LETTERS TO THE EDITOR

- McKenna KC, Chen PW (2010) Influence of immune privilege on ocular tumor development. *Ocular Immunol Inflamm* 18(2): 80–90.
- Moore AE, Rhoads CP, Southam CM (1957) Homotransplantation of human cell lines. Science 125(3239): 158–160.
- Murchison EP, Schulz-Trieglaff OB, Ning Z, Alexandrov LB, Bauer MJ, Fu B, Hims M, Ding Z, Ivakhno S, Stewart C, Ng BL, Wong W, Aken B, White S, Alsop A, Becq J, Bignell GR, Cheetham RK, Cheng W, Connor TR, Cox AJ, Feng ZP, Gu Y, Grocock RJ, Harris SR, Khrebtukova I, Kingsbury Z, Kowarsky M, Kreiss A, Luo S, Marshall J, McBride DJ, Murray L, Pearse AM, Raine K, Rasolonjatovo I, Shaw R, Tedder P, Tregidgo C, Vilella AJ, Wedge DC, Woods GM, Gormley N, Humphray S, Schroth G, Smith G, Hall K, Searle SM, Carter NP, Papenfuss AT, Futreal PA, Campbell PJ, Yang F, Bentley DR, Evers DJ, Stratton MR (2012) Genome sequencing and analysis of the Tasmanian devil and its transmissible cancer. *Cell* 148(4): 780–791.
- Murgia C, Pritchard JK, Kim SY, Fassati A, Weiss RA (2006) Clonal origin and evolution of a transmissible cancer. *Cell* 126(3): 477–487.
- Rebbeck CA, Thomas R, Breen M, Leroi AM, Burt A (2009) Origins and evolution of a transmissible cancer. *Evolution* **63**(9): 2340–2349.

*Correspondence: Dr Y Lazebnik; E-mail: yuri.lazebnik@gmail.com or GE Parris; E-mail: antimony_121@hotmail.com Published online 13 January 2015

© 2015 Cancer Research UK. All rights reserved 0007-0920/15

- Scanlon EF, Hawkins RA, Fox WW, Smith WS (1965) Fatal homotransplanted melanoma: a case report. Cancer 18: 782–789.
- Siddle HV, Kaufman J (2013) A tale of two tumours: comparison of the immune escape strategies of contagious cancers. *Mol Immunol* 55(2): 190–193.
- Simanovsky M, Berlinsky S, Sinai P, Leiba M, Nagler A, Galski H (2008) Phenotypic and gene expression diversity of malignant cells in human blast crisis chronic myeloid leukemia. *Differentiation* 76(8): 908–922.
- Strauss DC, Thomas JM (2010) Transmission of donor melanoma by organ transplantation. Lancet Oncol 11(8): 790-796.
- Torsvik Å, Rosland GV, Svendsen A, Molven A, Immervoll H, McCormack E, Lonning PE, Primon M, Sobala E, Tonn JC, Goldbrunner R, Schichor C, Mysliwietz J, Lah TT, Motaln H, Knappskog S, Bjerkvig R (2010) Spontaneous malignant transformation of human mesenchymal stem cells reflects crosscontamination: putting the research field on track - letter. *Cancer Res* 70(15): 6393–6396.
- Yang H, Lee J, Seed CR, Keller AJ (2010) Can blood transfusion transmit cancer? A literature review. Transfus Med Rev 24(3): 235–243.





http://creativecommons.org/licenses/by-nc-sa/4.0/

British Journal of Cancer (2015) 112, 1977–1978 | doi:10.1038/bjc.2015.156

Comment on: 'Evaluation of chemoresponse assays as predictive markers'

C Tian¹, M J Gabrin¹, S L Brower^{*,1} and D J Sargent²

¹Helomics Corporation, 2516 Jane Street, Pittsburgh, PA 15203, USA and ²Health Sciences Research, Mayo Clinic, 200 First Street SW, Rochester, MN 55905, USA

Sir,

We read with great interest the recent Short Communication by Korn and Freidlin (2015), which considers hypothetical examples challenging the 'match/mismatch' analysis presented in Tian et al (2014). In Tian et al (2014), we proposed and applied a novel match/mismatch analysis approach for evaluating the predictive value of a chemoresponse assay from an observational study, by investigating the assay's association with outcome. The match analysis was performed using the assay result for the administered therapy (assayed therapy = administered chemotherapy); the mismatch analysis was performed using the assay result for a randomly selected therapy from all assayed treatments for a given patient, not necessarily matching the administered therapy (assayed therapy \neq administered chemotherapy). If the match association is stronger than mismatch association, then the association is potentially drug specific and the assay may have predictive value. Using three examples in which a hypothetical chemoresponse assay is assumed to have only prognostic value, Korn and Freidlin (2015) have indicated that this analytical method may incorrectly conclude that the assay has predictive properties.

We agree with Korn and Freidlin (2015) that the match/mismatch method employed in Tian *et al* (2014) should be applied in limited circumstances and likely cannot be generalised to all chemoresponse, or more generally to all predictive biomarker assessment studies. As Korn and Freidlin (2015) point out, in situations where either (1) the treatments being considered have meaningful differences in efficacy in the unselected population or (2) specific treatment selection for a given patient is based on factors that have prognostic importance, the match/mismatch approach is inappropriate. However, we believe that neither of these cases are present in the clinical situation of recurrent ovarian cancer considered in the study by Rutherford *et al* (2013).

Specifically, in their hypothetical examples 2 and 3, Korn and Freidlin (2015) assumed different efficacies across treatments. This is inconsistent with the clinical situation in recurrent ovarian cancer (to which the match/ mismatch analysis was applied), where more than ten different drugs are recommended, but evidence from clinical trials fail to demonstrate that any one is superior to any other (National Comprehensive Cancer Network, 2014). In their hypothetical example 1, Korn and Freidlin (2015) assumed similar treatment effects for drugs A and B, but they also assumed that the patients treated by drug A were different from those treated by drug B in terms of patient prognostic profiles. In Korn and Freidlin's (2015) example, due to differences in subpopulations (pattern of assay results and sampling fraction can also be different), the match/mismatch analysis method is indeed inappropriate. However, in the study by Tian et al (2014), 15 drugs were evaluated and, as such, the heterogeneous pattern of assay results across treatments was far more complex than Korn and Freidlin's (2015) example that included two drugs. In addition, although it is possible that the treatment groups differ in prognostic profile, it is more likely, as demonstrated in clinical practice, that patients with similar prognoses have multiple therapeutic options, and there are no clear prognostic factors

Table 1. Comparison of prognostic profiles between ma	tch
and <i>mismatch</i> analyses (sensitivity vs resistance)	

	Match analysis		Mismatch analysis ^a		
	Sensitivity (28.6%)	Resistance (71.4%)	Sensitivity (25.2%)	Resistance (74.8%)	
MDRI ^b (mean)	0.68	0.10	0.71	0.11	
Age (mean, years)	57.3	63.3	58.9	62.5	
ECOG PS (%)					
0 1 or 2	68.0 32.0	70.6 29.4	69.8 30.2	71.0 29.0	
Cell type (%)					
Serous Others	65.3 34.7	69.0 31.0	65.9 34.1	69.8 30.2	
Tumour grade (%)					
1 or 2 3	15.9 84.1	23.3 76.7	17.9 82.1	23.1 76.9	
TFI ^c (%)					
<6 months ≥6 months	38.7 61.3	47.1 52.9	38.9 61.1	47.0 53.0	

Abbreviations: MDRI = multiple drug response index; TFI = treatment-free interval.

^aMismatch analysis: results representing the averages of 3000 simulations.

 ${}^{\mathbf{b}}\mathsf{MDRI}$ representing the percentage of all assayed therapies to which a patient scored as sensitive.

 ${}^{\mathsf{c}}\mathsf{TFI}$ defined as the time interval from the end of treatment until disease progression in the first-line treatment setting.

which dictate treatment decisions for individual patients. Taking all of these considerations together, after resampling, the likelihood that patients included in the mismatch analysis have similar prognostic profiles (on average), compared with those included in the match analysis, is quite high. Table 1 shows the comparison of patient prognostic profiles between match and mismatch analyses in the study by Tian *et al* (2014), demonstrating strong similarity between the two analysis groups. For the mismatch analysis used in Tian *et al* (2014), patients with heterogeneous patterns of *in vitro* response were assigned either 'sensitivity (S)' or 'resistance (R)' assay results by resampling. For match analysis, 28.6% were treated with an S drug and 71.4% were treated with an R drug, with mean multiple drug

response index (MDRI) of 0.68 vs 0.10, respectively; for mismatch, 25.2% were assigned as S and 75.8% were assigned as R, with mean MDRI of 0.71 vs 0.11, respectively. Other major prognostic factors also had similar distributions (S vs R) between match and mismatch. These results indicate that if the assay is only prognostic, the association with patient outcome should be consistent between match and mismatch analyses. In other words, the difference reported was unlikely explained by the confounding effects of prognostic factors. To further control the potential confounding factors, we also included a multivariate analysis in our study which further demonstrated the differences in patient outcome between the match and mismatch analyses (Tian *et al*, 2014).

We agree with Korn and Freidlin (2015) that evaluating the predictive value of chemoresponse assays is challenging. As with any observational study, it is impossible to entirely exclude bias, and a definitive answer relies on randomised clinical trials. However, randomised trials to evaluate predictive markers are highly challenging in rare tumour types such as recurrent ovarian cancer, particularly when a large number of treatment options are available. The length of time for patient accrual alone is likely to obfuscate clinical utility. Thus, observational studies and other non-randomised prospective studies must continue to have an important role in evaluating chemoresponse assays in this cancer type. We feel that in the appropriate circumstance, our proposed match/mismatch analysis can provide helpful information regarding an assay's potential prognostic and/or predictive value.

*Correspondence: Dr SL Brower; E-mail: sbrower@helomics.com Published online 19 May 2015

© 2015 Cancer Research UK. All rights reserved 0007-0920/15

ACKNOWLEDGEMENTS

We thank Drs Shuguang Huang and Victoria Plamadeala for their critical review and suggestions.

CONFLICT OF INTEREST

DJS received compensation for consulting work from Helomics Corporation. MJG, SLB and CT are paid employees of Helomics Corporation.

REFERENCES

- Korn EL, Freidlin B (2015) Evaluation of chemoresponse assays as predictive markers. Br J Cancer 112: 621–623.
- National Comprehensive Cancer Network (2014) NCCN Clinical Practice Guidelines in Oncology. Ovarian Cancer: Including Fallopian Tube Cancer and Primary Peritoneal Cancer. Version 3.2014.
- Rutherford T, Orr Jr J, Grendys Jr E, Edwards R, Krivak TC, Holloway R, Moore RG, Puls L, Tillmanns T, Schink JC, Brower SL, Tian C, Herzog TJ (2013) A prospective study evaluating the clinical relevance of a chemoresponse assay for treatment of patients with persistent or recurrent ovarian cancer. *Gynecol Oncol* 131: 362–367.
- Tian C, Sargent DJ, Krivak TC, Powell MA, Gabrin MJ, Brower SL, Coleman RL (2014) Evaluation of a chemoresponse assay as a predictive marker in the treatment of recurrent ovarian cancer: further analysis of a prospective study. *Br J Cancer* 111: 843–850.

BY NC SA

http://creativecommons.org/licenses/by-nc-sa/4.0/



BJC

British Journal of Cancer (2015) 112, 1978 | doi:10.1038/bjc.2015.157

Response to Comment on: 'Evaluation of chemoresponse assays as predictive biomarkers'

E L Korn^{*,1} and B Freidlin¹

¹Biometric Research Branch, Division of Cancer Treatment and Diagnosis, National Cancer Institute, MSC 9735, Bethesda, MD 20892, USA

Sir,

We thank Tian *et al* (2015) for their comments on our paper (Korn *et al*, 2015). They appear to agree with us that their analytic methods proposed in Tian *et al* (2014) do not work unless the following two assumptions hold: (1) the treatments have approximately equal efficacy in the overall population; and (2) the treatments the patients received were essentially assigned randomly (and not associated with factors that have prognostic importance). We note that these two assumptions are very strong, and, following Tian *et al* (2015), we review their plausibility in the context of recurrent ovarian cancer considered by Rutherford *et al* (2013). For assumption (1), one might question whether single-agent cisplatin or carboplatin works as well as the other treatments (e.g., combinations with platinum) on the population studied by Rutherford *et al* (2013), which contains ~45% of patients who were resistant to their initial platinum chemotherapy. If single-agent platinum drugs do not work as well, then assumption (1) is violated.

Assumption (2) allows one to treat observational data as if it were from a randomised clinical trial. It is impossible to prove that this assumption is satisfied, as there may always be important unmeasured prognostic characteristics of the patients that clinicians are implicitly using to help decide which treatments have to be given to which patients. However, it is possible to show that the assumption is questionable by finding a known important prognostic variable that is associated with the treatment the patients received. In the present case, consider the recognised important prognostic variable defined by whether patients are platinum sensitive or platinum resistant to their initial platinum chemotherapy (Jayson et al, 2014). It is known that patients with platinum-sensitive recurrent disease are more likely to be treated with combination of drugs including a platinum agent, whereas patients with platinum-resistant recurrent disease are more likely treated with a single (non-platinum) drug (Jayson et al, 2014). Indeed, this appears to be the case with data analysed by Rutherford et al (2013), where 27% of the platinum-sensitive patients received (non-platinum) single drugs whereas 50% of the platinum-resistant patients did (Table 1). This suggests a violation of assumption (2) that patients had their treatment chosen randomly.

It can be difficult to assess in any given clinical situation whether the required assumptions for the analytic methods of Tian *et al* (2014) are

*Correspondence: Dr EL Korn; E-mail: korne@ctep.nci.nih.gov Published online 12 May 2015

 $\ensuremath{\textcircled{\sc c}}$ 2015 Cancer Research UK. All rights reserved 0007 – 0920/15

Table 1. Distribution of patients cross classified by treatmentreceived and platinum status (data are abstracted fromSupplementary Table S1 of Rutherford et al (2013))

•••					
	Platinum sensitive	Platinum resistant			
Non-platinum single drugs ^a	35 (27%)	56 (50%)			
Platinum-containing combinations ^b	95 (73%)	57 (50%)			
Total	130 (100%)	113 (100%)			
^a PLD, topotecan, gemcitabine, paclitaxel. ^b Carboplatin/paxlitaxel, carboplatin/gemcitabine, carboplatin/docetaxel, cisplatin/gemci-					

tabine, cisplatin/paxlitaxel, carboplatin/topotecan.

reasonable. In particular, the required assumptions seem questionable in this recurrent ovarian cancer setting.

REFERENCES

- Jayson GC, Kohn EC, Kitchener HC, Ledermann JA (2014) Ovarian cancer. Lancet 284: 1376–1388.
- Korn EL, Freidlin B (2015) Evaluation of chemoresponse assays as predictive markers. Br J Cancer 112: 621–623.
- Rutherford T, Orr Jr J, Grendys Jr E, Edwards R, Krivak TC, Holloway R, Moore RG, Puls L, Tillmanns T, Schink JC, Brower SL, Tian C, Herzog TJ (2013) A prospective study evaluating the clinical relevance of a chemoresponse assay for treatment of patients with persistent or recurrent ovarian cancer. *Gynecol Oncol* 131: 362–367.
- Tian Ć, Gabrin MJ, Brower SL, Sargent DJ (2015) Comment on 'Evaluation of chemoresponse assays as pedictive markers'. Br J Cancer 112: 1977–1978.
- Tian C, Sargent DJ, Krivak TC, Powell MA, Gabrin MJ, Brower SL, Coleman RL (2014) Evaluation of a chemoresponse assay as a predictive marker in the treatment of recurrent ovarian cancer: further analysis of a prospective study. *Br J Cancer* 111: 843–850.



http://creativecommons.org/licenses/by-nc-sa/4.0/

 \odot