

Dissecting the molecular mechanism of nuclear receptor action: transcription coactivators and corepressors

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Abbreviations: HRE, hormone response elements; LBD, ligand binding domain; AF2, activation function; HAT, histone acetyl transferase; HDAC, deacetylase; CBP, CREB-binding protein; TRAPs, thyroid hormone receptor associated proteins; VDR, vitamin D₃; ASC-1, activating signal cointegrator-1; RAR, retinoic acid receptor

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Introduction

Over 150 members that belong to the nuclear receptor superfamily have been discovered since glucocorticoid receptor was first reported in 1985, which primarily regulate, in a ligand-dependent manner, transcriptional initiation of the target genes by directly binding to specific DNA sequences named hormone response elements (HRE) (reviewed in Mangelsdorf *et al.*, 1995). The C-terminus of the ligand binding domain (LBD) of these proteins harbors an essential ligand-dependent transactivation function (activation function 2, AF2), whereas the N-terminus of many nuclear receptors often includes AF1. Genetic studies implicated that transcription cofactors with no specific DNA-binding activity are essential components of transcriptional regulation, which ultimately led to identify a series of nuclear receptor-interacting coregulatory proteins (reviewed in Horwitz *et al.*, 1996). Thus far, these proteins have been shown to have a few characteristic features, as summarized in Figure 1. First, they bind to target transcription factors in a ligand-dependent manner. Second, many of them are capable of directly interacting with the basal transcriptional machinery. Third, some of them exhibit enzymatic function intrinsically linked to gene regulation, such as the

nucleosomal remodeling histone acetyl transferase (HAT) or deacetylase (HDAC) activities. Thus, these proteins appear to function by either remodeling chromatin structures and/or acting as adapter molecules between nuclear receptors and the components of the basal transcriptional apparatus. Herein, we discuss the recent progress in studies of these coactivators and corepressors of nuclear receptors.

The p160 Family

A group of related proteins were found to enhance ligand-induced transactivation function of several nuclear receptors, named the p160 family. These proteins are grouped into three subclasses based on their sequence homology; i.e., SRC-1/NCoA-1 (Hong *et al.*, 1997; Torchia *et al.*, 1997; Voegel *et al.*, 1998), TIF2/GRIP1/NCoA-2 (Hong *et al.*, 1997; Voegel *et al.*, 1998), and p/CIP/ACTR/AIB1/xSRC-3 (Anzick *et al.*, 1997; Chen *et al.*, 1997; Kim *et al.*, 1998; Torchia *et al.*, 1997). A distinctive structural feature of the p160 coactivators is the presence of multiple LXXLL signature motifs (Heery *et al.*, 1997; Torchia *et al.*, 1997). The AF2 core (helix 12) was recently shown to undergo a major restructuring upon ligand binding, forming part of a "charged clamp" that accommodates p160 coactivators within a hydrophobic cleft of the receptor LBD, through direct contacts with these LXXLL motifs (Darimont *et al.*, 1998; Nolte *et al.*, 1998). Loss of function studies using antibody microinjection technique also suggested that the p160 family proteins are required for nuclear receptor functions *in vivo* (Torchia *et al.*, 1997). In addition, these factors can also interact with CREB-binding protein (CBP)/p300 via a separate domain (Kamei *et al.*, 1996; Yao *et al.*, 1996). A weak intrinsic HAT activity has been reported in SRC-1 and ACTR, suggesting that a function of these factor may involve chromatin remodeling (Chen *et al.*, 1997; Spencer *et al.*, 1997). We have recently shown that SRC-1 also mediates transactivation by a series of other transcription factors, including AP-1 (Lee *et al.*, 1998), NF κ B (Na *et al.*, 1998), SRF (Kim *et al.*, 1998a), and p53 (Lee *et al.*, 1999). In particular, SRC-1 and p/CIP were strong coactivators for p53, whereas AIB1 and xSRC-3 were repressive (Lee *et al.*, 1999). It is also noted that the p160 family of proteins has a number of uncharacterized isoforms (Chen *et al.*, 1997; our unpublished results). These results suggest a provoking hypothesis that each member of the p160 family or isoforms may regulate a specific set of target transcription

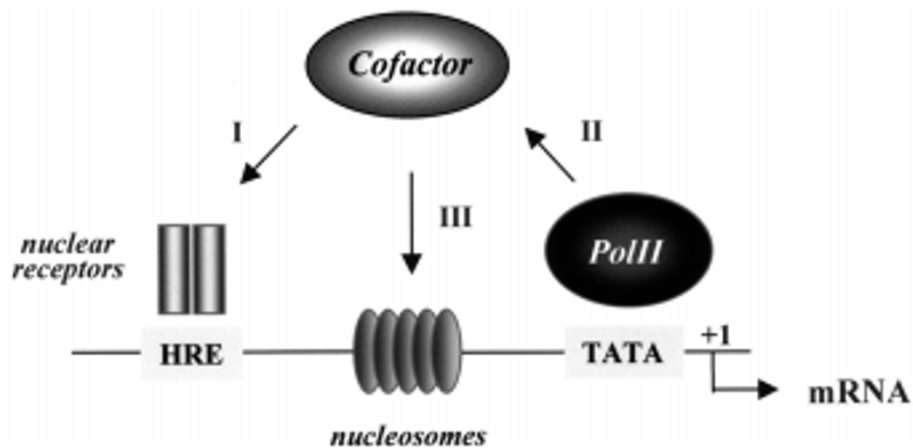


Figure 1. The role of transcriptional cofactors. Three general functions of known receptor cofactors are denoted as I, II, and III (see the text for details). HRE and +1 denote hormone response elements and transcription initiation site, respectively. Nuclear receptors, nucleosomes, cofactor and RNA polymerase II bound to TATA sequences are schematically depicted. Notably, RNA polymerase II and most cofactors exist as steady-state complex of multiple polypeptides, although each of them is represented as a single polypeptide for simplicity.

factors *in vivo*.

CBP/p300

CBP was originally isolated on the basis of its association with CREB in response to cAMP signaling, whereas its close homologue, p300, was purified as a cellular binding protein of the adenoviral protein E1A (Chrivia *et al.*, 1993; Eckner *et al.*, 1994). CBP and p300 have been implicated in functions of a large number of regulated transcription factors (reviewed in Goldman *et al.*, 1997). For nuclear receptors, the interaction with CBP/p300 is ligand- and AF2-dependent, although this direct interaction does not appear to be essential with many nuclear receptors (Westin *et al.*, 1998; Li *et al.*, 2000). In fibroblasts isolated from a p300^{-/-} mouse, however, RA-dependent transcription was severely impaired, clearly indicating that CBP/p300 are components of hormonal regulation of transcription *in vivo* (Yao *et al.*, 1998). Surprisingly, CBP and p300 harbor HAT activity (Bannister *et al.*, 1996; Ogryzko *et al.*, 1996). In addition, purified p300 was shown to potentiate ligand-induced ER function only on chromatinized template, strongly indicating that a major function of CBP/p300 could be to modify chromatin structure via histone acetylation (Kraus and Kadonaga, 1998). However, it is notable that CBP/p300 can also acetylate and functionally modulate, either in a negative or positive manner, non-histone proteins, including TFIIIE β (Imhof *et al.*, 1997), p53 (Gu and Roeder, 1997), hematopoietic transcription factor GATA-1 (Boyes *et al.*, 1998) and erythroid Krüppel-like factor (Zhang and Bieker, 1998). These results suggest that CBP/p300 may also target different aspects of gene activation, in addition to their roles in chromatin remodeling.

p/CAF

This protein was first discovered on the basis of sequence homology to the yeast HAT protein Gcn5p (Yang *et al.*, 1996). The N-terminus of p/CAF interacts with CBP and members of the p160 family, whereas interaction interface between p/CAF and nuclear receptors was different from that mediating the binding with either CBP/p300 or p160s (Blanco *et al.*, 1998; Korzus *et al.*, 1998). A core p/CAF complex was recently isolated by exploiting an affinity purification approach, which contained human homologues of the yeast ADA proteins, TAFs or TAF homologs, and p/CAF associated factor 65 α which contains histone-like structure (Ogryzko *et al.*, 1998). These results suggest a possible link between the p/CAF complex and the RNA polymerase II core machinery. This p/CAF complex bears resemblance to the GCN5/SAGA complex in yeast. In particular, other subunits of the complex facilitate p/CAF to acetylate histones in the context of nucleosomes, although p/CAF alone is inert (Grant *et al.*, 1997).

TRAP/DRIP Complexes

The thyroid hormone receptor associated proteins (TRAPs), composed of at least 9 polypeptides, were immunopurified from cells stably transfected with flag-tagged thyroid hormone receptors (Fondell *et al.*, 1996). In reconstituted *in vitro* transcription assays utilizing naked DNA templates, the TRAP complex potentiated transactivation function of liganded TR. A highly homologous DRIP (vitamin D₃ receptor (VDR) interacting protein) complex was also isolated using VDR as the affinity matrix (Rachez *et al.*, 1998), which substantially potentiated ligand-dependent transactivation function of VDR

on a chromatinized template *in vitro* (Rachez *et al.*, 1999). Interestingly, constituents of DRIP complex are almost identical to another newly discovered ARC (activator recruited cofactor) complex, which is essential for a number of other transcription factors such as SREBP, NF κ B and VP16 (Naar *et al.*, 1999; Rachez *et al.*, 1999). This TRAP/DRIP complex is recruited to the LBD AF2 core in response to ligand-binding through a single subunit (DRIP205/TRAP220/TRIP2) via a single LXXLL motif (Lee *et al.*, 1995; Naar *et al.*, 1999; Rachez *et al.*, 1999). This protein anchors the other components of the DRIP/TRAP complex to the receptor, thereby conferring hormone-dependent recruitment of what appears to be a preformed complex. In addition, TRAP/DRIP/ARC also contain part of the "Mediator complex" (Kim *et al.*, 1994), strongly suggesting their direct connection to the RNA polymerase II core machinery.

ASC-1

We have recently discovered a novel nuclear receptor-interacting coactivator, termed activating signal cointegrator-1 (ASC-1) (Kim *et al.*, 1999). ASC-1 harbors an autonomous transactivation function and binds to basal transcription factors TBP and TFIIA and transcription integrators SRC-1 and CBP/p300. The ASC-1 binding sites involve the hinge domain of nuclear receptors. Nonetheless, ASC-1 appears to require the AF2-dependent factors to function (*i.e.*, CBP/p300 and SRC-1), as suggested by the ASC-1's ability to coactivate nuclear receptors, either alone or in conjunction with SRC-1 and p300, as well as its inability to coactivate a mutant receptor lacking the AF2 core region. The ASC-1-receptor bindings, a ligand-independent event *in vitro* when tested with purified components, are either ligand-dependent *in vivo* or become ligand-dependent in the presence of NCoR *in vitro* (Kim *et al.*, 1999). Thus, ASC-1 may have more active roles in replacing NCoR/SMRT from receptors upon ligand binding. In addition, ASC-1, a nuclear protein, was found to be cytoplasmic under serum-deprivation but retained in nucleus when serum-starved in the presence of ligand or coexpressed CBP or SRC-1, suggesting additional roles for ASC-1 in cellular signal transductions (Kim *et al.*, 1999). Recently, we have purified the steady-state ASC-1 complex from HeLa nuclei (our unpublished results), which consisted of four different polypeptides of 200, 100, 65 (ASC-1), and 50 kD. Isolation of their cDNAs revealed that P200 and P50 have multiple RNA-helicase domains and RNA-binding motifs, respectively. RNA helicase A was recently isolated as a transcriptional coactivator specific for the AF1 of estrogen receptor α (Endoh *et al.*, 1999), whereas p68 RNA helicase was found to be a factor that mediates association of CBP with RNA polymerase II (Nakajima *et al.*, 1997). Surprisingly, a novel transcrip-

tional coactivator, p52, interacted not only with transcriptional activators and general transcription factors to enhance activated transcription but also with the essential splicing factor ASF/SF2 both *in vitro* and *in vivo* to modulate ASF/SF2-mediated pre-mRNA splicing (Ge and Wolfe, 1998). These results, along with the notion that pre-mRNA splicing can take place cotranscriptionally *in vivo*, suggest that, in addition to functioning as a transcriptional coactivator, ASC-1 may also act as an adaptor to coordinate pre-mRNA splicing and transcriptional activation of class II genes.

ASC-2

Activating signal cointegrator-2 (ASC-2) is another novel transcriptional coactivator molecule of nuclear receptors, recently isolated from this lab (Lee *et al.*, 1999a). Similar or identical molecules were also reported by other groups, named TRBP (Ko *et al.*, 2000), PRIP (Zhu *et al.*, 2000), and RAP250 (Caira *et al.*, 2000). ASC-2 harbors an autonomous transactivation function and binds to basal transcription factors TBP and TFIIA and transcription integrators SRC-1 and CBP/p300. Accordingly, ASC-2, a typical ligand- and AF2-dependent interactor of nuclear receptors, enhances the receptor transactivation, either alone or in conjunction with SRC-1 and p300. Interestingly, the autonomous transactivation domain of ASC-2 served as the interaction interface with CBP and the function of ASC-2 was absolutely dependent on the integrity of CBP recruitment *in vivo* (Lee *et al.*, submitted). Consistent with an idea that ASC-2 is essential for the nuclear receptor function *in vivo*, microinjection of anti-ASC-2 antibody almost completely abrogated the ligand-dependent transactivation of retinoic acid receptor (RAR) (Lee *et al.*, 1999a). More recently, we have also found that ASC-2 exists as a stable complex of multiple polypeptides *in vivo* (our unpublished results), which shows distinct chromatographic profiles from either ASC-1 (our unpublished results) or the recently described CBP/p300- and SRC-1-complexes (McKenna *et al.*, 1998). In addition, the LXXLL-type receptor interaction domain of ASC-2 acted as a potent dominant negative mutant of the peroxisome proliferator-activated receptors (PPARs), RAR and TR transactivation (Zhu *et al.*, 2000; Lee *et al.*, submitted). These results suggested that ASC-2 should directly bind to receptors and recruit CBP to mediate the receptor transactivation *in vivo*. Surprisingly, ASC-2 was identical to AIB3, a gene previously identified during a search for genes amplified and over-expressed in breast and other human cancers (Guan *et al.*, 1996). ASC-2 was recently found to mediate transactivation by a series of mitogenic transcription factors, including SRF, AP-1, NF κ B and E2F (Lee *et al.*, 2000; our unpublished results). Thus, ASC-2 may directly participate in the tumorigenesis processes when overex-

pressed.

NCoR/SMRT

Unliganded-RAR and TR bind to their target genes and repress transcription. NCoR (Chen *et al.*, 1995) and SMRT (Horlein *et al.*, 1995) were originally isolated as factors associated with the hinge domain of these nuclear receptors in the absence of ligand. More recently, however, the molecular basis of NCoR/SMRT recruitment was shown to be similar to that of coactivator recruitment, involving cooperative binding of two helical interaction motifs within the NCoR carboxyl terminus to both subunits of an RAR-RXR heterodimer (Hu and Lazar, 1999; Perissi *et al.*, 1999). The NCoR/SMRT nuclear receptor interaction motifs exhibit a consensus sequence of LXXI/HIXXXI/L, representing an extended helix compared to the coactivator LXXLL helix (Heery *et al.*, 1997; Torchia *et al.*, 1997), which was able to interact with specific residues in the same receptor pocket required for coactivator binding. Thus, discrimination of the different lengths of the coactivator and corepressor interaction helices by the nuclear receptor AF2 motif may provide the molecular basis for the exchange of coactivators for corepressors, with ligand-dependent formation of the charge clamp that stabilizes LXXLL binding sterically inhibiting interaction of the extended corepressor helix. Interestingly, NCoR and SMRT harbor transferable repression domains and associate with HDAC, consistent with the concept that histone hypoacetylation correlates with gene repression (Heinzel *et al.*, 1997; Nagy *et al.*, 1997). Two groups have reported the isolation of HDAC core complexes (mSinA and the NuRD complex) that are critically involved in this transcriptional repression (Tong *et al.*, 1998; Zhang *et al.*, 1998). A few components of the NuRD complex are also present in the recently described Sin3 complex that consists of seven polypeptides (Laherty *et al.*, 1998; Zhang *et al.*, 1998a). In particular, SAP30 was found to directly interact with NCoR (Laherty *et al.*, 1998). Antibody-blocking experiments and studies with HDAC inhibitors supported the idea that components of the NCoR/NuRD/mSin3 complex are required for repression mediated by these unliganded nuclear receptors. Thus, although NCoR doesn't appear to be stably associated with the mSin3 complex, it may serve as an adapter molecule between the core mSin3 complex and sequence-specific transcriptional repressors such as apo-nuclear receptors. Interestingly, the N-CoR/Sin3/HDAC complex is also known to mediate transcriptional repression from a wide variety of other non-receptor-mediated pathways. These include AP-1, NF κ B, SRF (Lee *et al.*, 2000a), MyoD (Bailey *et al.*, 1999), the bHLH-LZ proteins Mad and Mxi that mediate repression of myc activities and tumor suppression (Laherty *et al.*, 1997), E2F-repressive retino-

blastoma protein (Lai *et al.*, 1999), and the oncoproteins PLZF-RAR (Lin *et al.*, 1998) and LAZ3/BCL6 (Dhordain *et al.*, 1997), which are involved in acute promyelocytic leukemia and non-Hodgkin lymphomas, respectively.

Conclusion and Perspectives

Transcription coactivators and corepressors provide important insights into the mechanisms by which ligand mediates the transactivation function of nuclear receptors. In brief, ligand binding results in the dismissal of HDAC-containing corepressor complexes and the concomitant recruitment of coactivator complexes. The current model for the coactivator recruitment by nuclear receptor involves two step mechanisms, as depicted in Figure 2A. First, SRC-1 appears to be directly recruited to the liganded-receptors, which then serves as a platform to recruit CBP. Consistent with this idea, the receptor-interacting LXXLL motif located at the N-terminus of CBP was deleted without significantly affecting transactivation by RAR-RXR heterodimers, whereas the SRC-1 LXXLL motifs were absolutely essential (Li *et al.*, 2000; Westin *et al.*, 1998). These factors and associated proteins such as p/CAF, by using their HAT activities, remodel the nucleosomal structures so that TRAP/DRIP complex can replace SRC-1/CBP and bind the liganded-receptors. Subsequent recruitment of RNA polymerase II complex to TRAP/DRIP completes the second step in the nuclear receptor transactivation. However, this simple view is blurred by a large number of other nuclear receptor binding cofactor proteins or complexes, in particular with the increasing number of AF2-dependent coactivators (Horwitz *et al.*, 1996). Thus, one of the most immediate challenges to resolve is to unravel the inter-relationships between these distinct transcription cofactor proteins or complexes. These potential nuclear receptor cofactors may specifically function with different target genes as evidenced for CBP and p300 (Kawasaki *et al.*, 1998). Alternatively, these complexes may sequentially engage in different steps during the ligand-induced transactivation by nuclear receptors. For instance, we have recently suggested that ASC-2 may play a similar, essential role as SRC-1; *i.e.* direct bindings to nuclear receptors and recruitment of CBP to the receptor-ASC-2 complex (Lee *et al.*, submitted) (Figure 2B). Considering the fact that ASC-2 is expressed in relatively low amount in most cells but can be up-regulated in certain cells by various cytokines and growth factors (our unpublished results), ASC-2 may serve as an inducible factor that represents an alternative functional homologue of SRC-1. It is also noted that, from the results with ASC-1 (Kim *et al.*, 1999), RNA helicase A (Nakajima *et al.*, 1997), p68 (Endoh *et al.*, 1999), and p52 (Ge and Wolfe, 1998), transcription initiation was suggested to be directly

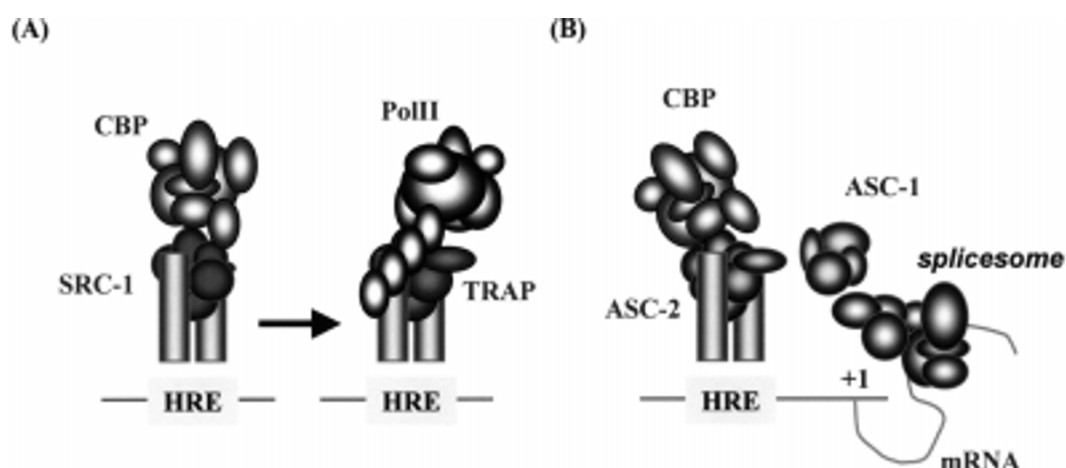


Figure 2. The model for the receptor function. **(A)** Two-step hypothesis for recruitment of coactivators is schematically explained, in which SRC-1 directly binds receptors and serves as a platform to recruit CBP. When CBP and associated proteins, using their HAT activities, remodel the nucleosomal structures, TRAP/DRIP complex is suggested to occupy receptors and subsequently recruit the RNA polymerase II complex. **(B)** ASC-2 may act as an alternative, functional homologue of SRC-1, whereas ASC-1 may link pre-RNA splicings to transcriptional initiations (see the text for details).

linked to post-transcriptional RNA processing events (such as pre-RNA splicings, as depicted in Figure 2B). Finally, MAP kinase-induced phosphorylation of SRC-1 was recently demonstrated to enhance its ability to function as a transcriptional coactivator (Rowan *et al.*, 2000), suggesting that signal transduction pathways may also modulate the assembly and/or functions of these cofactor complexes. Further characterization of these receptor coactivators and corepressors will undoubtedly unravel the fundamental mechanisms underlying the receptor function as well as the general transcription machinery.

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