in the sham-operated mice ($P < 0.01$) (Figures 5d and e). Fasudil or imidapril monotreatment significantly reduced the increased mRNA expression level of MCP-1/GAPDH ($P < 0.05$), but not the TNF- α /GAPDH ratio compared with those in the untreated mice. The combination of fasudil and imidapril further reduced the MCP-1/GAPDH ratio ($P < 0.05$) and significantly reduced the TNF- α /GAPDH ratio (Figures 5d and e).

The mRNA expression levels of NADPH subunits, as assessed by real-time PCR at 11 days after the operation, are shown in Figures 5f–j. The mRNA expression level of the NADPH subunits/GAPDH ratio was increased in the obstructed kidney in the UUO mice (about 2.5–30-fold) compared with that in the normal kidney in the sham-operated mice ($P < 0.01$) (Figures 5f–j). Fasudil or imidapril monotreatment did not decrease the mRNA expression level of NADPH subunits/GAPDH ratio. However, the combination of fasudil and imidapril significantly decreased the p22phox/GAPDH ratio, gp91phox/GAPDH ratio, p40phox/GAPDH ratio, p47phox/GAPDH ratio and p67phox/GAPDH ratio (Figures 5f–j).



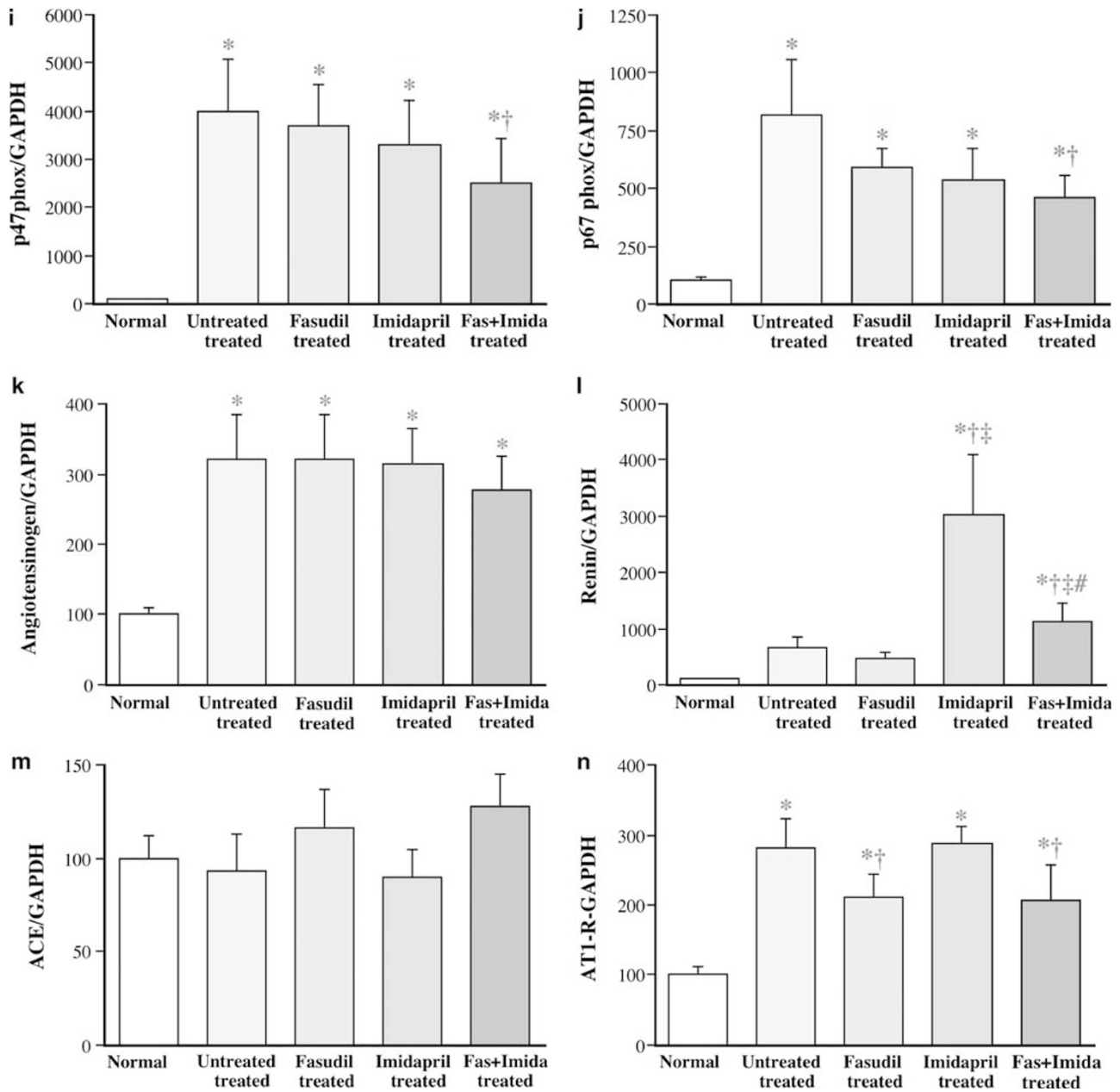


Figure 5 Continued.

Figure 5 (a–c) The mRNA expression levels of TGF- β (a), collagen-I (b) and collagen-III (c) normalized to GAPDH mRNA levels in normal kidney in sham-operated mice and in obstructed kidneys in untreated, fasudil-treated, imidapril-treated and fasudil- and imidapril-treated mice 11 days after production of the UUO are shown (normal kidney is shown in open bar, normalized as 100%). Data are expressed as mean \pm s.d. The number of rats was 6–8 in every group. * P <0.05 vs. normal, † P <0.05 vs. untreated, ‡ P <0.05 vs. fasudil-treated, # P <0.05 vs. imidapril-treated mice. (d, e) The mRNA expression levels of MCP-1 (d) and TNF- α (e) normalized to GAPDH mRNA levels in normal kidney in sham-operated mice and in obstructed kidneys in untreated, fasudil-treated, imidapril-treated and fasudil- and imidapril-treated mice 11 days after the production of UUO are shown (normal kidney is shown in open bar, normalized as 100%). Data are expressed as mean \pm s.d. The number of rats was 6–8 in every group. * P <0.05 vs. normal, † P <0.05 vs. untreated, ‡ P <0.05 vs. fasudil-treated, # P <0.05 vs. imidapril-treated mice. (f–j) The mRNA expression levels of p22phox (f), gp91phox (g), p40phox (h), p47phox (i) and 67phox (j) normalized to GAPDH mRNA levels in normal kidney in sham-operated mice and in obstructed kidneys in untreated, fasudil-treated, imidapril-treated and fasudil- and imidapril-treated mice 11 days after production of UUO are shown (normal kidney is shown in open bar, normalized as 100%). Data are expressed as mean \pm s.d. The number of rats was 6–8 in every group. * P <0.05 vs. normal, † P <0.05 vs. untreated, ‡ P <0.05 vs. fasudil-treated, # P <0.05 vs. imidapril-treated mice. (k–n) The mRNA expression levels of angiotensinogen (k), renin (l), ACE (m) and AT1-R (n) normalized to GAPDH mRNA levels in normal kidney in sham-operated mice and in obstructed kidneys in untreated, fasudil-treated, imidapril-treated and fasudil- and imidapril-treated mice 11 days after the production of UUO are shown (normal kidney is shown in open bar, normalized as 100%). Data are expressed as mean \pm s.d. The number of rats was 6–8 in every group. * P <0.05 vs. normal, † P <0.05 vs. untreated, ‡ P <0.05 vs. fasudil-treated, # P <0.05 vs. imidapril-treated mice.

The mRNA expression levels of ANG, renin, ACE and the AT1-R, as assessed by real-time PCR at 11 days after the operation, are shown in Figures 5k–n. The mRNA expression levels of ANG and the AT1-R/GAPDH ratio were increased in obstructed kidney in the UUO mice (about threefold) compared with that in normal kidney in the sham-operated mice, but renin and the ACE/GAPDH ratio were not significantly increased (Figures 5k–n). Fasudil monotherapy decreased the mRNA expression level of the AT1-R/GAPDH ratio. In contrast, imidapril monotherapy significantly increased the mRNA expression levels of the renin/GAPDH ratio (Figures 5l and n). The combination of fasudil and imidapril significantly decreased the renin/GAPDH ratio compared with imidapril monotherapy; however, it was still higher than that in the untreated mice (Figure 5l). The combination of fasudil and imidapril decreased the mRNA expression AT1-R/GAPDH ratio as observed for fasudil monotherapy (Figure 5n).

DISCUSSION

This study provides the first experimental evidence that combination therapy involving a Rho-kinase inhibitor and an ACE inhibitor is more beneficial than either agent alone for treating renal fibrosis induced by UUO.

In this study, the mRNA expressions of RhoA, RhoB and RhoC were significantly increased in the obstructed kidney in UUO mice, suggesting that a Rho/Rho-kinase pathway is activated in this model.⁷ Long-term fasudil or imidapril treatment did not change these mRNA levels, which are consistent with the earlier reports.^{7,12}

It has been well established that UUO causes renal fibrosis through many molecular mechanisms.²¹ Although renal fibrosis caused by chronic kidney disease is a clinically very important problem, only a few useful therapeutic drugs for renal fibrosis exist and its efficacy is limited. We and other investigators recently reported that Rho-kinase inhibitors attenuate glomerulosclerosis in salt-induced hypertensive rats,⁹ subtotal nephrectomized spontaneously hypertensive rats,¹⁰ severely hypertensive rats,¹² aldosterone-induced glomerular injury in rats¹³ and diabetic renal injury in rats.¹⁴ Therefore, in this study, to investigate whether the combination of a Rho-kinase inhibitor and an ACE inhibitor is beneficial for treating renal fibrosis, we used a renal fibrosis model induced by UUO.

We showed that renal fibrosis induced by UUO caused an increase in myofibroblast differentiation, as shown by the α -SMA-positive area, which causes excess deposition of the extracellular matrix and renal fibrosis.²¹ Interestingly, the combination therapy suppressed renal α -SMA-positive area to a larger extent than each monotherapy, suggesting that the combination therapy may be more effective for reducing renal myofibroblast differentiation in this model. In addition, renal fibrosis, as assessed by Mason's trichrome staining, was increased in untreated-UUO mice. Given that collagen accumulation is responsible for renal fibrosis,²² the renal fibrosis in UUO can be explained, at least in part, by the enhancing of the TGF- β /collagen pathway.²³ A great deal of investigation has been conducted to understand the cellular and molecular mechanisms of renal fibrosis in this model.²⁴ At present, one of the most important molecular mechanism of renal fibrosis in this model is increase of TGF- β .^{23–25} Earlier studies have shown that macrophage/monocyte infiltration occurred in the kidney 4 h after acute ureteral obstruction and its peak response occurred at 24 h after UUO and stabilized thereafter.²⁶ Another study also showed a highly significant correlation between the increasing number of interstitial macrophage/monocyte and the cortical TGF- β mRNA levels.²⁷ Taken together, these results suggest that macrophage/monocyte is one potential source for the increased

gene expression of TGF- β . In this study, F4/80-positive cells in the interstitium was significantly decreased in fasudil and combination group. Therefore, one possible mechanism of reduction of TGF- β mRNA levels induced by fasudil or combination may be due to the reduction of macrophage/monocyte infiltration.

Recent studies have shown that an important mechanism underlying the development of tubulointerstitial fibrosis is epithelial-to-mesenchymal transition.^{28,29} Epithelial-to-mesenchymal transition, a process by which differentiated epithelial cells undergo a phenotypic conversion that gives rise to the matrix-producing fibroblasts and myofibroblasts, is increasingly recognized as an integral part of tissue fibrogenesis after injury. Emerging evidence suggests that TGF- β initiates the transition of renal tubular epithelial cells to myofibroblasts. Thus, fasudil or combination therapy is more effective at reducing renal fibrosis than monotherapy because of inhibition of macrophage/monocyte infiltration, leading to inhibition of the TGF- β /collagen gene expression, epithelial-to-mesenchymal transition and myofibroblast differentiation pathway.

Cellular reactive oxygen species are reported to be increased in renal disease, and they have been shown to contribute to the pathogenesis of chronic kidney disease.^{30,31} Oxidative stress is known to enhance the TGF- β /collagen cascade.³² In this study, the mRNA expression of NADPH oxidase subunits such as p47phox, p40phox, p67phox, p22phox and gp91phox was increased in the UUO kidney compared with the sham-operated kidney.³³ Consistent with these findings, superoxide anion generation in the renal interstitium was increased as evaluated by DHE staining. Rho-kinase inhibitor or ACE inhibitor monotherapy did not attenuate the DHE-positive area or the mRNA expression levels of NADPH oxidase subunits. Only the combination of Rho-kinase inhibitor and ACE inhibitor significantly decreased the DHE-positive area. This reduction of the DHE-positive area appeared to be due to the change in the synthesis of superoxide by NADPH oxidase because the mRNA expression levels of NADPH oxidase subunits were only significantly reduced by the combination of the Rho-kinase inhibitor and the ACE inhibitor.

Earlier studies showed that macrophage/monocyte infiltration has a critical function in the development of renal interstitial fibrosis in a UUO model.²¹ MCP-1 mRNA expression is closely associated with macrophage/monocyte infiltration.⁷ Therefore, in this study, we measured macrophage/monocyte infiltration through the F4/80-positive area using immunohistochemistry techniques. We found that F4/80-positive area significantly increased in UUO model with the increase of MCP-1 mRNA levels. Importantly, this increase in F4/80-positive area was significantly attenuated by fasudil or imidapril monotherapy and was further reduced by the combination of fasudil and imidapril treatment. These findings, together with the fact that macrophage/monocyte infiltration can stimulate the oxidative stress, the TGF- β /collagen cascade and cytokine production,³⁴ support the notion that the reduction of renal macrophage/monocyte infiltration induced by the combination therapy is involved in the amelioration of renal fibrosis.

As shown above, fasudil and imidapril have similar actions on TGF- β /collagen cascade, myofibroblast differentiation and oxidative stress generation. However, the effect of these drugs on renal tissue RAS was different. Imidapril increased the expression of renin mRNA probably through a negative feedback mechanism; however, it did not change the level of ANG, ACE or AT1-R mRNA, which is consistent with the results of an earlier report.³⁵ Fasudil did not affect the expression of ANG, renin or ACE mRNA, but slightly decreased that of AT1-R mRNA. We earlier reported that fasudil improved renal impairment in a salt-induced hypertensive rat model without changing plasma renin

activity.⁹ Thus, regarding the effect of Rho-kinase inhibitors on the tissue RAS level, further study is required.

We have a limitation in this study. Although we have shown that mRNA level of Rho-kinase was increased, we did not measure tissue Rho-kinase activity in this study. However, we earlier showed that increased mRNA levels of Rho-A, B and C was associated with phosphorylated MYPT level, a marker of Rho-kinase activity.¹⁵ Collectively, these results suggest that Rho-kinase tissue activity was increased in this model. We did not measure ACE activity in this study, either. However, the BP reduction and increased mRNA levels of renin in ACEI treatment group strongly suggest that tissue ACE activity was deduced to be increased in UUO model.

Renal interstitial fibrosis is a progressive and potentially lethal disease caused by diverse clinical entities including urinary tract obstruction, chronic inflammation and diabetes.³⁶ However, there are few renoprotective drugs at present that have been proved by large clinical trial. In addition, their effects are limited. Therefore, a new drug or combination therapy including ACE inhibitors is desired. In this study, we examined the effect of a combination of an ACE inhibitor and a Rho-kinase inhibitor on renal fibrosis induced by UUO and found that combination therapy is more effective than either monotherapy. These results suggest that the combination of an ACE inhibitor and a Rho-kinase inhibitor may be a new therapeutic approach in the renal interstitial fibrosis in the future.

In conclusion, we first showed that the combination of a Rho-kinase inhibitor and an ACE inhibitor improved renal interstitial fibrosis more than monotherapy and that the effects of the combination therapy were associated with the inhibition of myofibroblast differentiation, the TGF- β /collagen cascade, oxidative stress and cytokine production.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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