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# Bidirectional modulation of endogenous EpCAM expression to unravel its function in ovarian cancer

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**Background:** The epithelial cell adhesion molecule (EpCAM) is overexpressed on most carcinomas. Dependent on the tumour type, its overexpression is either associated with improved or worse patient survival. For ovarian cancer, however, the role of EpCAM remains unclear.

**Methods:** Cell survival of ovarian cancer cell lines was studied after induction or repression of endogenous EpCAM expression using siRNA/cDNA or artificial transcription factors (ATF) consisting of engineered zinc-fingers fused to either a transcriptional activator or repressor domain.

**Results:** Two ATFs were selected as the most potent down- and upregulator, showing at least a two-fold alteration of EpCAM protein expression compared with control. Downregulation of EpCAM expression resulted in growth inhibition in breast cancer, but showed no effect on cell growth in ovarian cancer. Induction or further upregulation of EpCAM expression decreased ovarian cancer cell survival.

**Conclusion:** The bidirectional ATF-based approach is uniquely suited to study cell-type-specific biological effects of EpCAM expression. Using this approach, the oncogenic function of EpCAM in breast cancer was confirmed. Despite its value as a diagnostic marker and for immunotherapy, EpCAM does not seem to represent a therapeutic target for gene expression silencing in ovarian cancer.

The epithelial cell adhesion molecule (EpCAM; CD326) is a transmembrane glycoprotein, overexpressed on the vast majority of carcinomas compared with healthy epithelium (Went *et al*, 2004). Epithelial cell adhesion molecule is used as a diagnostic marker, has prognostic value for some tumours (Went *et al*, 2008) and even serves as a therapeutic target in antibody-based therapies (Baeuerle and Gires, 2007). Recently, EpCAM has been identified as a marker for cancer-initiating stem cells (Visvader and Lindeman, 2008) and was shown to be involved in the Wnt-signaling pathway (Maetzel *et al*, 2009). Upon intramembrane

proteolysis of EpCAM, the intracellular domain functions with  $\beta$ -catenin as part of a transcriptional complex inducing *c-myc* and cyclin A/E expression. In breast cancer, high EpCAM expression is associated with poor prognosis, (Spizzo *et al*, 2004), which is consistent with the finding that induction of EpCAM in the EpCAM-negative breast cancer cell line Hs578T showed increased proliferation compared with the empty vector control (Gostner *et al*, 2011). In addition, RNAi-based silencing of EpCAM expression in breast cancer cell lines reduced the oncogenic potential of the treated cells (Osta *et al*, 2004) and EpCAM

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expression rescued by cDNA constructs increased cell invasion (Sankpal *et al*, 2011).

Despite this oncogenic function of EpCAM in breast carcinogenesis, for several tumour types the biological role of EpCAM is far from clear (van der Gun *et al*, 2010). Clinical observations in ovarian cancer suggest that for this tumour type EpCAM overexpression correlates with decreased overall survival, especially in patients with FIGO stage III/IV (Spizzo *et al*, 2006), although this could not be confirmed by independent studies (Kim *et al*, 2003; Heinzelmann-Schwarz *et al*, 2004). In addition, EpCAM has been described as a marker of human ovarian cancer stem/progenitor cells (Meirelles *et al*, 2012) and introduction of human EpCAM cDNA in EpCAM-negative mouse ovarian stem-like tumour cells enhanced the tumour-initiating ability (Motohara *et al*, 2011). Although these studies are far from definitive, they do suggest that downregulation of endogenous EpCAM expression might decrease the oncogenic potential in ovarian cancer.

To our knowledge, so far no data exist on EpCAM expression modulation in human ovarian cell culture systems in contrast to breast and many other tumour types. To investigate the functional role of EpCAM, a method to modulate bidirectionally endogenous expression levels would provide biologically relevant insights. To this end, artificial transcription factors (ATFs) can be constructed by fusing a gene-specific DNA-binding domain to transcriptional modulators (Uil *et al*, 2003). DNA-binding domains often used are engineered six zinc-fingers targeting an 18-base-pair unique sequence, thereby, enabling specific targeting of virtually any gene (Sera, 2009). Artificial transcription factors are appealing because they allow for both up- (Beltran and Blancafort, 2011) and downregulation (Stolzenburg *et al*, 2012) of gene expression, by fusing an activator or repressor to the zinc-fingers. Furthermore, compared with siRNA approaches where many mRNA copies have to be targeted, only two gene copies need to be targeted by ATFs. In addition, compared with gene delivery approaches, which often involve the expression of only one cDNA variant, ATFs can induce all splice variants in their natural ratios. Importantly, using this approach, epigenetic effector domains can be targeted to the gene of interest to induce mitotically stable gene expression modulation (De Groote *et al*, 2012). In this study, bidirectional endogenous EpCAM modulation by ATFs was exploited to investigate the biological role of EpCAM overexpression in ovarian cancer.

## MATERIALS AND METHODS

**Cell culture.** HEK293T cells, and the cancer cell lines HeLa (cervix), MCF7, MDA-MB231, SKBR3 (breast) and OVCAR3, CaOV3, SKOV3 and A2780 (ovarian) were cultured in DMEM (BioWhittaker, Walkersville, MD, USA) with 50  $\mu\text{g ml}^{-1}$  gentamicin sulphate, 2 mM L-glutamine and 10% FBS.

**ATF retroviral transduction.** Epithelial cell adhesion molecule targeting zinc-finger proteins (ZFPs): one 3-finger (3F-ZF<sub>A</sub>) and two 6-fingers (referred to as ZF<sub>A</sub> and ZF<sub>B</sub>) (Gommans *et al*, 2007) were subcloned into the retroviral vector pMX-IRES-GFP (pMX) (Beltran and Blancafort, 2011) containing the activator VP64 (a tetrameric repeat of herpes simplex VP16) (Beltran and Blancafort, 2011) or the repressor SKD (Kruppel-associated box) (Stolzenburg *et al*, 2012). Retroviral particles were produced by co-transfection of pMX with the VSV envelope protein (pMD2.G) and the gag/pol proteins (pMDLg/pRRE) into HEK293T cells using CaPO<sub>4</sub>. After 2 and 3 days, the viral supernatant was used to transduce the host cell lines in the presence of 5  $\mu\text{g ml}^{-1}$  polybrene (Sigma, St Louis, MO, USA). Transduction was optimized by, for example, diminishing toxicity to 293 producer cells (lowering the amount of plasmids in the transfection) and by varying the number of host cells seeded. Cells were harvested for analysis, 4 days after

the first transduction. As the IRES-GFP present in the pMX vector showed a relative low GFP signal (Beerli *et al*, 2000), co-transduction with a strong GFP reporter was used to monitor toxicity by downregulation of EpCAM expression in SKBR3 cells.

**siRNA or cDNA transfection.** siRNA-EpCAM 750 ng (sense, 5'-GGAGAUCACAACGCGUUAUUU and antisense, 5'-AUAA CGCGUUGUGAUCUCCUU) or irrelevant-siRNA (irr-siRNA) (1022076; Qiagen, Venlo, Netherlands) or plasmid DNA (generously provided by Giepmans, UMCG and Cirulli (Washington, DC, USA)) (Schnell *et al*, 2012) was complexed in 200  $\mu\text{l}$  HBS with 20  $\mu\text{l}$  SAINT-2:DOPE (SD; 0.75 mM) (Synvolux, Groningen, Netherlands) and pipetted onto the SKOV3 or A2780 cells.

**Gene expression analysis.** Q-RT-PCR was performed (ABIPrism 7900HT, Applied Biosystems, Nieuwekerk, Netherlands) for EpCAM (Hs00158980\_m1, Applied Biosystems) and GAPDH (F, 5'-CCACATCGCTCAGACACCAT-3'; R, 5'-GCGCCCAATA CGACCAAAT-3'; probe: 5'-CGTTGACTCCGACCTTCACCTT CCC-3'; Eurogentec, Maastricht, Netherlands) in triplicate. Results are shown as relative expression compared with GAPDH levels, using  $\Delta\text{Ct}$  method (pMX was set at 1). Epithelial cell adhesion molecule protein was detected by APC-CD326 (BioLegend, Uithoorn, Netherlands). The mean fluorescence intensity (pMX was set at 100%) was measured on a Calibur flow cytometer (Beckton Dickinson Biosciences, San Jose, CA, USA).

**Cell growth assays.** Transduction was carried out directly in 96-well plates and cell viability MTT assay (M2128; Sigma) was performed at days 4, 5 and 6 after transduction, or transductions were carried out in 6-well plates and host cells were harvested at day 4 after transduction, replated at equal numbers in 96-well plates and MTT assay was performed for the four following days. After 3.5 h of MTT incubation, plates were centrifuged, the medium was aspirated and 200  $\mu\text{l}$  DMSO was added. Absorption was measured at 560 nm (Varioskan microplate spectrophotometer; Thermo Scientific, Breda, Netherlands), and background was subtracted at 670 nm. To estimate cell numbers after treatment, cells were fixed and stained with methanol/acetic acid/water mixture (50%, 20% and 30% respectively), containing 0.01% Coomassie brilliant blue (Sigma) visualizing protein content. In addition, clonogenic survival assay was performed by replating cells in 6-well plates at day 4 after transduction, colonies were allowed to grow for 15–21 days and visualized as described above.

## RESULTS

**Bidirectional modulation of EpCAM expression.** To identify which ATF could effectively modulate EpCAM expression, ATFs were screened in a panel of carcinoma cell lines by using sub-optimal retroviral transductions (Supplementary Figure 1). This screening approach identified ZF<sub>A</sub>SKD and ZF<sub>B</sub>VP64 as the most potent down- and upregulators of EpCAM expression, respectively. ZF<sub>B</sub>VP64 showed no effect on Her2neu and ICAM expression (data not shown), whereas an ATF designed to target the *Her2neu* gene (Beerli *et al*, 2000) effectively up- (E2C-VP64) or downregulated (E2C-SKD) Her2neu expression in HeLa, but showed no effect on EpCAM expression (Supplementary Figure 1D and E).

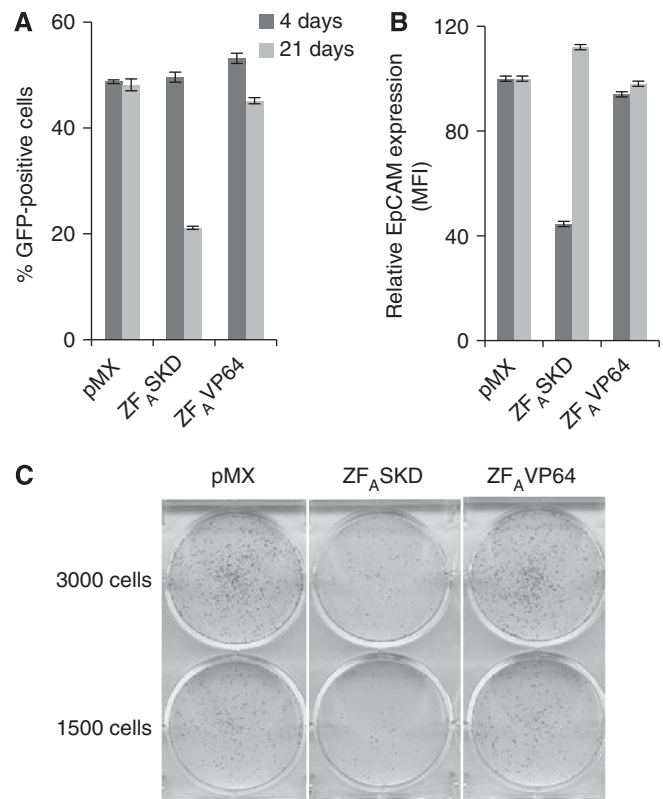
**Functional validation of downregulation of EpCAM expression by ATFs.** Downregulation of EpCAM expression by RNAi in breast cancer reduces the oncogenic potential (Osta *et al*, 2004; Sankpal *et al*, 2011). Therefore, we validated that downregulation of EpCAM expression by ATFs in breast cancer also induced similar phenotypic alterations. SKBR3 cells were co-transduced with ZF<sub>A</sub>SKD and a strong GFP reporter vector to track the percentage of transduced cells. As negative controls, ZF<sub>A</sub>VP64 and

pMX (empty vector control) were used. At 4 days after transduction, all conditions displayed approximately 50% GFP-positive cells (Figure 1A). Compared with pMX, ZF<sub>A</sub>SKD-transduced cells showed a 2.3-fold reduction in EpCAM protein expression, whereas the EpCAM expression of ZF<sub>A</sub>VP64- and pMX-transduced cells was similar (Figure 1B). After normalizing for cell numbers and subculturing, ZF<sub>A</sub>SKD-transduced cells proliferated less rapidly than the cells transduced with pMX or ZF<sub>A</sub>VP64, requiring less cell passaging. Indeed, after 17 days of subculturing, the ZF<sub>A</sub>SKD-transduced cell population only contained 20% GFP-positive cells, whereas the cells transduced with pMX or ZF<sub>A</sub>VP64 still displayed 50% GFP-positive cells (Figure 1A). This finding indicates that the unaffected EpCAM-expressing cell population overgrew the ZF<sub>A</sub>SKD-expressing SKBR3 population, which is also reflected in the restored EpCAM expression of the cell population (Figure 1B). The reduced proliferation displayed by the reduced EpCAM-expressing breast cancer cells was also confirmed by a colony-forming assay. The number of colonies formed by SKBR3 cells expressing ZF<sub>A</sub>SKD was clearly less than the number of colonies formed by the cells transduced with pMX and ZF<sub>A</sub>VP64 (Figure 1C).

**Biological role of EpCAM expression in ovarian cancer.** As the biological role of EpCAM in ovarian cancer is unclear (van der Gun *et al*, 2010), we set out to provide such insights by bidirectionally modulating EpCAM expression in intermediate EpCAM-expressing ovarian cancer cells (SKOV3). The transduction protocol was improved to yield at least 80% of GFP-positive ovarian cancer cells expressing ZF<sub>A</sub>SKD or ZF<sub>B</sub>VP64. Compared with cells transduced with pMX, ZF<sub>A</sub>SKD-expressing cells displayed a two-fold reduction of EpCAM expression on RNA ( $0.5 \pm 0.1$ ;  $P < 0.001$ ) and on protein level ( $41 \pm 2\%$  expression left;  $P < 0.001$ ) (Figure 2A). The ZF<sub>A</sub>VP64- and ZF<sub>B</sub>VP64-transduced cells demonstrated increased EpCAM protein expression ( $213 \pm 22\%$ ;  $P < 0.01$  and  $340 \pm 33\%$ ;  $P < 0.001$ ), respectively). Cells transduced to express ZF<sub>B</sub>NoED (NoED = no effector domain) without an effector domain also exhibited upregulation ( $178 \pm 4\%$ ;  $P < 0.001$ ), which was not observed for ZF<sub>A</sub>NoED (Figure 2A).

Because ZF<sub>A</sub>SKD and ZF<sub>B</sub>VP64 modulated EpCAM expression most effectively, these ATFs were further selected to investigate the phenotypical change due to EpCAM gene downregulation *vs* overexpression. Unexpectedly, at day 4 after transduction, not ZF<sub>A</sub>SKD- but ZF<sub>B</sub>VP64-transduced cells showed less cell survival as shown by staining (Figure 2B, left inset) compared with the cells transduced with pMX (empty vector control). Cells transduced with pMX, ZF<sub>B</sub>NoED or ZF<sub>A</sub>SKD showed comparable survival. This observation was confirmed by cell viability MTT assays performed on days 4, 5 and 6 after transduction: less cell survival was observed after transduction with ZF<sub>B</sub>VP64, whereas pMX, ZF<sub>A</sub>SKD and ZF<sub>B</sub>NoED displayed the same cell growth rate (Figure 2B, left). Also, after harvesting the different transduced cell populations at day 4 after transduction, MTT assays performed on the equal numbers of replated cells showed exactly the same trend: ZF<sub>B</sub>VP64-transduced cells did poorly attach to the wells and were hardly capable to proliferate, whereas ZF<sub>A</sub>SKD and ZF<sub>B</sub>NoED showed the same growth rate as pMX (Figure 2B, right). Furthermore, enforced ectopic cDNA expression of EpCAM did not result in growth induction of ovarian cells (Supplementary Figure 2A). To exclude that the expression of VP64 itself was toxic for the cells, transductions were performed with an irrelevant ZFP fused to VP64 (ZF<sub>irr</sub>VP64), with ZF<sub>A</sub>VP64 and the controls ZF<sub>A</sub>NoED and pMX. Again, ZF<sub>B</sub>VP64-transduced cells clearly showed less cell survival (Supplementary Figure 2B), whereas all other constructs displayed comparable cell numbers with pMX.

**siRNA-based silencing of EpCAM expression in ovarian cancer has no effect on cell growth.** The biological effect of increased EpCAM expression was in contrast with our initial hypothesis that

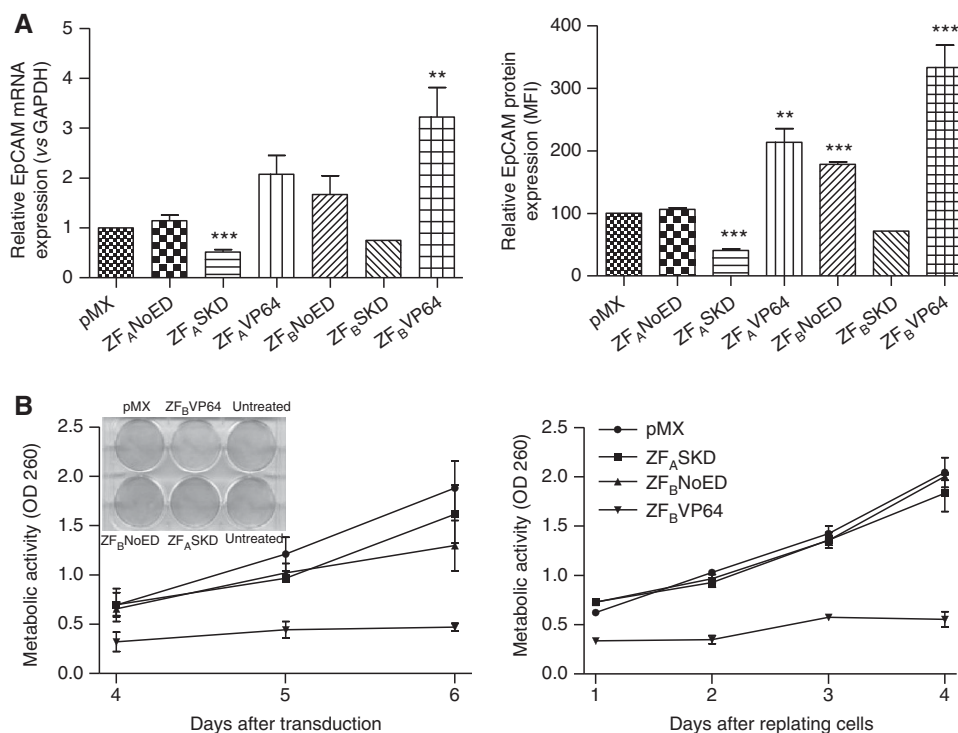


**Figure 1.** Downregulation of EpCAM expression by ATFs inhibits cell growth in breast cancer cells. SKBR3 cells were transduced to express the indicated ATFs. At days 4 and 21 after transduction, the percentage of GFP-positive (A) and the relative EpCAM protein expression (B; pMX set at 100) was measured by flow cytometry. (C) Clonogenic survival assay: 4 days after transduction cells were equally replated in 6-well plates and allowed to grow colonies for 16 days when surviving colonies were stained. MFI, mean fluorescence intensity.

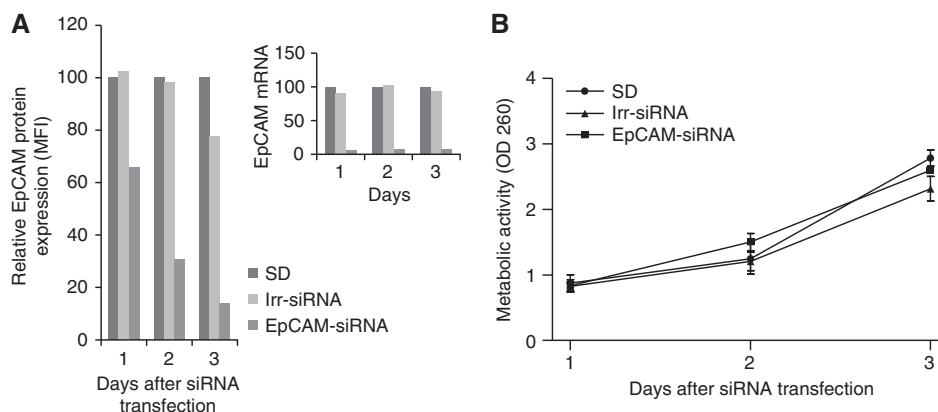
reduced EpCAM expression in ovarian cancer would result in growth inhibition. To confirm that indeed downregulation of EpCAM expression had no effect on cell proliferation, we validated our finding with the conventional approach of silencing EpCAM expression by siRNA. After transfection of EpCAM-siRNA into SKOV3 cells, EpCAM RNA expression was almost completely silenced and protein expression was reduced by 95% (Figure 3A), whereas irr-siRNA had no effect on EpCAM expression. Despite the efficient downregulation of EpCAM, again equal growth rates were observed for EpCAM-siRNA- and irr-siRNA-treated cells (Figure 3B).

**Increased EpCAM expression levels correlated with cell growth inhibition.** To investigate whether the growth inhibition associated with induced EpCAM overexpression could be phenotypically rescued, ZF<sub>B</sub>VP64-transduced SKOV3 cells were transfected with EpCAM-siRNA. At day 1 after siRNA transfection, the ZF<sub>B</sub>VP64-induced 3.3-fold upregulation of EpCAM protein expression was reduced by the combination with siRNA to only 1.8-fold upregulation compared with ZF<sub>B</sub>VP64 and irr-siRNA treatment (Figure 4A). At day 3, only 16% EpCAM expression compared with mock-treated cells was left. Despite the efficient downregulation of EpCAM expression at day 3, this reduction could not rescue the growth inhibition caused by the initial ATF-induced EpCAM overexpression (Figure 4B).

As downregulation of EpCAM expression after upregulation of EpCAM expression had no effect on cell survival, we investigated whether we could prevent the induced upregulation by simultaneous downregulation of EpCAM expression. SKOV3 cells were transduced with a constant amount of ZF<sub>B</sub>VP64 virus supernatant



**Figure 2.** Phenotypical effect of up- and downregulation of EpCAM expression in ovarian cancer. SKOV3 cells were transfected to express the indicated ATFs in 6- and 96-well plates. **(A)** At day 4 after transduction, cells were harvested and analysed for EpCAM mRNA (left; pMX set at 1) and protein (right; pMX set at 100) expression (mean  $\pm$  s.e.m.,  $n \geq 5$ ;  $t$ -test  $**P < 0.01$ ,  $***P < 0.001$ ). **(B)** At day 4 after transduction, cells were fixed, stained (inset) and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay was performed at days 4, 5 and 6 after transduction (left) or cells were replated at equal cell numbers and MTT assay was performed for the four following days (right). GAPDH, glyceraldehyde 3-phosphate dehydrogenase; MFI, mean fluorescence intensity; OD, optical density.

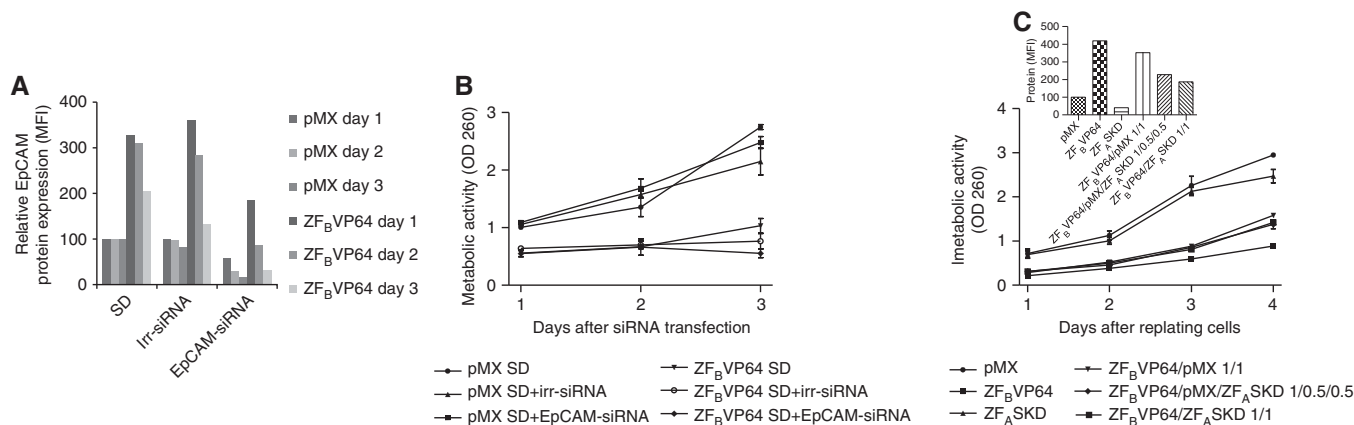


**Figure 3.** SiRNA-based silencing of EpCAM expression in ovarian cancer has no effect on cell growth. SKOV3 cells were transfected with irr-siRNA or EpCAM-siRNA. At days 1 to 3 after transfection, EpCAM mRNA, protein expression **(A)** and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay **(B)** were analysed (SD set at 100 = delivery agent). MFI, mean fluorescence intensity; OD, optical density.

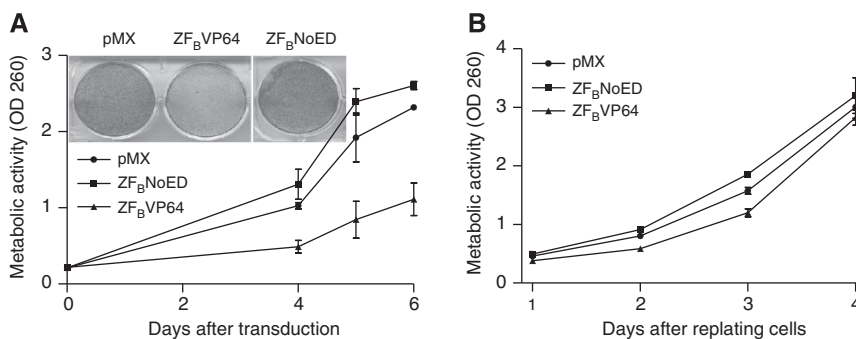
mixed with pMX and or ZF<sub>A</sub>SKD. Although the pMX-ATF mixtures induced a gradual decrease of EpCAM upregulation (Figure 4C, inset), the lowest induced upregulation was still 1.9-fold and only a slight recovery from growth inhibition was observed (Figure 4C). Subsequent induction of different levels of endogenous EpCAM expression (e.g. by titration of pMX-6-ZF-B-VP64 with ZF<sub>A</sub>SKD) confirmed that exceeding a threshold of EpCAM expression resulted in irreversible growth inhibition (Supplementary Figure 3).

**Induction of EpCAM expression in EpCAM-negative ovarian cancer cell line.** To further validate our unexpected finding that up-regulation of EpCAM expression resulted in growth inhibition

in ovarian cancer, we investigated the phenotypical changes due to induction of EpCAM in the EpCAM-negative ovarian cancer cell line A2780. Indeed, ZF<sub>B</sub>VP64-transduced A2780 cells clearly displayed a lower number of surviving cells than pMX-transduced cells and again growth inhibition was observed (Figure 5A). Transfection of an EpCAM cDNA-expressing plasmid also induced growth inhibition (Supplementary Figure 2). When surviving infected cells were harvested at day 4 and replated at equal densities, the same growth rate was obtained for cells transfected with pMX, ZF<sub>B</sub>NoED or ZF<sub>B</sub>VP64 (Figure 5B). The difference in growth capacity of direct treated cells (Figure 5A) compared with the replated surviving cell population (Figure 5B) can be explained by the cytotoxic effect of the induced EpCAM expression. At day 4,



**Figure 4.** Increased EpCAM expression levels correlate with cell growth inhibition. SKOV3 cells were transduced with pMX and ZF<sub>B</sub>VP64. At day 4 after transduction, cells were equally replated. Next day, siRNA transfection was performed, and subsequently at 3 days, EpCAM protein expression (A) and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay (B) were analysed. (C) SKOV3 cells were transduced with a constant amount of ZF<sub>B</sub>VP64 virus supernatant mixed with pMX and/or ZF<sub>A</sub>SKD. At day 4 after transduction, EpCAM protein expression was measured (inset) and transduced cells were replated at equal cell numbers, and MTT assay was performed for the four following days.



**Figure 5.** Induction of EpCAM expression in EpCAM-negative ovarian cancer cell line. A2780 cells were transduced to express the indicated ATFs in 6- and 96-wellsplates. (A) At day 4, transduced cells were fixed and stained, or 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay was performed at days 4, 5 and 6 after transduction. (B) Transduced cells were harvested at day 4 after transduction, replated at equal cell numbers in 96-well plates and MTT assay was performed for the four following days. OD, optical density.

only  $35 \pm 3\%$  of the A2780 cells was still GFP-positive compared with  $85 \pm 2\%$  and  $80 \pm 10\%$  for ZF<sub>B</sub>NoED and ZF<sub>A</sub>SKD, respectively. In contrast, ZF<sub>B</sub>VP64-transduced SKOV3 cells displayed at day 4 after transduction  $84 \pm 5\%$  GFP-positive cells compared with  $96 \pm 1\%$  and  $93 \pm 6\%$  for ZF<sub>B</sub>NoED and ZF<sub>A</sub>SKD, respectively.

## DISCUSSION

Dependent on the tumour type, EpCAM overexpression has been associated with either decreased or increased overall survival of patients (van der Gun *et al*, 2010). Although this dual role is also reflected by EpCAM modulation studies, for human ovarian cancer no such siRNA/cDNA studies have been reported and the biological role of EpCAM here remains unclear. This study is the first to report that downregulation of EpCAM expression in human ovarian cancer does not decrease growth rate, not when using a repressive ATF nor after siRNA-mediated downregulation. Importantly, ATF-induced EpCAM downregulation did result in cell growth inhibition for breast cancer, confirming previous RNA interference studies for this carcinoma type (Osta *et al*, 2004; Sankpal *et al*, 2011).

As our findings suggest the absence of an oncogenic role for EpCAM in human ovarian cancer, we set out to induce or further

upregulate EpCAM expression in EpCAM-negative or low expressing ovarian cancer cells. As has also been reported for renal cell (Klatte *et al*, 2009) and thyroid carcinomas (Ralhan *et al*, 2010), we observed an inhibition in cell growth by inducing EpCAM expression. As ablation of p53 expression has been associated with an increase in EpCAM expression (Sankpal *et al*, 2009), the commonly observed EpCAM overexpression in ovarian cancer might be a consequence of p53 dysfunction, ubiquitously observed for ovarian cancers (Ahmed *et al*, 2010). Indeed, although EpCAM expression enhanced the tumour-initiating ability of ovarian stem-like cells, a significant outgrowth of tumour mass was only observed for EpCAM-positive cells when p53 was down-regulated (Motohara *et al*, 2011). In addition, a growth-promoting role of EpCAM via Wnt signalling, an important pathway in cancers, might not be of general importance in the development of cancer. Whereas EpCAM overexpression in MDA-MB231 could activate Wnt signalling, this was not observed in Hs578T (Gostner *et al*, 2011). Alternatively, the high EpCAM expression commonly observed on ovarian cancer cells could be explained by a recent postulation stating that high-grade serous ovarian cancer arises from the fallopian tube instead of the ovary (Reitsma *et al*, 2012). The relative increase of EpCAM protein expression in ovarian cancer (2.8 on a scale of 0–3) compared with healthy fallopian tube epithelium (1.8) is only 1.5-fold, whereas the increase of EpCAM

protein expression compared with ovarian surface epithelium (0.2) is 14-fold (Emmanuel *et al*, 2011).

As EpCAM is frequently overexpressed in ovarian cancer, anti-EpCAM antibodies are successfully exploited to recruit immune effector cells, resulting in ovarian cancer cell lysis (Richter *et al*, 2010). Despite this important role of EpCAM in ovarian cancer therapy, we show here that inhibition of EpCAM expression does not offer a promising therapeutic approach for ovarian cancer.

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