

* Down field relative to 3-(trimethylsilyl) propylsulphonic acid (Na salt) as internal standard.



to 12.3 c.p.s. for the olefinic protons of the trans and cis isomers respectively⁵. It is of interest that the extinction coefficient of the *cis* isomer of cinnamic acid is much less than the trans⁶, while those of urocanic acid are quite similar.

Because of the large amount of material needed for the nuclear magnetic resonance studies it was necessary to use isotope techniques to identify the new ultra-violet light absorbing product formed in epidermis following Guinea-pigs were injected with 20 µc. of irradiation. L-histidine-14C (uniformly labelled) intraperitoneally and 24 h later they were depilated with wax and given a minimal erythema dose of ultra-violet light with a 'Hanovia' hot quartz lamp. The epidermis was separated by heating the skin to 50° C for 1 min and the urocanic acid isolated on a 'Dowex-1 formate' column as previously described⁷. Two radioactive compounds were identified by chromatography and autoradiography. The spots were eluted from the paper and chromatographed with the cis and trans isomers to which they corresponded exactly.

The data presented confirm that natural urocanic acid consists of the trans isomer contaminated with some cis and that following irradiation there is a trans to cis isomerization. This same reaction also occurs in the epidermis where urocanic acid is present at a high concentration.

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¹ Masuda, K., Jap. J. Pharm., 6, 152 (1957).

² Kurogochi, Y., Fukui, Y., and Nakagawa, Y. I., Jap. J. Pharmacol., 6, 147 (1957).

³ Edlbacher, S., and Heitz, F., Hoppe-Seyl. Z., 279, 63 (1943).

⁴ Anglin, J. H., and Everett, M. A., Biochim. Biophys. Acta, 88, 492 (1964).
⁵ Roberts, J. D., and Caserio, M. C., Basic Principles of Organic Chemistry, 161 (W. A. Benjamin, Inc., New York, 1964).

⁶ Brande, E. A., Ann. Rep. Chem. Soc., 42, 105 (1945).

⁷ Baden, H. P., and Pathak, M. A., Biochim. Biophys. Acta (in the press).

Cortisone-induced Lipaemia and Hepatic Steatosis in the Male Rat

Ir has previously been reported that female rats develop lipaemia and fatty livers during treatment with cortisone¹, and, more recently, the pathogenesis of this has been traced to a markedly enhanced mobilization of fatty acids from adipose tissue². The mechanism thus seems quite different from that accounting for the fat accumulation in the livers of female, but not of male, rats during ethionine administration³, and it is not reversed or prevented by measures which are effective during ethionine treatment². Nevertheless, it is important to clarify whether the effect could also be observed in male rats.

Male albino 190-200 g rats were treated in a manner identical to that used previously for female rats¹. All rats received tube feedings three times daily of a liquid diet; control rats gained weight well (average weight 247 g after 3 weeks). Experimental rats received 6.25 mg cortisone intramuscularly daily, while controls received saline. The experimental rats lost weight (average weight 170 g after 3 weeks). Animals were killed after 3 weeks; serum and liver lipids were extracted and total glycerides estimated by the van Handel–Zilversmit procedure⁴. Liver protein content was determined on a saline homogenate by the method of Oyama and Eagle⁵.

Results of lipid analysis are shown in Table 1. The magnitude of rise of serum and hepatic lipids is similar to that seen in female rats¹. Thus, it seems clear that no sex difference can be demonstrated for the fatty acid mobilizing properties of cortisone.

Table 1

	Serum glyceride*	Liver glyceride†
Untreated controls (6) Cortisone-treated, pair-fed animals (3)	$\begin{array}{c} 1{\cdot}42\pm0{\cdot}24\\ 3{\cdot}42\pm0{\cdot}18\end{array}$	$\begin{array}{c} 0.208 \pm 0.06 \\ 0.850 \pm 0.30 \end{array}$
+ 1 U C		

* mmoles/1. of serum. † mmoles/g of liver protein. All figures are given ± one S.D. Figures in parenthesis represent number of animals.

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¹ Hill, jun., R. B., and Droke, W. E., Proc. Soc. Exp. Biol. and Med., 114, 766 (1963).

² Hill, jun., R. B., Droke, W. E., and Hays, A. P. Exp. Mol. Path., 4, 320 (1965).

³ Villa-Trevino, S., Shull, K. H., and Farber, E., J. Biol. Chem., 238, 1757 (1963)

⁴ van Handel, E., and Zilversmit, D. B., J. Lab. Clin. Med., 50, 152 (1957).

⁵ Oyama, V. I., and Eagle, H., Proc. Soc. Exp. Biol. and Med., 91, 305 (1956).

In vivo Conversion of Y-Hydroxybutyrate into **γ-Aminobutyrate**

Y-AMINOBUTYRATE (GABA) transaminates in normal brain to succinic semialdehyde^{1,2} which in turn oxidates to succinic acid³

Moreover, Fishbein and Bessman⁴ report that in the soluble fraction of brain homogenate, succinic semialdehyde is reduced to γ-hydroxybutyrate by action of a DPN+ depending enzyme, indistinguishable from lacticdehydrogenase; y-hydroxybutyrate is then lactonized to γ-butyrolactone⁵. γ-Hydroxybutyrate and γ-butyrolactone have been recognized in large amounts in the normal nervous system and they seem the only physiological compounds in the nervous system which have an anaesthetic activity6.

This communication reports on the variation of γ -aminobutyrate content in rat brain after injection of γ -hydroxybutyrate.

Male albino rats (200 g weight) were injected intraperitoneally with 500 mg/kg of γ -hydroxybutyrate. The rats were killed 2 h after injection and the brain removed and homogenized with 70 per cent alcohol (20 ml./g brain). After centrifugation and extraction with chloroform (10 ml./g brain), the alcohol layer was dried in vacuo at 40° C; the residue was dissolved with 1 ml. water/g brain.

 γ -Aminobutyrate was separated by paper electrophoresis; 25 or 50 µl. of residue were utilized. Paper: Whatman No. 1; pyridine-acetic acid-water, buffer (250:50:900); pH 6, 12 V/cm, for 1.5 h. Paper strips, dried with hot air, were sprayed with 0.1 per cent ninhydrin in butanol (Fig. 1). A Chromoscan apparatus was used for reading the quantitative measurements. The standard curve