

Hepatic cytochrome P450 3A drug metabolism is reduced in cancer patients who have an acute-phase response

LP Rivory^{*,1,2}, KA Slaviero² and SJ Clarke¹

¹Medical Oncology, Level 6 Gloucester House, Sydney Cancer Centre, Missenden Road, Camperdown, NSW 2050, Australia; ²Department of Pharmacology, University of Sydney, 2006, NSW, Australia

Inflammatory disease states (infection, arthritis) are associated with reduced drug oxidation by the cytochrome P450 3A system. Many chemotherapy agents are metabolised through this pathway, and disease may therefore influence inter-individual differences in drug pharmacokinetics. The purpose of this study was to assess cytochrome P450 3A function in patients with advanced cancer, and its relation to the acute-phase response. We evaluated hepatic cytochrome P450 3A function in 40 patients with advanced cancer using the erythromycin breath test. Both the traditional $C_{20\text{min}}$ measure and the recently proposed $1/T_{\text{MAX}}$ values were estimated. The marker of acute-phase response, C-reactive protein and the pro-inflammatory cytokines IL-6, IL-1 β , TNF α and IL-8 were measured in serum or plasma at baseline. Cancer patients with an acute phase response (C-reactive protein > 10 mg l⁻¹, $n=26$) had reduced metabolism as measured with the erythromycin breath test $1/T_{\text{MAX}}$ (Kruskal–Wallis Anova, $P=0.0062$) as compared to controls (C-reactive protein ≤ 10 mg l⁻¹, $n=14$). Indeed, metabolism was significantly associated with C-reactive protein over the whole concentration range of this acute-phase marker ($r=-0.64$, Spearman Rank Correlation, $P<0.00001$). C-reactive protein serum levels were significantly correlated with those of IL-6 (Spearman coefficient=0.58, $P<0.0003$). The reduction in cytochrome P450 3A function with acute-phase reaction was independent of the tumour type and C-reactive protein elevation was associated with poor performance status. This indicates that the sub-group of cancer patients with significant acute-phase response have compromised drug metabolism, which may have implications for the safety of chemotherapy in this population.

British Journal of Cancer (2002) 87, 277–280. doi:10.1038/sj.bjc.6600448 www.bjcancer.com

© 2002 Cancer Research UK

Keywords: acute phase response; drug metabolism; erythromycin breath-test (EBT)

Viral and bacterial infection, severe trauma and degenerative diseases are known to cause significant reductions in hepatic drug clearance, mostly through decreased expression of drug-metabolising cytochrome P450 enzymes (Morgan, 1997). This is mediated largely through down-regulation of gene transcription by the pro-inflammatory cytokines such as IL-6 and TNF α (Muntane-Relat *et al*, 1995; Morgan, 1997; Guillen *et al*, 1998; Pascussi *et al*, 2000). These cytokines also induce the synthesis of acute-phase reactants, such as C-reactive protein, by the liver (O’Riordain *et al*, 1999). This is usually accompanied by a decreased synthesis of albumin and pre-albumin. Most commonly, reduction in drug metabolism in the presence of acute-phase reactants involves the cytochrome P450 3A family (CYP3A), which is responsible for the metabolism of ~60% of drugs used in medicine. This includes many of those used in cancer chemotherapy (e.g., taxanes, vinca alkaloids, camptothecins, tamoxifen, etoposide and oxazaphosphorines (Kivisto *et al*, 1995)).

The manner in which patients tolerate chemotherapy in general is remarkably unpredictable, and some experience significant morbidity leading to hospitalisation and, occasionally, mortality. Pro-inflammatory cytokines and acute-phase reactants are elevated

in many patients with advanced cancer (Heys *et al*, 1998; Martin *et al*, 1999) and there is, therefore, the possibility that part of the inter-individual variability in drug clearance and toxicity could relate to the effects of these cytokines on CYP3A expression (Moreno *et al*, 1991; Craig *et al*, 1993; Chen *et al*, 1994; Muntane-Relat *et al*, 1995; Morgan, 1997; Pascussi *et al*, 2000). Further falls in CYP3A function may occur with age (Hunt *et al*, 1990) and reduced hepatic drug clearance may contribute to a greater risk of adverse events in elderly cancer patients (Yancik *et al*, 1998).

Pharmacokinetic variability in the disposition of anticancer drugs is responsible for a significant proportion of inter-individual variability in their activity and toxicity (Gurney, 1996). Surprisingly, the impact on drug metabolism of the acute-phase response that often accompanies cancer has not been explored in this setting. Hence, the purpose of the presented study was to estimate liver CYP3A function in patients with advanced cancer, to examine its association with the acute-phase response and to identify the key cytokines involved in the initiation of the latter. Serum levels of basic fibroblast growth factor (bFGF) and vascular endothelium growth factor (VEGF) have been shown to correlate strongly with tumour stage and outcome in several malignancies (Chen *et al*, 1999; Graeven *et al*, 1999; Ugurel *et al*, 2001). These may predict for more aggressive tumours. Hence, a secondary aim was to examine the relationship between the acute-phase response and circulating levels of bFGF and VEGF.

*Correspondence: LP Rivory; E-mail: lrivory@canc.rpa.cs.nsw.gov.au
Received 9 January 2002; revised 24 April 2002; accepted 12 May 2002

METHODS

Subjects

This was a prospective, single-centre study of the influence of acute-phase response on drug metabolism in cancer patients. It was open to all subjects >18 years of age with biopsy-proven, advanced malignancy who were about to receive chemotherapy. The entry criteria were: ECOG performance status (PS) 0-3, neutrophils $>3.0 \times 10^9 \text{ l}^{-1}$, bilirubin $<2.0 \text{ ULN}$ (upper limit of normal) and transaminases $<2.5 \times \text{ULN}$ unless evidence of liver involvement ($<5 \times \text{ULN}$). The Ethics Committee of the Central Sydney Area Health Service approved the study, and written informed consent was obtained from all subjects.

Experimental protocol

Blood samples were collected for routine evaluation of haematological and biochemistry parameters within 72 h prior to administering the EBT. Serum samples were also collected for the analysis of the acute-phase reactants C-reactive protein (CRP) and α_1 acid glycoprotein (AAG) as well as albumin and pre-albumin and frozen at -70°C until analysis. The latter were performed by the Biochemistry Department of the Royal Prince Alfred Hospital using standard turbidimetric and nephelometry assays. Presence of an acute-phase response was defined as CRP $>10 \text{ mg l}^{-1}$.

The cytokines IL-1 β , IL-6, TNF α and IFN γ were analysed in serum and IL-8, VEGF and bFGF in plasma using commercial ELISA kits (R&D, Minneapolis, MN, USA). Standard curves were run with each batch and only values greater than the lowest standard were reported ($>15.6 \text{ pg ml}^{-1}$ for IFN γ , TNF α , VEGF; $>31.2 \text{ pg ml}^{-1}$ for IL-8; $>3.9 \text{ pg ml}^{-1}$ for IL-1 β , $>1 \text{ pg ml}^{-1}$ for bFGF and $>3.13 \text{ pg ml}^{-1}$ for IL-6).

The erythromycin breath test was performed as recently described (Rivory *et al*, 2000). Briefly, 4 μCi of ^{14}C erythromycin (*N*-methyl- ^{14}C , 55 mCi mmole $^{-1}$, NEN Life Science Products Inc, Boston, MA, USA) was injected intravenously and breath samples were collected into gas-tight balloons (Pytest[®], Ballard Medical Products, Utah, USA) 5, 10, 15, 20, 25, 30 and 40 min later. These were processed by bubbling the collected gas through a capture solution consisting of hyamine hydroxide 10X (Packard, Sydney, NSW, Australia) in 50:50 methanol/ethanol v v $^{-1}$ to which a trace of phenolphthalein was added. After the addition of scintillant (Ultima Gold[®], Packard, Sydney, NSW, Australia) and counting, the data were expressed in terms of per cent of dose exhaled per minute at each time point by assuming a CO $_2$ output of 5 mmoles $\text{min}^{-1} \text{ m}^{-2}$ (Watkins *et al*, 1989). The widely used measure of CYP3A activity, the flux at 20 min ($C_{20\text{min}}$), was recorded (Hirth *et al*, 2000). In addition, the novel parameter, $1/T_{\text{MAX}}$, which correlates with total drug clearance of erythromycin (Rivory *et al*, 2000) was estimated from a fitting of a bi-exponential equation to the data as described recently (Rivory *et al*, 2000, 2001). In some cases, the profiles were extremely flat or had not reached a maximum at 40 min. In these cases, T_{MAX} was set at 50 min.

Statistical analysis

The association between categorical (e.g. gender, ECOG) and continuous variables (e.g. EBT results, cytokine concentrations) was examined by Kruskal-Wallis Anova. Regression analyses between continuous variables were performed with the Spearman rank-order test.

The frequency distributions of CRP and AAG data were evaluated using the Kolmogorov-Smirnov One Sample Test. All tests were carried out using SYSTAT v 7.0.1 (SPSS Inc, Chicago, IL, USA) and $P < 0.05$ was considered as significant.

RESULTS

Between July 2000 and April 2001, a total of 40 subjects were investigated. These patients had mostly lung and breast cancer (see Table 1) and ranged in age from 38 to 83, with a median of 64 years. There were similar numbers of males ($n=21$) and females ($n=19$).

The erythromycin breath test results were found to vary widely in this population. The median (and range in parentheses) for the $C_{20\text{min}}$ measure was $0.050\% \text{ min}^{-1}$ (0.002–0.101) whereas it was 0.050 min^{-1} (0.02–0.12) for $1/T_{\text{MAX}}$. There was no significant effect of age or sex on either of the EBT parameters in this cancer population (Spearman Rank-Order and Kruskal-Wallis Anova, respectively).

Baseline serum CRP, AAG, albumin and pre-albumin were also variable and are summarised in Table 2. Only two patients had quantifiable serum TNF α (16.2, 18.5 pg ml^{-1}), another two had quantifiable IFN γ (37.5, 77.9 pg ml^{-1}), whereas most had quantifiable serum IL-1 β (median: 10.6 pg ml^{-1}). IL-6 was quantifiable in over half ($n=33$) with a median of 5.6 pg ml^{-1} (range: <3.2 –193.5 pg ml^{-1}). VEGF was found to range from <32.1 to 1537 pg ml^{-1} with a median of 274.6 pg ml^{-1} . The range observed for bFGF was $<1 \text{ pg ml}^{-1}$ to 12.2 pg ml^{-1} .

When the cancer patients were divided into control ($\leq 10 \text{ mg l}^{-1}$) and acute-phase response ($>10 \text{ mg l}^{-1}$) groups based on the upper normal limit of serum CRP, those in the acute-phase group had an average 30% reduction in drug metabolism (0.070 ± 0.024 vs $0.049 \pm 0.022 \text{ min}^{-1}$, respectively). This was statistically significant ($P=0.0062$, Kruskal-Wallis Anova).

Further examination revealed that the effect occurred as a continuum with acute-phase response over the entire patient group. Indeed, the EBT $1/T_{\text{MAX}}$ values negatively correlated with both CRP and AAG with Spearman coefficients of -0.64 ($P < 0.00001$) and -0.45 ($P < 0.005$), respectively. Weaker correla-

Table 1 Patients' demographics ($n=40$)

Characteristic	n
Histology	
Lung ^a	19
Breast	9
Carcinoma of unknown primary	3
Head and neck	3
Others	6
ECOG performance status	
0	4
1	21
2	12
3	3
Age (year)	
30–50	7
51–65	14
66–90	19

^aMostly non-small cell lung cancer.

Table 2 Base-line serum protein levels in study population ($n=40$)

Protein	Median	Range	Reference range	Units
Albumin	38.0	18–44	40–50	g l^{-1}
Pre-albumin	0.22	0.05–0.39	0.17–0.35	g l^{-1}
C-reactive protein	13.0	<1 –291	<10	mg l^{-1}
α_1 acid glycoprotein	1.23	0.5–3.18	0.5–1.0	g l^{-1}

tions were observed against albumin and pre-albumin (data not shown). The EBT $C_{20\min}$ was only correlated with pre-albumin (Spearman coefficient=0.38, $P<0.02$). The distribution of the CRP values appeared to be log-normal and log-CRP was significantly correlated with $1/T_{\text{MAX}}$ as the independent variable ($r^2=0.44$, $P<0.00002$, Figure 1). In comparison, a similar regression with the $C_{20\min}$ of the EBT yielded $r^2=0.15$ and $P=0.012$. CRP serum levels were significantly correlated with those of IL-6 (Spearman coefficient=0.58, $P<0.0003$) but not with any of the other cytokines. Also, the CRP levels were significantly different across the ECOG performance status categories (Kruskal–Wallis, $P<0.006$, Figure 2). Because of the heterogeneous nature of the population in terms of disease site, the correlation between CRP and $1/T_{\text{MAX}}$ was also examined in the sub-groups of breast and lung cancer patients. The Spearman correlation values were -0.63 ($P=0.07$, $n=9$) and -0.53 ($P<0.02$, $n=19$), respectively, indicating that the effect is not likely to be tumour-type specific.

Co-medication may affect CYP3A activity either by induction or inhibition. Examination of the treatment files of the patients in this study revealed that three were being treated with inhibitors of CYP3A (diltiazem and clarithromycin, respectively), while four

were on long-term treatment with the inducer dexamethasone (daily doses 2–4 mg). The mean \pm s.d. of $1/T_{\text{MAX}}$ for this latter group was $0.073 \pm 0.028 \text{ min}^{-1}$ as compared to the study average of $0.057 \pm 0.024 \text{ min}^{-1}$. The correlation between $1/T_{\text{MAX}}$ and CRP remained significant even after removal of the data from the seven patients on CYP3A-modifying medication (Spearman $Rho=-0.55$, $P=0.002$).

DISCUSSION

These results indicate that CYP3A function in patients with advanced cancer is highly variable and correlates with markers of the acute-phase response. Those patients with an acute-phase response (CRP $>10 \text{ mg l}^{-1}$) had on average a 30% decrease in their metabolic activity as compared to the control group. This decrease in CYP3A activity with acute-phase response was best detected using the recent $1/T_{\text{MAX}}$ parameter of the erythromycin breath-test, which is a better predictor of total drug clearance (Rivory *et al*, 2001). However, similar trends occurred with the $C_{20\min}$ data, although these were not as significant. One of the disadvantages of using the $C_{20\min}$ approach is that values of this parameter are often significantly different between male and female subjects, possibly because of a flawed assumption regarding CO_2 output (Rivory *et al*, 2001). This phenomenon, however, was not observed with our patient data. This suggests that the extreme variability in CYP3A metabolism observed in cancer patients obscures this possible bias.

The source of the variability in CYP3A function is not known but our observation of a significant correlation between acute-phase response and the EBT $1/T_{\text{MAX}}$ suggests that the pro-inflammatory cytokines, which are increased in malignancy (Heys *et al*, 1998; Barber *et al*, 1999; Martin *et al*, 1999), not only trigger the acute-phase response but also result in compromised drug metabolism by CYP3A in some cancer patients. The strong correlation between the IL-6 and CRP serum levels is in strong agreement with this interpretation, although other cytokines may have contributed. In fact, the biological effect of cytokines is modulated by complex inter-relationships with both their soluble and membrane-bound receptors. We argue that serum CRP, which is an indicator of hepatic gene regulation in the presence of inflammatory cytokines, reflects the overall biological effect of this inflammatory response.

AAG, which is one of the acute-phase reactants, was also increased and there is the possibility that the EBT was modified through the effects of protein-binding. Indeed, erythromycin is highly bound to this protein (Prandota *et al*, 1980). In our study, however, CRP was a more significant predictor of CYP3A activity than AAG. Also, it has been noted that the clearance of hepatically metabolised drugs is sometimes reduced in xenograft-bearing animals, even when these are not bound to AAG (Zamboni *et al*, 1998). Finally, there is evidence in support of a direct effect of pro-inflammatory cytokines on CYP3A expression, activity and drug clearance (Moreno *et al*, 1991; Craig *et al*, 1993; Chen *et al*, 1994; Muntane-Relat *et al*, 1995; Morgan, 1997; Pascucci *et al*, 2000).

The implications of this observation are many and of direct relevance to the chemotherapy of cancer. Firstly, the variability of CYP3A drug metabolism in cancer patients may justify the need for doses to be 'individualised', using measures such as the EBT (Hirth *et al*, 2000). Second, we found an association between acute-phase response and poor performance status in concert with other studies (O'Gorman *et al*, 1999). Hence, the link between acute-phase response and impaired drug metabolism may partly explain the observation of increased toxicity of drugs in patients with poor performance status (Krikorian *et al*, 1978; Freyer *et al*, 2000) although this is very likely a multi-factorial phenomenon.

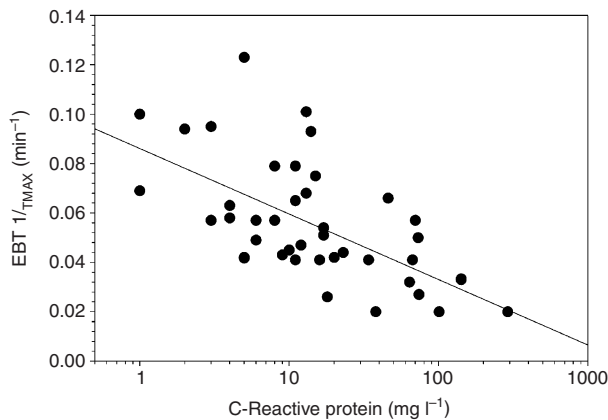


Figure 1 The relationship between the $1/T_{\text{MAX}}$ parameter of the erythromycin breath test and serum C-reactive protein in 40 patients with advanced cancer. The upper limit of normal of CRP is 10 mg l^{-1}

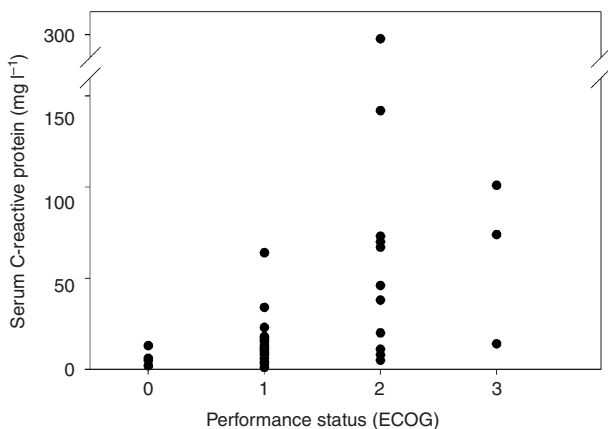


Figure 2 The distribution of the baseline serum C-reactive protein concentration in 40 cancer patients according to their performance status (ECOG). Analysis according to Kruskal–Wallis Anova test indicates significant differences between the four groups ($P<0.006$)

ACKNOWLEDGEMENTS

This study was supported by grants from the New South Wales Cancer Council and the National Health and Medical Research

Council of Australia. We are grateful for the support of the clinical and nursing staff who assisted with this work.

REFERENCES

- Barber MD, Ross JA, Fearon KC (1999) Changes in nutritional, functional, and inflammatory markers in advanced pancreatic cancer. *Nutr Cancer* **35**: 106–110
- Chen YL, Le Vraux V, Leneveu A, Dreyfus F, Stheneur A, Florentin I, De Sousa M, Giroud JP, Flouvat B, Chauvelot-Moachon L (1994) Acute-phase response, interleukin-6, and alteration of cyclosporine pharmacokinetics. *Clin Pharmacol Ther* **55**: 649–660
- Chen Z, Malhotra PS, Thomas GR, Ondrey FG, Duffey DC, Smith CW, Enamorado I, Yeh NT, Kroog GS, Rudy S, McCullagh L, Mousa S, Quezada M, Herscher LL, Van Waes C (1999) Expression of proinflammatory and proangiogenic cytokines in patients with head and neck cancer. *Clin Cancer Res* **5**: 1369–1379
- Craig PI, Tapner M, Farrell GC (1993) Interferon suppresses erythromycin metabolism in rats and human subjects. *Hepatology* **17**: 230–235
- Freyer G, Rougier P, Bugat R, Droz JP, Marty M, Bleiberg H, Mignard D, Awad L, Herait P, Culine S, Trillet-Lenoir V (2000) Prognostic factors for tumour response, progression-free survival and toxicity in metastatic colorectal cancer patients given irinotecan (CPT-11) as second-line chemotherapy after 5FU failure. CPT-11 F205, F220, F221 and V222 study groups. *Br J Cancer* **83**: 431–437
- Graeven U, Andre N, Achilles E, Zornig C, Schmiegel W (1999) Serum levels of vascular endothelial growth factor and basic fibroblast growth factor in patients with soft-tissue sarcoma. *J Cancer Res Clin Oncol* **125**: 577–581
- Guillen MI, Donato MT, Jover R, Castell JV, Fabra R, Trullenque R, Gomez-Lechon MJ (1998) Oncostatin M down-regulates basal and induced cytochromes P450 in human hepatocytes. *J Pharmacol Expl Ther* **285**: 127–134
- Gurney H (1996) Dose calculation of anticancer drugs: a review of the current practice and introduction of an alternative. *J Clin Oncol* **14**: 2590–2611
- Heys SD, Ogston KN, Simpson WG, Walker LG, Hutcheon AW, Sarkar TK, Eremin O (1998) Acute phase proteins in patients with large and locally advanced breast cancer treated with neo-adjuvant chemotherapy: response and survival. *Int J Oncol* **13**: 589–594
- Hirth J, Watkins PB, Strawderman M, Schott A, Bruno R, Baker LH (2000) The effect of an individual's cytochrome CYP3A4 activity on docetaxel clearance. *Clin Cancer Res* **6**: 1255–1258
- Hunt CM, Strater S, Stave GM (1990) Effect of normal aging on the activity of human hepatic cytochrome P450IIE1. *Biochem Pharmacol* **40**: 1666–1669
- Kivisto KT, Kroemer HK, Eichelbaum M (1995) The role of human cytochrome P450 enzymes in the metabolism of anticancer agents: implications for drug interactions. *Br J Clin Pharmacol* **40**: 523–530
- Krikorian JG, Daniels JR, Brown BW, Hu MS (1978) Variables for predicting serious toxicity (vinblastine dose, performance status, and prior therapeutic experience): chemotherapy for metastatic testicular cancer with cis-dichlorodiammineplatinum(II), vinblastine, and bleomycin. *Cancer Treat Rep* **62**: 1455–1463
- Martin F, Santolaria F, Batista N, Milena A, Gonzalez-Reimers E, Brito MJ, Oramas J (1999) Cytokine levels (IL-6 and IFN-gamma), acute phase response and nutritional status as prognostic factors in lung cancer. *Cytokine* **11**: 80–86
- Moreno JJ, Castellote MC, Queralt J (1991) Effect of Mycobacterium butyricum on the hepatic cytochrome P-450 system of the mouse: influence of anti-inflammatory drug. *Comp Biochem Physiol C* **99**: 7–10
- Morgan ET (1997) Regulation of cytochromes P450 during inflammation and infection. *Drug Metab Rev* **29**: 1129–1188
- Muntane-Relat J, Ourlin JC, Domergue J, Maurel P (1995) Differential effects of cytokines on the inducible expression of CYP1A1, CYP1A2, and CYP3A4 in human hepatocytes in primary culture. *Hepatology* **22**: 1143–1153
- O'Gorman P, McMillan DC, McArdle CS (1999) Longitudinal study of weight, appetite, performance status, and inflammation in advanced gastrointestinal cancer. *Nutr Cancer* **35**: 127–129
- O'Riordain MG, Falconer JS, Maingay J, Fearon KC, Ross JA (1999) Peripheral blood cells from weight-losing cancer patients control the hepatic acute phase response by a primarily interleukin-6 dependent mechanism. *Int J Oncol* **15**: 823–827
- Pascussi JM, Gerbal-Chaloin S, Pichard-Garcia L, Daujat M, Fabre JM, Maurel P, Vilarem MJ (2000) Interleukin-6 negatively regulates the expression of pregnane X receptor and constitutively activated receptor in primary human hepatocytes. *Biochem Biophys Res Comm* **274**: 707–713
- Prandota J, Tillement JP, d'Athis P, Campos H, Barre J (1980) Binding of erythromycin base to human plasma proteins. *J Int Med Res* **8**(Suppl 2): 1–8
- Rivory L, Slaviero K, Hoskins JM, Clarke SJ (2001) The erythromycin breath test for the prediction of drug clearance. *Clin Pharmacokin* **40**: 151–158
- Rivory LP, Slaviero K, Seale JP, Hoskins JM, Boyer M, Beale PJ, Millward MJ, Bishop JF, Clarke SJ (2000) Optimizing the erythromycin breath test for use in cancer patients. *Clin Cancer Res* **6**: 3480–3485
- Ugurel S, Rapp G, Tilgen W, Reinhold U (2001) Increased serum concentration of angiogenic factors in malignant melanoma patients correlates with tumor progression and survival. *J Clin Oncol* **19**: 577–583
- Watkins PB, Murray SA, Winkelman LG, Heuman DM, Wrighton SA, Guzelian PS (1989) Erythromycin breath test as an assay of glucocorticoid-inducible liver cytochromes P-450. Studies in rats and patients. *J Clin Invest* **83**: 688–697
- Yancik R, Wesley MN, Ries LA, Havlik RJ, Long S, Edwards BK, Yates JW (1998) Comorbidity and age as predictors of risk for early mortality of male and female colon carcinoma patients: a population-based study. *Cancer* **82**: 2123–2134
- Zamboni WC, Houghton PJ, Thompson J, Cheshire PJ, Hanna SK, Richmond LB, Lou X, Stewart CF (1998) Altered irinotecan and SN-38 disposition after intravenous and oral administration of irinotecan in mice bearing human neuroblastoma xenografts. *Clin Cancer Res* **4**: 455–462