DEVELOPMENT AND CANCER

Tampering with the cell cycle's brakes

To stop the cell cycle, tumour suppressors such as the retinoblastoma protein (Rb) apply the brakes. A *Nature* paper by Anna Lasorella and colleagues describes how Id2 — a dominant-negative inhibitor of helix–loop–helix DNA-binding proteins — gets the wheels turning again.

Retinoblastoma protein is essential for mammalian development: knockouts die during embryogenesis. Lasorella *et al.* report that knocking out *Id2* rescues Rb^{-l-} embryos. Defective myogenesis kills $Id2^{-l-}Rb^{-l-}$ mice shortly after birth, but they show none of the hallmarks of Rb^{-l-} mice — too much proliferation and apoptosis in the haematopoietic and nervous systems. Id2 therefore perpetrates the Rb^{-l-} phenotype; but how? Immunoprecipitates revealed that active, hypophosphorylated Rb binds Id2 and, to tip the balance against tumour suppression, Id2 has to be in molar excess of Rb.

An intact Rb pathway is needed to prevent tumorigenesis, so Lasorella and colleagues

asked whether Id2 is overexpressed in neuroblastoma cell lines, in which N-*myc* amplification typically bypasses Rb. Remarkably, Id2 expression correlated with N-*myc* amplification. What's more, Myc's effect on Id2 expression (which also extends to c-Myc) is direct, owing to two high-affinity Myc-binding sites in the Id2 promoter. Deletion of these sites abolished Id2 expression in response to Myc.

By producing a surfeit of Id, then, Myc can override Rb's attempts to halt the cell cycle. But what targets of Id are responsible for the $Rb^{-/-}$ phenotype? And what about the other Id family members, Id1 and Id3? These questions, and others, await the next cycle tour.

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W References and links

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APOPTOSIS

Viral pirates hijack Bcl-2



Viruses are modern-day buccaneers. They ride the cellular seas, hijacking proteins and using them to promote their own survival. But how? Ojala *et al.*, reporting in the November issue of *Nature Cell Biology*, describe how the Kaposi's sarcoma herpesvirus (aka human herpesvirus 8; HHV8) interferes with apoptotic signalling pathways in its host. The virus encodes a pirated cyclin (vcyclin), which forms a complex with a cellular cyclin-dependent kinase, CDK6. This complex can induce apoptosis, and Ojala *et al.* now show that it probably does so by phosphorylating — and inactivating — the cellular anti-apoptotic molecule Bcl-2. The authors find that Bcl-2 and CDK6 associate in cell extracts, and they show that the targets for phosphorylation are two serine residues in an unstructured 'loop' region of cellular Bcl-2.

But herein lies a paradox. Why should the virus inactivate a molecule that protects the host cell from apoptotic death? One might think it in the best interests of the virus to keep its host alive. It turns out, however, that cellular Bcl-2 has other functions that the virus finds less than savoury; for instance, it has been reported to impair cell-cycle progression when overexpressed. Moreover, host cell death is advantageous to the virus in that it allows the spread of viral particles. There is a problem, though — the risk that, when the virus blocks cellular Bcl-2, the host cell will die before the viral replication cycle is complete. Here, in an ultimate act of viral skulduggery, HHV8 produces its own Bcl-2 homologue. This virus-encoded protein lacks the crucial unstructured loop, so it cannot be phosphorylated or inactivated by v-cyclin–CDK6.

Infection of host cells with HHV8 has previously been shown to be linked to apoptosis, and the lesions associated with Kaposi's sarcoma contain some apoptotic cells. The next step, then, will be to work out how the acts of viral piracy uncovered by Ojala *et al.* link HHV8 to tumour formation and to the development of Kaposi's sarcoma.

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