

Letter to the Editor

Regarding: Koilocytes indicate a role for human papilloma virus in breast cancer

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Sir,

I welcome the contributions of Heng et al and Lawson et al in the October 2009 issue of the British Journal of Cancer. I would like to comment on the article submitted by Lawson et al, 'Koilocytes indicate a role for human papilloma virus in breast cancer'. The authors cite an earlier article in which I was a participating author and refer to the koilocytes identified in our article as 'putative (supposed) HPV-associated koilocytes'. The concept of koilocytosis was initially and arguably still remains a morphological construct (Reid et al, 1982). In the early 1990s, based on a systematic and careful review of routine breast cancer specimens, which included areolar and nipple tissue, Buck and I submitted, for publication, a select series of breast cancer cases with koilocytosis in the nipple and areola. The initial attempts at publication were rejected because of an absence of molecular confirmation of the presence of human papilloma virus (HPV). Subsequently, with de Villiers (Buck et al, 2001), we reported a limited series of areola and nipple specimens with histological evidence of koilocytosis confirmed by polymerase chain reaction (PCR). In a larger series, joined by zur Hausen (de Villiers et al, 2005), we published a series of cases including both nipple and areolar specimens and associated breast cancer tissue. In that report, both PCR and in situ hybridization were used to confirm the presence of the viral DNA in koilocytes. We demonstrated the co-localization by in situ hybridization with histological evidence of koilocytosis. Our confirmatory methods were in essence identical to those used by Lawson et al.

Recent identification of HPV by PCR in breast milk, ductal lavage and colostrum specimens is not cited by Lawson et al (Sarkola et al,

2008; Cazzaniga et al, 2009). Human papilloma virus was detected in 4 and 14% of non-selected breast milk specimens. Although the authors identified low numbers of positive HPV samples, the timespecific incidence of HPV positivity is of the same order of magnitude as encountered in cervical specimens of sexually active women and, to my mind, implies a relatively high aggregate lifetime likelihood of HPV colonization of the nipple and/or areola. Cazzaniga et al have argued that scraping the superficial layers of the nipple decreased the number of positive samples. However, the argument can be made that this may reflect the natural history of HPV in that virus accumulates through the course of keratinizing cell maturation from the basal layer to the superficial layer. It has been documented that sampling from the surface of skin tumours after striping the biopsy with an adhesive tape decreases the likelihood of isolating the virus by PCR (Forslund et al, 2004). The presence of the virus in breast milk specimens supports the likelihood that HPV infects the nipple and areolar tissue and indirectly supports the concept of HPV-specific koilocytosis at these sites. I hope I will be allowed some latitude for speculation in this regard. There is the possibility that HPV transmission in the neonatal period in breast milk or colostrum through immune modulation and oral induced tolerance (Strobe et al, 2001; Lawrence and Lawrence, 2004) may modify host immune response to the virus encountered later in life. This speculative construct may further complicate and enrich the pathophysiological mechanism that Buck et al initially postulated that HPV infection as evidenced by koilocytosis affecting the nipple, areola and duct structures may be related to the induction of neoplasia in breast tissue by retroductular spread of the virus.

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