NEWS AND COMMENTARY

Heredity (2007) 98, 61–62 © 2007 Nature Publishing Group All rights reserved 0018-067X/07 \$30.00

www.nature.com/hdy

Conserved gene order belies rapid genome turnover

The dynamic interplay between genomic DNA and the outside world

AR Leitch

Heredity (2007) **98,** 61–62. doi:10.1038/sj.hdy.6800924; published online 20 December 2006

here is an increasing understanding of the extraordinary scale of the dynamic change occurring in the eukaryote genome. Part of this dynamism involves retroelement reorganization and replacement. In a recent paper published in Nature, Tarlinton et al. (2006) suggest that a retrovirus has integrated into the koala genome remarkably recently - within the last 100 years - and is now part of the endogenous retrovirus populations in some koalas. Their data is contributing to a growing realization that a model of intragenomic dynamism must be extended to the outside world: to extragenomic environments.

When chromosomes were first discovered, they were considered as stable units of inheritance, whose internal structure changed little over long time periods. Of course it was noted that gross chromosomal changes did occur (e.g. translocations, deletions, inversions), but these changes were thought to involve large chunks of chromatin moving en block (Darlington, 1939). This view persisted into recent times with the arrival of the field of synteny, the study of conserved gene order in the genomes of species with distant common ancestors (Devos and Gale, 1997). But these data belie a dynamic mobility in the DNA sequences within the chromosome that is not apparent from traditional cytogenetics or by comparisons of the marker sequences used in synteny studies.

The realization that DNA can jump within the genome began with McClintock's discovery of transposons in Zea mays (maize). But the extent to which mobile elements influence the eukaryote genome is only now being realized. Much of the eukaryote genome is composed of retroelements, for example 42% of the human (Lander et al., 2001) and over 70% of some plant genomes (Kumar and Bennetzen, 1999). Čut-andpaste DNA transposons constitute another 3% of the human genome and recent in silico analysis of genomic DNA sequence from species as diverse as Arabidopsis and Caenorhabdites indicate that up to 2% is composed of Helitrons (Kapitonov and Jurka, 2001), which are elements that multiply using rolling circle replication. As was the case for other transposons, evidence for helitron mobility is emerging in *Z. mays* (Lal and Hannah, 2005). If we include tandemly repeated DNA (e.g. satellite DNA and nuclear ribosomal DNA), then a model of the eukaryotic chromosome emerges that is comprised almost entirely of repetitive and transposable elements.

Comparative genomic studies are revealing that the abundance and distribution of repetitive sequences can be highly labile. Retrotransposon mobility can lead to copy number amplifications, while excisions, perhaps mediated by illegitimate recombination, act to reduce copy numbers (Devos et al., 2002). The consequence can be a fast turnover of elements, estimated to be within 8 million years, with an element half-life of less than 6 million years (Ma et al., 2004). Studies on tandem repeated satellite sequences also reveal rapid change, with amplifications and reductions, probably mediated by recombination-based homogenization mechanisms, replacing units with variants over time-frames of 3 million years or less (Lim et al., 2006).

The realization of these high levels of genomic dynamism led Brookfield (2005) to consider the genome as an ecological community of elements. This analogy is useful since it can be extended to include the invasion of new sequences (species in his analogy) from the outside world. Sequence analysis and phylogenetic reconstructions point to the occurrence of horizontal gene transfer (HGT) or lateral gene transfer (LGT) between species, involving the exchange of DNA between viruses, prokaryotes and eukaryotes. Although the extent to which HGT/LGT is occurring (Keeling et al., 2005) remains controversial. Nevertheless there are examples of well-characterized horizontally transferred material becoming fixed in eukaryote genomes. Three examples are considered here because

they relate to the integration of viral sequences to eukaryote genomes. In some species of Nicotiana, geminivirus related DNA (GRD) is found as tandem repeats. It is thought that these sequences are derived from two independent integration events of geminivirus DNA (Murad et al., 2004). Geminivirus genomes, like Helitrons, amplify using rolling circle replication and some GRD sequences show sequence identity with Helitrons. It is possible that a mobile Helitron captured geminivirus DNA and a recombinant sequence integrated and amplified in the genome. Integrated pararetroviruses sequences (banana streak virus, tobacco vein clearing virus and petunia vein clearing virus) are also known in plant genomes. They are not simply relics of past events, but can actually be released from the genome under certain conditions to form infectious particles (Lheureux et al., 2003). Similarly, a clear relationship between retroviruses and LTR retrotransposons has been recognized. The independent acquisition of env-like genes into some retrotransposons lineages appears to have occurred independently several times, and these events may have helped mediate transmission between species and, in some instances, infectious ability (Malik et al., 2000). Thus, there are examples of two-way traffic into and out of the genome, extending the flow of genetic information between the genome and the free-living, extra-genomic world.

In the paper by Tarlinton et al. (2006), the authors describe the integration of a koala retrovirus (KoRV) into the koala genome. They suggest that this event is sufficiently recent that not all koalas carry the sequence. Indeed, by studying populations of koalas, including those established on Kangaroo Island off mainland Australia in the early 1900s, they propose sequence integration probably begun within the last 100 years. If so they have identified a sequence right at the beginning of its evolution from a free-living koala retrovirus (KoRV) to an endogenous sequence. The finding promises an important model to study early retrovirus/retroelement evolutionary dynamics and insights into the exchange of DNA between genomic DNA and the outside world.

AR Leitch is at the School of Biological and Chemical Sciences, Queen Mary University of London, London, UK.

e-mail: A.R.Leitch@qmul.ac.uk

Brookfield JFY (2005). The ecology of the genome – mobile DNA elements and their hosts. *Nat Rev Genet* 6: 128–136.

Darlington CD (1939). Evolution of Genetic Systems. Oliver and Boyd: Edinburgh.

- Devos KM, Gale MD (1997). Comparative genetics in the grasses. *Plant MolBiol* **35**: 3–15.
- Devos KM, Brown JKM, Bennetzen JL (2002). Genome size reduction through illegitimate recombination counteracts genome expansion in Arabidopsis. *Genome Res* 12: 1075–1079.
- Kapitonov VV, Jurka J (2001). Rolling-circle transposons in eukaryotes. Proc Natl Acad Sci USA 98: 8714–8719.
- Keeling PJ, Burger G, Durnford DG, Lang BF, Lee RW, Pearlman RE *et al.* (2005). The tree of eukaryotes. *Trends Ecol Evol* **20**: 670–676.
- Kumar A, Bennetzen JL (1999). Plant retrotransposons. Annu Rev Genet **33**: 479–532.
- Lal SK, Hannah LC (2005). Plant genomes massive changes of the maize genome are caused by Helitrons. *Heredity* **95**: 421-422.
- Lander ES, Linton LM, Birren B, Nusbaum C, Zody MC, Baldwin J et al. (2001). Initial

sequencing and analysis of the human genome. *Nature* **409**: 860–921.

- Lheureux F, Carreel F, Jenny C, Lockhart BEL, Iskra-Caruana ML (2003). Identification of genetic markers linked to banana streak disease expression in inter-specific Musa hybrids. *Theor Appl Genet* **106**: 594–598.
- Lim KY, Kovarik A, Matyasek R, Chase MW, Knapp S, McCarthy E et al. (2006). Comparative genomics and repetitive sequence divergnece in the species of diploid Nicotiana section Alatae. Plant J 48: 907–919.
- Ma JX, Devos KM, Bennetzen JL (2004). Analyses of LTR-retrotransposon structures reveal recent and rapid genomic DNA loss in rice. *Genome Res* 14: 860–869.
- Malik HS, Henikoff S, Eickbush TH (2000). Poised for contagion: Evolutionary origins of the infectious abilities of invertebrate retroviruses. *Genome Res* **10**: 1307–1318.
- Murad L, Bielawski JP, Matyasek R, Kovarik A, Nichols RA, Leitch AR *et al.* (2004). The origin

and evolution of geminivirus-related DNA sequences in *Nicotiana*. *Heredity* **92**: 352–358.

Tarlinton RE, Meers J, Young PR (2006). Retroviral invasion of the koala genome. *Nature* **442**: 79–81.

Editor's suggested reading

- Belshaw R, Bensasson D (2006). The rise and falls of introns. *Heredity* **96**: 208–213.
- Buchon N, Vaury C (2005). RNAi: a defensive RNA-silencing against viruses and transposable elements. *Heredity* 96: 195–202.
- Filatov D (2005). Evolutionary genetics: Stickleback's view of sex chromosome evolution. *Heredity* **94**: 275–276.
- Schulman AH, Sabot F (2006). Parasitism and the retrotransposon life cycle in plants: a hitchhiker's guide to the genome. *Heredity* doi: 10.1038/sj.hdy.6800903.
- Volff JN (2005). Genome evolution and biodiversity in teleost fish. *Heredity* **94**: 280–294.