Short Communication

Germline *TP53* alterations in Finnish breast cancer families are rare and occur at conserved mutation-prone sites

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Summary We have screened for germline *TP53* mutations in Finnish *BRCA1* and *BRCA2* mutation-negative families. This study represents the largest survey of the entire protein-encoding portion of *TP53*, and indicates that mutations are only found at conserved domains in breast cancer families also meeting the criteria for Li-Fraumeni/Li-Fraumeni-like syndrome, explaining only a very small additional fraction of the hereditary breast cancer cases. © 2001 Cancer Research Campaign http://www.bjcancer.com

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Approximately 5–10% of breast cancer patients have some degree of family history and may be carriers of an inherited susceptibility to the disease (Claus et al, 1996). However, mutations in the two major breast cancer susceptibility genes, *BRCA1* and *BRCA2* (Miki et al, 1994; Wooster et al, 1995), have been found only in about 20% of Finnish high-risk breast cancer families (Vehmanen et al, 1997a, b; Huusko et al, 1998). Very recently, the chromosomal location of a new susceptibility gene was identified (Kainu et al, 2000). This and other genes, such as *TP53*, *PTEN*, and possibly *ATM*, are responsible for an additional fraction of breast cancer predisposition (reviewed in Easton, 1999).

Somatic mutations in the *TP53* gene are found in almost all kinds of tumours (Hollstein et al, 1991). Mutations are usually clustered within the most conserved regions of exons 4, 5, 7 and 8, corresponding to the sequence-specific DNA binding domain of the protein (Levine, 1997). Germline *TP53* mutations have been found in patients with the rare Li-Fraumeni syndrome (LFS), characterized by breast cancer, osteosarcoma, leukaemia, brain and adrenocortical tumours at an early age (Birch et al, 1994; Varley et al, 1997b; reviewed in Eng et al, 1997 and Varley et al, 1997a). A similar cancer phenotype has recently been observed in patients carrying hereditary mutations in the checkpoint kinase gene, *CHK2*, indicating genetic heterogeneity within LFS (Bell et al, 1999).

In the previous study by Huusko et al (1999), we screened 7 Finnish LFS and Li-Fraumeni-like (LFL) families for *TP53* exon 5–8 mutations. Two changes were identified (Tyr220Cys and Asn235Ser), both of which appeared to associate with accumulation of female breast cancer. Based on these results, we wanted to see whether germline *TP53* mutations could also explain familial breast cancer cases without LFS/LFL phenotype and if mutations

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could be found outside the conserved segment. We also anticipated finding geographical clustering, because of the strong founder effects regarding *BRCA1* and *BRCA2* mutations in Finland (Huusko et al, 1998; Sarantaus et al, 2000). Here, a large cohort of 130 breast cancer patients from 108 Finnish *BRCA1* and *BRCA2* mutation-negative breast cancer families were screened for *TP53* germline alterations covering the entire protein encoding region (exons 2–11) of the gene.

We analysed 130 subjects from 108 Finnish BRCA1 and BRCA2 mutation-negative breast cancer families (Vehmanen et al, 1997a, b; Huusko et al, 1998). Of these families, 5 had previously been studied for TP53 exon 5-8 mutations (Huusko et al, 1999). Geographically, the families originated from 3 regions of Finland: 79 from the Oulu University Hospital area, 13 from the Tampere University Hospital area and 16 from the Helsinki University Central Hospital area. The criteria for inclusion were 3 or more cases of breast cancer in first- or second-degree relatives, early disease onset (≤35 years), bilateral breast cancer, or multiple tumours including breast cancer in the same individual. 75 families met the criteria for moderate- to high-risk hereditary breast cancer only, 32 for both hereditary breast cancer and LFL, and 1 for both hereditary breast cancer and LFS (Table 1; Birch et al, 1994; Eng et al, 1997). To search for founder effects, additional BRCA1 and BRCA2 mutation-negative breast cancer families (50 for Tyr220Cys and Asn235Ser, and 5 for Arg248Gln), originating from the same geographical regions as those with the identified mutations, were analysed. In addition, DNAs from 500 unselected consecutive breast cancer patients from the Tampere region were used to establish the mutation frequency in the population. Informed consent to obtain pedigree data and a blood specimen for the study was obtained from all patients. The Ethical Boards of the participating hospitals and the Finnish Ministry of Social Affairs and Health have approved the study.

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Table 1 Summary classification of all studied 108 cancer families according to occurrence of breast, other LFS/LFL-associated (osteosarcoma, leukaemia, brain or adenocortical tumours) and other cancers in 1st and 2nd degree relatives

No. of breast cancer cases within a family			canc	of of ers w fami	No. of LFS/LFL- associated cancers within a family			
	0	1	2	3	4	5	6	_
1	1	2 1 1	1	2		1		0 1 2 3 4
2	6 2	9	9	1	1		1	0 1 2 3 4
3	5 1 1	5 3	10 4	1 2	2 1	1	3	0 1 2 3 4
4	1	1 2	3 1 1	1	1		1	0 1 2 3 4
5	1	2	1		1		1	0 1 2 3 4
≥6	1			1	1			0 1 2 3 4
		To	otal no	o. of f	amili	es: 1	08	

Numbers in bold indicate the total number of families in each class. Of the 130 breast cancer cases, 12 (9%) were identified at or below age 35, 86 (66%) between ages 36-60, and 32 (25%) at or above age 61.

Using genomic DNA, TP53 exons 2-11 were screened by conformation sensitive gel electrophoresis (CSGE) or fluorescense-CSGE as described earlier (Ganguly et al, 1998; Huusko et al, 1998). Samples with a band shift were sequenced as instructed by the apparatus manufacturer Li-Cor (Lincoln, USA). DNAs from 500 unselected consecutive breast cancer cases were screened using DNA minisequencing (Syvänen, 1999). Oligonucleotide sequences were designed based on GenBank sequence information (U94788).

In the present study, 108 breast cancer families were screened for TP53 mutations. This is a considerably larger number of families fulfilling the criteria of hereditary breast cancer than in any previous study. The CSGE and F-CSGE analyses of exons 2-11 revealed a constitutional TP53 alteration, Arg248Gln, in one of the studied families. In addition, one silent variant (Arg213Arg) and other known polymorphisms in the intronic region between the exons 2 and 3, as well as in exon 4, were detected (data not shown).

The subject with the Arg248Gln ($CGG \rightarrow CAG$) mutation showed strong family history of breast cancer. The cancer spectrum of this family (#6001, Figure 1A) also matches the LFL criteria (Birch et al, 1994; Eng et al, 1997). Immunohistochemical analysis results from patient records revealed positive p53 staining in the tumours of all three of the mutation carriers.

Arg248 resides in the highly conserved region of exon 7 and is the most frequently altered residue of the p53 protein, being essential for DNA binding functions (Cho et al, 1994). Therefore, the Arg248Gln substitution leads to defective contacts with target DNA. Agr248Gln has most frequently been detected in colorectal and breast tumours (Lasky and Silbergeld, 1996). It is also the most commonly found alteration in LFS (Shibata et al, 1996). In the family carrying Arg248Gln (Figure 1A), we were unable to determine the mutational status of the proband's (case 19) paternal grandparents. Since they both died at an old age without evidence of cancer, it is possible that this is a de novo mutation.

Screening of the Arg248Gln mutation and the previously detected two Finnish germline TP53 mutations (Tyr220Cys, Asn235Ser; Huusko et al, 1999) was expanded to include 500 unselected consecutive breast cancer cases. No mutations were found, suggesting that the frequency of these mutations is very low in the general breast cancer population. Our results support the observation that TP53 alterations are rare and explain only a negligible fraction of breast cancer cases at the population level, mainly associated with strong family history of breast cancer. There was no evidence of founder effects that otherwise are common in hereditary diseases in Finland. This may be due to the rareness/young age/more severe effect on survival of the studied TP53 mutations, as compared to the situation in for instance HNPCC- (Nyström-Lahti et al, 1995) and BRCA1/BRCA2-related cancer predisposition (Huusko et al, 1998; Sarantaus et al, 2000). Evidence against founder effects was also obtained by studying additional breast cancer cases originating from the same geographical areas as those with mutations.

Taking together the two studies, we have analysed the entire TP53 protein-encoding region in 108 breast cancer families and found mutations in three (2.8%) of them (Figure 1). This frequency is in agreement with the concept that mutations preferentially occur in LFS/LFL-related patients. Zeleda-Hedman et al (1997), Warren et al (1992), Prosser et al (1991) and Patel et al (1995) studied 109, 25, 5 and 4 breast cancer families, respectively, without finding any mutations. Boerresen et al (1992) studied 237 women with breast cancer, of which 30 had at least one first-degree relative with breast cancer, 40 had breast cancer before age 35, and 167 represented unselected breast cancer patients. Only one unselected and one early-onset patient were found to carry a mutation. On more detailed review, both of these cases had a family history of breast cancer and other malignancies suggestive of LFS. Lidereau and Soussi (1992) studied 19 unrelated cases with bilateral breast cancer, but no TP53 mutations were detected. Breast and other cancers had occurred in the relatives of 7 of the 12. Sidransky et al (1992) found 1 of 126 patients with early-onset breast cancer carrying a germline change. These earlier mutation analyses have usually been limited to exons 5–8. Due to recent observations that the mutations could also reside in domains responsible for transcription and oligomerization control (exons 2-4 and 9-11) (Varley et al, 1997a), we decided to investigate these regions too. However, our results support the existing view (Toguchida et al, 1992; Birch et al, 1994; Cornelis et al, 1997; Varley et al, 1997b), suggesting that the breast cancer

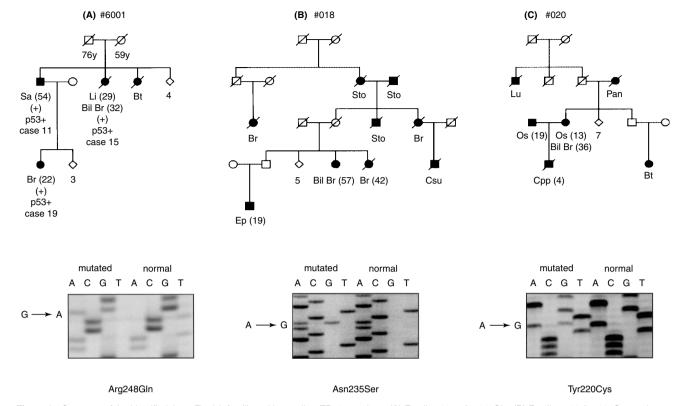


Figure 1 Summary of the identified three Finnish families with germline *TP53* mutations: (**A**) Family #6001 Arg248Gln, (**B**) Family #018 Asn235Ser, and (**C**) Family #020 Tyr220Cys. Tumours: Bil Br, bilateral breast; Br, breast; Bt, brain; Cpp, choroid plexus papilloma; Csu, cancer site unknown; Ep, ependyoma; Li, liver; Lu, lung; Os, osteosarcoma; Pan, pancreas; Sa, sarcoma; Sto, stomach. The age at diagnosis, when known, is marked after the malignancy. The number of siblings is shown below the diamonds. For the new family #6001, (+) = mutation carrier, p53+ = positive staining in immunohistochemical analysis. The case numbers of the individuals analysed are shown below the carrier status

predisposing *TP53* mutations mainly occur at specific mutation-prone regions (exons 6–7) of the conserved parts of the gene.

All 3 Finnish *TP53* germline mutations affect mutation-prone sites and have been observed previously: Tyr220Cys and Arg248Gln in LFS/LFL families (Birch et al, 1994; Varley et al, 1997b), and Asn235Ser in LFS-associated malignancies, but with an uncertain familial background of cancer predisposition (Wagner et al, 1994; Diller et al, 1995; Cornelis et al, 1997). Available data may therefore suggest that Asn235Ser could be associated with variable cancer susceptibility and reduced penetrance. Results from both our (Figure 1) and other studies indicate that these mutations may be associated in particular with female breast cancer, frequently bilateral, and young age of onset, in combination with other signatures of LFS or LFL.

In conclusion, this systematic, large-scale study of germline *TP53* alterations among breast cancer families indicates that mutations in the conserved regions of the gene seem to explain only a very small fraction of the Finnish *BRCA1* and *BRCA2* mutation-negative breast cancer cases, and that additional and more important hereditary breast cancer susceptibility genes remain to be identified.

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