IL-12 p40 prevents the development of chronic enterocolitis in IL-10-deficient mice

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T-helper-1 (Th1) cytokines play an important role in Crohn's disease, and interleukin-12 (IL-12), which is composed of two subunits, p40 and p35, drives Th1 differentiation. In previous reports, IL-12 p40 was shown to prevent IL-12 from binding to the receptor. We demonstrate here the effect of IL-12 p40 overexpression in intestinal epithelia on enterocolitis mediated by Th1 cells in IL-10-deficient (IL-10^{-/-}) mice on a C57BL/6J background. IL-10 deficient (IL-10^{-/-})/T3^b-IL-12 p40⁺ (IL-12 p40⁺) mice and IL-10^{-/-}/T3^b-IL-12 p40⁻ (IL-12 p40⁻) mice were generated by crossing T3^b-IL-12 p40 transgenic mice and IL-10^{-/-} mice. At 8 weeks of age, IL-12 p40⁺ mice did not show any clinical manifestations of colitis. The colon length of IL-12 p40⁻ mice became shorter than that of IL-12 p40⁺ mice. The histological score of IL-12 p40⁺ mice was lower. Interferon-gamma (IFN- γ) production was suppressed in both the mesenteric lymph node cell culture and colon tissue culture of IL-12 p40⁺ mice. There was no significant difference in IL-4 production and tumor necrosis factor-alpha (TNF- α) production between the two groups. These results show that overexpression of IL-12 p40 in intestinal epithelia prevents enterocolitis in IL-10^{-/-} mice by suppressing IFN- γ production, and suggest a potential clinical application of IL-12 p40 for Crohn's disease. Furthermore, these results also suggest that local gene transduction in the intestinal epithelium may be a potent therapeutic approach for Crohn's disease. *Laboratory Investigation* (2004) **84**, 1491–1500, advance online publication, 27 September 2004; doi:10.1038/labinvest.3700175

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Inflammatory bowel disease (IBD), which includes both Crohn's disease and ulcerative colitis, is an idiopathic chronic disease.¹ The precise pathogenesis of IBD is still unclear, but T-helper-1 (Th1) cytokines have been reported to play a pivotal role in the development of Crohn's disease.^{2,3} Recently, various mice models of intestinal inflammation mediated by a Th1 response have been established. These mice models, including interleukin (IL)-2),^{4,5} IL-10, and Stat3⁶-deficient mice, IL-7 transgenic mice,⁷ $Tnf^{\Delta ARE}$ mice,^{8,9} senescence accelerated mice P1/Yit strain,^{10,11} 2,4,6-trinitrobenzenesulfonic acid (TNBS)-treated mice,¹² and CD4+CD45RB^{high} T cell transferred SCID mice,^{13,14} have provided new insight concerning the intestinal inflammation of Crohn's disease and its pathogenesis. The methods

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by which the inflammation is prevented in these models could lead to new cytokine therapies for Crohn's disease.

IL-12, which is a 70 kDa heterodimeric cytokine composed of two disulfide-linked subunits, p40 and p35, is a key cytokine that drives naïve T cells to Th1 differentiation.^{15–17} In Crohn's disease, it has also been reported that the excessive expression of IL-12 produced predominantly by macrophages and dendritic cells promotes the Th1 response.^{18,19} Furthermore, IFN- γ induced by IL-12 can promote the upregulation of IL-12 production.²⁰ This cycle promotes the chronic intestinal inflammation of Crohn's disease. The prevention of this cycle, therefore, could be an effective therapy for Crohn's disease. Although several ways to prevent this cycle have been reported, we focused on IL-12 p40 in the present study. Because IL-12 p40, especially a homodimeric form of IL-12 p40, has affinity for the IL-12 receptor, prevents the binding of a heterodimeric form of IL-12,^{21,22} and is not a foreign protein in vivo, IL-12 p40 would not likely make an unwelcome antibody.

We have used $T3^b$ gene promotor to overexpress IL-12 p40 in the intestine of mice. The $T3^b$ gene encodes one of the thymic antigens, which are nonclassical major histocompatibility complex class I molecules, and in the C57BL/6 mouse strain it is expressed exclusively in the intestinal epithelium.²³ Using the $T3^b$ gene promoter thus enables us to verify whether the intestinal epithelium is the proper target of the gene therapy in mice with experimental transluminal colitis resembling Crohn's disease. We previously reported transgenic mice overexpressing IL-12 p40 in the epithelial cells of the small and large intestines, specifically under the control of the $T3^b$ gene promoter.²⁴ There were no abnormalities in T3^b-IL-12 p40 transgenic mice in a specific pathogen-free condition.

In this study, we generated double-mutant mice by crossing IL-10-deficient (IL- $10^{-/-}$) mice, which develop spontaneous chronic colitis mediated by Th1 cells expressing CD4,^{25–28} and transgenic mice overexpressing IL-12 p40 to assess the therapeutic effects of IL-12 p40 in IL- $10^{-/-}$ mice. We demonstrate that the overexpression of IL-12 p40 in the gastrointestinal tract can suppress the spontaneous chronic enterocolitis of IL- $10^{-/-}$ mice, which suggests the potential of using IL-12 p40 as a therapy for Crohn's disease.

Materials and methods

Mice

The generation of T3^b-IL-12 p40 transgenic mice on a C57BL/6J background has been previously described.²⁴ IL-10^{-/-} mice on a C57BL/6J background were purchased (The Jackson Laboratory, ME, USA). IL- $10^{-/-}/T3^{b}$ -IL-12 p40⁺ (IL-12 p40⁺) mice and IL-10^{-/-}/T3^b-IL-12 p40⁻ (IL-12 p40⁻) mice were generated by crossing IL-10^{-/-} mice and T3^b-IL-12 p40 transgenic mice. The genotyping of the mice was performed by PCR using tail DNA isolated by the sodium dodecyl sulfate-proteinase K method. All mice were maintained under specific pathogen-free conditions in the animal facility at Tohoku University Graduate School of Medicine. At weekly intervals, we inspected the mice for clinical symptoms such as diarrhea and anal prolapse from 3 weeks of age. The body weights of the mice were also measured. Animals were killed by cervical dislocation at 8 weeks of age. The length of the colon was measured for the assessment of morphologic change. The entire tissue sections of the colon were made for histological analysis and colon tissue culture. Mesenteric lymph node cells were also collected for cell culture. All mice received humane care in accordance with the law concerning the protection and control of animals in Japan.

Histology

The colon tissues were fixed in 10% formalin. Paraffin sections were then made and stained with hematoxylin and eosin. Histological analysis was performed in a blinded fashion by one pathologist. The tissue sections were divided into five regions: cecum, ascending, transverse, descending colon, and rectum. The evaluation of inflammation in the entire colon was performed by the summation of the scores for each region, which were graded semiquantitatively from 0 to 4.27,29 The grading represents the degree of severity of mucosal inflammation and epithelial cell hyperplasia as follows: 0, no signs of inflammation; 1, very low level of leukocytic infiltration in the lamina propria with mild epithelial hyperplasia and without the depletion of goblet cells; 2, mild inflammatory cell infiltration in the lamina propria with mild epithelial hyperplasia and depletion of goblet cells; 3, inflammation involved the submucosa and crypt abscess were occasionally found. Moderate epithelial hyperplasia and depletion of goblet cells were also seen; 4, inflammation involved the submucosa, and was sometimes transluminal. Marked epithelial hyperplasia and elongated glands with few goblet cells were found.

Immunohistochemistry

Colon tissues were fixed in periodate-lysine-4% paraformaldehyde overnight at 4°C and then frozen in Optimal Cutting Temperature compound (Sakura Finetechnical Co., Ltd, Tokyo, Japan). Serial sections (6 μ m thick) were cut with a cryostat and placed on 3-amino-propyltriethoxysilane-coated slides. Blocking was performed in Histofine blocking anti-goat serum (Nichirei Tokyo, Japan) for 12 min at room temperature. Samples were washed twice in phosphate-buffered saline (PBS) and then incubated overnight 4°C with a rat anti-mouse IL-12 (p40/p70) monoclonal antibody, C15.6 (Pharmingen, San Diego, CA, USA), at a density of $2.5 \,\mu\text{g/ml}$ in PBS supplemented with 0.5% FBS and 0.05% sodium azide. These slides were washed three times in PBS. Endogenous peroxidase was quenched by incubating the samples in 0.3% H₂O₂/methanol for 12 min at room temperature. And then the slides were incubated for an hour at room temperature with secondary antibody, F(ab') fragments of anti-rat immunoglobulin coupled to horseradish peroxidase (Zymed, San Francisco, CA, USA) at a density of $0.025 \,\mu$ g/ml in PBS. Following subsequent washes in PBS, the color reaction was developed with diaminobenzidine. Slides were counterstained with hematoxylin.

Mesenteric Lymph Node Cell Cultures

Mesenteric lymph node cell cultures were performed in RPMI medium 1640 (Invitrogen, Carlsbad, CA, USA) supplemented with 10% FBS, $50 \,\mu\text{M}$ 2mercaptoethanol, 100 U/ml penicillin/streptomycin (Invitrogen, Carlsbad, CA, USA) and $10 \,\mu\text{g/ml}$ concanavalin A (conA) at 37°C in humidified 5% CO₂ for 24 h using 24-well flat bottom plates (Becton Dickinson Labware, Franklin Lakes, NJ, USA). The density of the mesenteric lymph node cells was adjusted at 2×10^6 cells/ml. The supernatants of the mesenteric lymph node cell culture were collected for cytokine analysis.

Cell Isolation and Cell Culture of Mesenteric CD4⁺ T Cells

Collected mesenteric lymph node cells were centrifuged for 7 min at 1400 rounds per minute, and CD4⁺ T cells were isolated from mesenteric lymph node cells by the MACS magnetic cell sorting system (Militenyi Biotech, Auburn, CA, USA) according to the manufacturer's protocols. The validity of cell isolation was verified by flow cytometry. In all, 96-well flat bottom plates (Becton Dickinson Labware, Franklin Lakes, NJ, USA) were coated with $10 \,\mu \text{g/ml}$ murine anti-CD3 ε antibody (Pharmingen, San Diego, CA, USA) in PBS overnight at 4° C. CD4⁺ T cells at 1×10^{6} cells/ml were then cultured in RPMI medium 1640 supplemented with 10% FBS, 50 μ M 2-mercaptoethanol, 100 U/ml penicillin/streptomycin, and $5 \mu g/ml$ soluble anti-CD28 antibody (Pharmingen, San Diego, CA, USA) in the precoated wells for 24 h. The supernatants of the isolated mesenteric CD4⁺ T-cell culture were collected for cytokine analysis.

Colon Tissue Cultures

Colon tissue biopsy specimens were obtained by the following method:³⁰ The harvested colon tissue was cleared of fecal matter by flushing with PBS, split longitudinally, and six full-thickness colon biopsy specimens with a diameter of 3 mm were obtained from the distal and proximal colon using a dermal punch biopsy instrument (Kai Industries Co., Ltd, Seki, Japan). Each biopsy specimen was then transferred onto a stainless-steel mesh with the mucosal side up in a well of a 24-well tissue culture plate containing RPMI medium 1640 supplemented with 10% FBS, 50 µM 2-mercaptoethanol, 100 U/ml penicillin/streptomycin, and $10 \,\mu \text{g/ml}$ conA. The tissue was cultured for 24 h using the same method as for the mesenteric lymph node cell culture. The supernatants of the colon tissue culture were collected for cytokine analysis.

Western Blot Analysis

The harvested colon tissue was cultured as mentioned above, but the culture medium did not contain 2-mercaptoethanol. The culture supernatants were separated by the sodium dodecyl sulfate-polyacrylamide gel electrophoresis (10%) under reducing or non-reducing conditions. The separated proteins were transferred to a $0.45 \,\mu m$ nitrocellulose membrane (Bio-Rad, CA, USA). Transblots were incubated with a rat anti-mouse IL-12 (p40/p70) monoclonal antibody, C17.8 (Pharmingen, San Diego, CA, USA) and then incubated for an hour with secondary antibody, F(ab') fragments of anti-rat immunoglobulin coupled to horseradish peroxidase (Zymed, San Francisco, CA, USA). The antibody-reactive proteins were visualized using ECL Western Blotting Detection Reagents (Amersham Bioscience Buckinghamshire, England) according to the manufacturer's protocol.

Cytokine Analysis

To detect the cytokine production of IFN- γ , IL-12, IL-4, and TNF- α by the mesenteric lymph node cells, by the CD4 $^{\scriptscriptstyle +}$ T cells and colon tissues, ELISA was performed using a mouse ELISA kit (ENDOGEN, Woburn, MA, USA) according to the manufacturer's protocols.

Statistical Analysis

All data are expressed as the mean+standard deviation. Statistical difference was assessed using Mann–Whitney test. The level of significance was set at *P*<0.05.

Results

Detection of Overexpression of IL-12 p40 in IL-12 p40⁺ mice

We confirmed the overexpression of IL-12 p40 in IL-12 p40⁺ mice. Immunohistochemical analysis of the colon tissues demonstrated that the epithelial cells of the colon in IL-12 p40⁺ mice were strongly stained by anti-IL-12 monoclonal antibody (Figure 1a). Close examination of the sections revealed that the IL-12 p40 expression was mainly by mature absorptive cells facing to the lumen and by lymphocytes (Figure 1b). The epithelial cells of the colon of IL-12 $p40^{-}$ mice were not stained (Figure 1c). However, lymphocytes found in the colonic mucosa were stained in a granular pattern (Figure 1d). Next we performed Western blot analysis to determine what form of IL-12 p40 was present. Western blot analysis showed two bands around 80 and 40 kDa in the supernatant of the colon tissue culture of IL-12 $p40^+$ mice under a nonreducing condition (Figure 2). The position of the upper band was the same as that of the control recombinant IL-12 p40 homodimer, which is the upper one. Moreover, the bands of the IL-12 p40⁺ mice, control recombinant IL-12 p40 homodimer, and recombinant IL-12 heterodimer around 80 and 70 kDa shifted downward to around 40 kDa under a reducing condition. One very weak band of the IL-12 p40⁻ mice could be seen around 40 kDa. We also confirmed that the overexpression of IL-12 p40 was found not in lymph node cells but mainly in the colonic mucosa using ELISA.



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Figure 1 Immunohistochemical analysis. Sections from the colon were incubated with rat anti-mouse IL-12 monoclonal antibody, C15.6, and subsequently incubated with anti-rat immunoglobulin conjugated to HRP. The color reaction was developed with diaminobenzidine. Slides were counterstained with hematoxylin. (a) Colon of IL-12 p40⁺ mice. (\times 200). (b) Close inspection of the colon of IL-12 p40⁺ mice (\times 400). (c) Colon of IL-12 p40⁻ mice (\times 200). (d) Close inspection of the colon of IL-12 p40⁻ mice (\times 400).

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Figure 2 Western blot analysis. IL-12 p40 produced by cultured colon tissue and control protein, which is a mixture of mouse recombinant IL-12 p40 homodimer and recombinant IL-12 heterodimer, were separated by SDS-PAGE (10%) under reducing or nonreducing conditions, and transferred to a nitrocellulose membrane. Transblots were incubated with a rat anti-mouse IL-12 (p40/p70) monoclonal antibody, C17.8, and then incubated with secondary antibody coupled to horseradish peroxidase.

ELISA showed the massive production of IL-12 p40 in the colon tissue culture of IL-12 $p40^+$ mice (Figure 3a). The IL-12 p40 production in colon tissue of IL-12 p40⁻ mice was also induced, but the amount was far less than that of IL-12 p40⁺ mice. The amount of IL-12 measured by ELISA consisted of the IL-12 p40 produced by the epithelium and IL-12 produced by inflammatory cells in the colon. However, we have reported previously the IL-12 p40 expression in the intestine of T3^b-IL-12 p40 transgenic mice that were not crossed IL- $10^{-/-}$ mice was restricted to the epithelial cells.²⁴ Thus, the IL-12 p40 in the colonic tissue culture of IL-12⁺ mice measured by ELISA would be produced mainly by the colonic epithelial cells. IL-12 p40 production by mesenteric lymph node cells was induced in IL-12 p40⁺ mice and IL-12 p40⁻ mice. But no remarkable difference was found between the two groups (Figure 3b).

In contrast to IL-12 $p40^-$ Mice, IL-12 $p40^+$ Mice Gained Weight Normally

At 8 weeks of age, the IL-12 $p40^-$ mice showed clinical manifestations such as diarrhea, rectal prolapse, and melena, but the IL-12 $p40^+$ mice did not show any manifestations of colitis. In fact, IL-12 $p40^+$ mice observed for 24 weeks did not develop histologic evidence of chronic enterocolitis (data not shown). Disease manifestations of the IL-12 $p40^-$ mice appeared nearly at the same age as IL-10⁻⁷⁻ mice that were not crossed T3^b-IL-12 p40 mice under specific pathogen-free conditions of our animal facility. We measured the body weight of the mice to assess the progression of colitis. IL-12 $p40^+$ mice and IL-12 $p40^-$ mice gained weight similarly until 5 weeks of age. However, the IL-12 $p40^-$ male mice gained less



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Figure 3 ELISA using IL-12 total ELISA Kit. The manufacturer described that this assay is specific for mouse IL-12 p40 and p70. The measured value of mouse IL-12 homodimer was about twice the true value in this assay. Results are expressed as the mean±standard deviation of each group. (a) IL-12 production of mesenteric lymph node cells. n.s. indicates no remarkable difference between p40⁻ and p40⁺ mice. (b) IL-12 production of colon tissue culture. **P<0.01 between IL-12 p40⁺ mice and IL-12 p40⁻ mice.

weight than the IL-12 $p40^+$ male mice from 6 weeks of age. At 8 weeks of age, the IL-12 $p40^+$ male mice had gained weight to a similar extent as the wild-type mice (data not shown), but the body weights of the IL-12 $p40^-$ male mice were significantly lower than those of the IL-12 $p40^+$ mice (Figure 4). IL-12 $p40^+$ female mice also steadily gained weight similarly as the wild type (data not shown) and IL-12 $p40^-$ female mice. At 8 weeks of age, there was no statistical difference between the mean body weight of IL-12 $p40^-$ and that of IL-12 $p40^+$ female mice.

Influence of Inflammation on the Colon was Found Macroscopically Only in the IL-12 p40⁻ Mice

The colon of IL-12 $p40^-$ was edematous and thickened by inflammation with hyperplasia and contained loose stool, while the colon of IL-12 $p40^+$



Figure 4 Change in body weights. The ratios of body weight to the weight at 3 weeks of age were plotted. Each group consisted of 10 mice. IL-12 p40⁺ male mice gained weight similarly to the wild type, but IL-12 p40⁻ male mice gained less weight than IL-12 p40⁺ mice from 6 weeks of age. The weights of p40⁻ female mice were similar to those of p40⁺ mice during the observation period. *P < 0.05 between IL-12 p40⁺ mice and IL-12 p40⁻ mice.

seemed to be normal macroscopically and the stool in the colon was also normal. The colon length of IL-12 $p40^-$ mice was shortened by inflammation, and there was significant difference between IL-12 $p40^+$ and IL-12 $p40^-$ mice in the colon length (Figure 5).

Histological Score in IL-12 p40⁺ Mice was Significantly Lower

We investigated the influence of IL-12 p40 overexpression on intestinal inflammation by histological examination. Histological sections of the colon of IL-12 p40⁺ mice revealed few findings of inflammation (Figure 6a). Close examination of the sections revealed slight hyperplasia and scant infiltration of inflammatory cells into the mucosa (Figure 6b). Histological sections of the colon of IL-12 p40⁻ mice revealed infiltration of inflammatory cells into the mucosa and submucosa, severe hyperplasia, depletion of goblet cells (Figure 6c), and mucosal injuries (Figure 6d). The disease score distribution showed that these inflammatory changes could be found in the IL-12 p40⁻ group even if the mice did not manifest any clinical symptoms. In contrast, the histological observation of IL-12 p40⁺ mice showed little inflammation of the colon. The mean histological score was significantly lower in the IL-12 p40⁺ mice (Table 1). The intestinal inflammation was patchy and the regional disease score revealed that the inflammation was severer in the proximal side of the colon than in the distal side of colon. However, there was no difference in the suppressive effect of IL-12 p40 overexpression among the regions. In addition, there was no statistical difference between male mice and female mice in severity of colitis or any other parameters measured in contrast with body weight changes.



Figure 5 Mean length of the colon. Upper, male mice (n=8). Lower, female mice (n=7). The length of the colon of IL-12 p40⁻ mice was significantly shorter than that of IL-12 p40⁺ mice. *P < 0.05 between IL-12 p40⁺ mice and IL-12 p40⁻ mice.

Number of Mesenteric Lymph Node Cells and IFN- γ Production in IL-12 p40⁺ Mice were Significantly Reduced

To investigate the effect of IL-12 p40 overexpression we assessed mesenteric lymph node cell numbers as well as IFN- γ production by the mesenteric lymph node cells and colon tissue. The number of lymph node cells was significantly lower in IL-12 p40⁺ mice than in IL-12 $p40^{-}$ mice (Table 2), and it was slightly higher as compared with wild-type mice and T3^b-IL-12 p40 transgenic mice (data not shown). We then performed cell isolation of CD4⁺ T cells. The number of CD4⁺ T cells in IL-12 p40⁺ mice was three-fold lower than that in IL-12 $p40^-$ mice (Table 2). These data may indicate that IL-12 p40 suppressed the expansion of pathogenic T cells. ELISA showed that IFN- γ production by the mesenteric lymph node cells was significantly suppressed in IL-12 p40⁺ mice (Figure 7a). The mean concentrations of IFN- γ in the culture of mesenteric lymph node cells of IL-12 p40⁺ mice and IL-12 p40⁻ mice (n = 9) were 660 ± 1539 and 3341 ± 2208 pg/ml, respectively. This tendency was also found in the culture of isolated CD4⁺ T cells $(91.0 \pm 43.6 \text{ and } 1930 \pm 794 \text{ pg/ml for IL-12 p40}^+ \text{ mice}$ and IL-12 p40⁻ mice, respectively (n=5)) and colon tissue culture (716+639 vs 3442+1007 pg/ml for IL-12) $p40^+$ mice and IL-12 $p40^-$ mice, respectively (n=7)). Next we assessed the production of other cytokines. IL-4, which is one of the Th2 cytokines that prevent Th1 differentiation, was secreted at very low levels by mesenteric lymph node cells, colon tissue and isolated $CD4^+$ T cells (Figure 7b). There was no difference in IL-4 secretion between IL-12 $p40^+$ mice and IL-12 $p40^-$ mice in any of these cultures. The mean concentrations of IL-4 in the cultures of mesenteric lymph node cells were 5.10 ± 0.272 vs 11.1 ± 14.1 pg/ ml (n = 9), in the cultures of isolated CD4⁺ T cells IL-4 was lower than detection limit vs 21.4 ± 36.8 pg/ml (n=5), and in the colonic tissue cultures it was $6.78 \pm 4.70 \text{ pg/ml } vs$ lower than detection limit (n = 7) for IL-12 p40⁺ mice and IL-12 p40⁻ mice, respectively. We then investigated TNF- α , another proinflammatory mediator (Figure 7c), but there was no significant

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Figure 6 Histological sections of the colon of IL-12 p40⁺ and IL-12 p40⁻ mice. Sections were stained by hematoxylin and eosin. (a) Colon of IL-12 p40⁺ mice (\times 40). (b) Higher magnification of the section from IL-12 p40⁺ mice (\times 200). (c) Colon of IL-12 p40⁻ mice (\times 40). (d) Higher magnification of the section from IL-12 p40⁻ mice (\times 100).

Table 1 Effect of IL-12 p40 overproduction on spontaneous enterocolitis of IL-10 $^{-/-}$ mice

Mice	Number of mice affected ^a	Disease score distribution ^b					Mean disease score
		1-4	5–8	9–12	13–16	17–20	(0–20)
IL-12 p40 ⁺ mice IL-12 p40 ⁻ mice	0/5 3/5	3 0	2 1	0 0	0 2	0 2	$\begin{array}{c} 4.8 \pm 4.7^{**} \\ 14 \pm 2.2 \end{array}$

^aNumber of mice showing some clinical manifestations such as diarrhea, hematochezia, and rectal prolapse. Each group consisted of five mice. ^bSeverity of enterocolitis graded semiquantitatively from 0 to 4.

**P < 0.01 compared with IL-12 p40⁻ mice.

difference in the production of TNF- α between IL-12 p40⁺ mice and IL-12 p40⁻ mice (236±91.6 and 283±140 pg/ml, respectively, in the cultures of the mesenteric lymph node cells (n=9), 335±145 vs 289±146 pg/ml, respectively in the cultures of isolated CD4⁺ T cells (n=5), and 260±166 vs 462±175 pg/ml, respectively in the colonic tissue cultures (n=7)).

Discussion

We demonstrated in the present study that the overexpression of IL-12 p40 in the intestinal epithelium prevented the development of chronic enterocolitis mediated by a Th1 response in IL- $10^{-/-}$ mice in all parameters examined.

Mice	Number of mesenteric lymph node cells; $n = 10 (\times 10^7)$	Number of isolated $CD4^{+}$ T cells; n = 5 (× 10 ⁶)
IL-12 p40+ mice	3.0 ± 1.7 **	$1.2 \pm 0.70^*$
IL-12 p40- mice	6.3 ± 2.7	3.7 ± 2.8

Table 2 Number of mesenteric lymph node cells and isolated CD4* $T\ cells^a$

^aValues are given as mean \pm standard deviation.

*P < 0.05 and **P < 0.01 compared with IL-12 p40⁻ mice.

IL-12 promotes Th1 differentiation and IFN- γ production,¹⁵ and consequently promotes chronic enterocolitis in IL- $10^{-/-}$ mice, especially during the early phase of colitis.³¹ IL-12 p40 has been reported to have the opposite effect on IL-12 heterodimer,^{21,32} especially when in a homodimeric form.²¹ Furthermore, Yoshimoto et al³³ previously reported that transgenic mice overexpressing IL-12 p40 regulated by the liver-specific promoter showed reduced Th1 responses.³³ Thus, the overexpression of IL-12 p40 could suppress the Th1 response and consequently suppress the development of spontaneous chronic enterocolitis in IL- $10^{-/-}$ mice. Our data are consistent with this scenario insofar as we demonstrated a suppressed Th1 reaction. The numbers of mesenteric lymph node cells were reduced, as were the mesenteric CD4⁺ T cells in IL-12 p40⁺ mice. ELISA revealed the suppression of IFN- γ production in the cultures of mesenteric lymph node cells and colon tissue in IL-12 p40⁺ mice. In addition, we confirmed the overexpression of IL-12 p40 exclusively in the epithelium of IL-12 p40⁺ mice by immunohistochemical analysis and ELISA. Western blot analysis revealed that most of IL-12 p40 existed in a homodimeric form.

Many reports have shown that overexpression of p40 suppresses Th1-mediated inflammation. However, there are several reports describing that IL-12 p40 promotes Th1-mediated inflammation. In mice with experimental autoimmune encephalomyelitis, which is Th1 cell-mediated autoimmune demyelinating disease, and human multiple sclerosis, IL-12 p40 was overexpressed in the central nervous system, and that the level of IL-12 p40 was such far higher than that of p35.34-36 We think IL-12 p40 could be produced as a damper in such situations. van Boxel-Dezaire *et al*³⁴ performed the cytokine analysis in patients with relapsing-remitting multiple sclerosis treated with IFN- β 1b and described that the baseline levels of IL-12 p35 mRNA were lower in the clinical responders than in the nonresponders. Such a result may indicate that patients with suppressed bioactive IL-12 under the influence of overexpressed IL-12 p40 could be clinical responders. Further studies are required to determine the role of p40 in experimental autoimmune encephalomyelitis and human multiple sclerosis.



Figure 7 Cytokine analysis. Primary cultures of mesenteric lymph node cells and colon tissue were performed for 24 h. The supernatants were collected for cytokine analysis. Cytokine production was determined by ELISA in culture supernatants from mesenteric lymph node cells (n=9), isolated CD4⁺ T cells and colon tissue (n=7). Data are shown as mean±standard deviation pg/ml per group. (a) IFN- γ , (b) IL-4, (c) TNF- α . **P<0.01 between IL-12 p40⁻ mice and IL-12 p40⁻ mice. n.s. means no significant difference. ND means not detectable.

Recently, IL-23, which is composed of two subunits, p19 and p40, was discovered.³⁷ It could be a novel cytokine that promotes the proliferation of memory T cells, and has been reported to possess to some extent a similar function to IL-12.³⁸ The p40 subunit of IL-23 is a protein common to IL-12. Thus, the possibility that the effects of p40 on the IL-23 pathway prevented the development of colitis in the present study cannot be excluded. For example, IL-12 p40 might act a competitor against IL-23 and thereby prevent the development of colitis.

Toxicity from the overexpression of IL-12 p40 was not seen in this study. Some investigators have reported that mice with blocked IL-12 activity were susceptible to infection. Tripp *et a*¹³⁹ have reported that anti-IL-12 antibody-treated mice showed increased susceptibility to *Listeria burden*. Our T3^b-IL-12 p40 transgenic mice did not show any abnormalities and their longevity was the same as that of wild-type mice in specific pathogen-free conditions.

In this study, we also demonstrated the potential use of local gene transduction in the intestinal epithelium as a therapy for Crohn's disease. Some investigators have reported the efficacy of gene therapy⁴⁰⁻⁴² with adenovirus enemas. The target cells of the adenovirus are ambiguous (epithelium, fibroblast, lymphocyte, etc). Thus, it remains to be shown which kinds of cells in the intestine are the best targets of gene transduction to suppress transluminal colitis. In this study, the overexpression of IL-12 p40 was found exclusively in the epithelial cells of the IL-12 p40⁺ mice. Although the inflammation of IL- $10^{-/-}$ mice was transluminal, the overexpression of IL-12 p40 in the epithelium prevented this transluminal inflammation. This result, therefore, might indicate that local gene transduction in the intestinal epithelium alone could be a potent therapy (including the prevention of relapse) for Crohn's disease.

The clinical benefits of cytokine therapies such as with an anti-TNF- α agent have been reported,⁴³ and some investigators have reported the efficacy of anti-IL-12 antibodies in preventing the development of the intestinal inflammation mediated by a Th1 response.^{12,17,29,44} In contrast, it has been anticipated that the use of antibody agent could induce the production of an anti-drug antibody such as an antiidiotype antibody. We think that IL-12 p40 would not induce the synthesis of an unwelcome antibody because it exists naturally *in vivo*.

In conclusion, the present data demonstrated that the overexpression of IL-12 p40 by the T3^b-IL-12 p40 transgene in the gastrointestinal tract can suppress the production of IFN- γ by CD4⁺ Th1 cells, which mediate intestinal inflammation. These data suggest a potential clinical application of IL-12 p40 for IBD. Furthermore, we also demonstrated the possibility of employing local gene transduction in the intestinal epithelium as a therapy for Crohn's disease.

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