parts of the polypeptide (8 among the first 22 residues and 8 among the last 18 residues), while a stretch of 10 acidic residues is found in an internal region, between amino acids 135 and 163. These features may be significant for the biological activity of the spoIIG gene product.

The encoded protein seems to be synthesized in a very low amount as we have been unable to express it in B. subtilis maxicells in various experimental conditions; this suggests that the protein may have a regulatory role in sporulation. Note that the spoIIG gene is transcribed only in the early stages of sporulation and not during vegetative growth. This was demonstrated by hydridization experiments in which the gene was used to probe mRNAs isolated at different stages of growth and sporulation (results not shown).

To elucidate the nature of the polypeptide encoded by the spoIIG gene, we searched for homologies with known proteins stored in the Dayhoff protein sequence data bank (~2,300 sequences). An important homology was found with the product of the E. coli  $rpoD^{13}$  gene, the sigma factor. Figure 3a shows the alignments of the two related segments of the B. subtilis spoIIG gene product and of the E. coli sigma factor. These aligned sequences exhibit a remarkable similarity: of the 65 positions compared, 29 (44.6%) are occupied by identical amino acids. When chemically similar pairs of amino acids are taken into account<sup>14</sup>, this number rises to 44 (67.7%). It was not necessary to introduce gaps to maximize this homology. Further-more, statistical tests<sup>15,16</sup> show that the similarities between the aligned sequences are highly significant; the probability that such a similarity is fortuitous is  $<10^{-9}$  (Fig. 3*a*).

Analysis of the corresponding nucleotide sequences also revealed a high degree of similarity at the DNA level (100 matches over 195 positions, that is 51.3%; see Fig. 3b). A striking feature of the similarities is their clustering into three blocks of 34, 35 and 30 bp, where 73.5%, 80% and 70% of the respective positions are occupied by identical nucleotides. As the codon usage is not the same in E. coli and in B. subtilis (as indicated, for example, by the respective usages of the trpE genes of these two organisms<sup>17,18</sup>), such similarities strongly suggest that the two genes are evolutionarily related.

The regions shown in Fig. 3 correspond to different internal parts of the two proteins: residues 58-127 of the 239-amino acid polypeptide encoded by spoIIG and residues 375-439 of the E. coli sigma factor, a polypeptide of 613 amino acids<sup>13</sup>. However, their high degree of similarity strongly suggests a common function for these two protein domains. This leads to the attractive hypothesis that the spoIIG gene product may act as a specific sigma-like factor and that the region it has in common with the genuine E. coli sigma factor may similarly be involved in binding to the core RNA polymerase. This hypothesis is strengthened by the known functional and structural relationship between the E. coli and B. subtilis core RNA polymerases<sup>19-21</sup>. Moreover, recent nucleotide sequence analysis of the *B. subtilis*  $\sigma^{55}$  structural gene (R. H. Doi, personal communication) indicates that the homologous region in the E. coli sigma factor and the B. subtilis spoIIG gene product is conserved in the major sigma factor of *B. subtilis*,  $\sigma^{55}$ , confirming previous immunological experiments<sup>22</sup>. One obvious possibility is that the *spo*IIG gene encodes  $\sigma^{29}$ , a polypeptide associated with RNA polymerase purified from B. subtilis sporulating cells and conferring on it new promoter recognition specificity <sup>23</sup>.  $\sigma^{29}$  is a  $M_r$  27,000-29,000 polypeptide<sup>4,23,24</sup> and appears about 1 h after the onset of sporulation<sup>23</sup>, the exact moment at which the temperaturesensitive period starts in strain 279.1, a strain mutated in the spoIIG gene<sup>8</sup>. However, sporulation involves the expression of multiple gene sets and could rely on the presence of several alternative sigma-like proteins. The spoIIG gene product could equally be one of these, as yet uncharacterized, transcriptional factors.

We thank Françoise de la Torre for technical assistance in cloning the spoIIG gene, Isabelle Giri for help with computer sequence analysis and Jocelyne Mauger for typing the manuscript. This work was supported by a CNRS grant (LA 136) to P.S. and J.B.

Note added in proof: The region of homology between the B. subtilis spoIIG gene product and the E. coli sigma factor has recently been found to be conserved also in the heat shock regulatory protein of E. coli<sup>27</sup>.

Received 18 June; accepted 6 September 1984.

- 1. Piggot, P. J. & Coote, J. G. Bact. Rev. 40, 908-962 (1976)
- Henner, D. J. & Hoch, J. A. Microbiol. Rev. 44, 57-82 (1980).
  Losick, R. & Pero, J. Cell 25, 582-584 (1981).
- 4. Fukuda, R. & Doi, R. H. J. Bact. 129, 422-432 (1977)
- Shimotsu, H., Kawamura, F., Kobayashi, Y. & Saito, H. Proc. natn. Acad. Sci. U.S.A. 80, 658-662 (1983).
- Ramakrishna, N., Dubnau, E. & Smith, I. Nucleic Acids Res. 12, 1779-1790 (1984).
- Bouvier, J., Stragier, P., Bonamy, C. & Szulmajster, J. Proc. natn. Acad. Sci. U.S.A. (in the press)
- Young, M. J. Bact. 126, 928-936 (1976).
- Foung, M. J. Bact. 126, 925-950 (1970).
  Ayaki, H. & Kobayashi, Y. J. Bact. 158, 507-512 (1984).
  Bonamy, C. & Szulmajster, J. Molec. gen. Genet. 180, 57-65 (1982).
- Maxam, A. M. & Gilbert, W. Meth. Enzym. 65, 499-560 (1980).
  McLaughlin, J. R., Murray, C. L. & Rabinowitz, J. C. J. biol. Chem. 256, 11283-11291 (1981).
- Burton, Z. et al. Nucleic Acids Res. 9, 2889-2903 (1981).
  Schwartz, R. M. & Dayhoff, M. O. Atlas of Protein Sequence and Structure Vol. 5, Suppl. 3 (ed. Dayhoff, M. O. )353-358 (National Biomedical Research Foundation, Washington, D.C., 1978).
- 15. Needleman, S. B. & Wunch, C. D. J. molec. Biol. 48, 443-453 (1970).
- Brechtman, S. D. & Walter, C. D. J. molec. Biol. 40, 40-50 (1978).
  Smith, T. F., Waterman, M. S. & Fitch, W. M. J. molec. Evol. 18, 38-46 (1981).
  Nichols, B. P., Van Cleemput, M. & Yanosfsky, C. J. molec. Biol. 146, 45-54 (1981).
  Band, L., Shimotsu, H. & Henner, D. J. Gene 27, 55-65 (1984).
- Losick, R., Shorenstein, R. G. & Sonenshein, A. L. Nature 227, 910-913 (1970).
  Shorenstein, R. G. & Losick, R. J. biol. Chem. 248, 6170-6173 (1973).
- 21. Achbeyer, E. C. & Whiteley, H. R. J. biol. Chem. 255, 11957-11964 (1980).
- Wong, S. L. & Doi, R. H. J. biol. Chem. 257, 11932-11936 (1982). Haldenwang, W. G., Lang, N. & Losick, R. Cell 23, 615-624 (1981) 22. 23.
- Linn, T., Greenleaf, A. L. & Losick, R. J. *biol. Chem.* **250**, 9256–9261 (1975).
  Ehrlich, S. D. *Proc. natn. Acad. Sci. U.S.A.* **75**, 1433–1436 (1978).
- Vieira, J. & Messing, J. Gene 19, 259-268 (1982).
- 27. Landick, R. et al. Cell 38, 175-182 (1984)

## Erratum

New evidence that growth in 3T3 cell cultures is a diffusion-limited process

G. A. Dunn & G. W. Ireland Nature 312, 63-65 (1984)

BECAUSE of an editorial error, an incorrect figure was used in this letter. The correct Fig. 3 is shown below.

