The finback whale vasopressin has been compared with beef vasopressin by electrophoresis on paper at different pH's and by structural studies. The amino-acid sequence, determined by the chemical and enzymatic methods used earlier in the study of beef vasopressin10, is the same in the two peptides, the structure being:

Cys. Tyr. Phe. Glu(NH₂). Asp(NH₂). Cys. Pro. Arg. Gly(NH₂).

It seems, therefore, that the posterior pituitary of the finback whale contains the same oxytocin and argininevasopressin found for most other mammals, but the difference resides in the proportions of the two hormones. the arginine-vasopressin being in the whale gland about 5 times more abundant than oxytocin.

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¹ Acher, R., Symp. Zool. Soc. Lond., No. 9, 83 (1963).

- ² Acher, R., Chauvet, J., Chauvet, M. T., and Crepy, D., Biochim. Biophys. Acta, 51, 419 (1961); 58, 624 (1962).
 ³ Heller, H., and Pickering, B. T., J. Physiol., 155, 98 (1961).
- ⁴ Rasmussen, H., and Craig, L., Endocrinol., 68, 1051 (1961).
- ⁵ Popence, A. E., Lawler, H. C., and du Vigneaud, V., J. Amer. Chem. Soc., 74, 3713 (1952).
- Geiling, E. M. K., Bull. Johns Hopkins Hospital, 57, 123 (1935).
- ⁷ McClosky, W. T., Miller, L. C., and Le Mesurier, D. N., J. Pharmacol. Exp. Therap., 57, 132 (1936).
 ⁸ Acher, R., Light, A., and du Vigneaud, V., J. Biol. Chem., 233, 116 (1958).

* Light, A., and du Vigneaud, V., Proc. Soc. Exp. Biol. N.Y., 98, 692 (1958). ³³ Acher, R., Chauvet, J., and Lenci, M. T., Bull. Soc. Chim. Biol., **40**, 2005 (1958).

²¹ Chauvet, J., Lonci, M. T., and Acher, R., Biochim. Biophys. Acta, 38, 266 (1960).

¹² Spackman, D. H., Stein, W. H., and Moore. S., Anal. Chem., 30, 1190 (1958).

Inhibition of Dehydrogenases by Salicylate

Euler and Ahlstrom¹ found that salicylate inhibited glucose and lactate dehydrogenases and that the inhibitory effect of the drug on the latter enzyme could be removed by adding cozymase. Salicylate has also been shown to inhibit malate and isocitrate dehydrogenase activities in vitro and the inhibitions were reversed by either NAD or NADP².

The results in Table 1 show that salicylate inhibits the activity of several other dehydrogenases in vitro. The inhibitions were reversed by the further addition of the appropriate coenzyme. Thus the drug appears to exert a general inhibitory action on dehydrogenase enzymes by a mechanism involving a reversible competition with the pyridine nucleotides required as co-factors. An exception is glutamate dehydrogenase, which is inhibited by salicylate by a mechanism not involving competition with either NAD or glutamate³.

Table 1. INHIBITION OF DEHYDROGENASES BY SALICYLATE

| Dehydrogenase | Coenzyme | Salicylate concentration (mM) | Inhibition (per cent) |
|----------------------------|-------------|-------------------------------------|--------------------------|
| Glucose-6-phosphate | NADP+ | 15 | 20 |
| Glyceraldchyde-3-phosphate | NAD+ | 10 | 29 |
| a-Glycerophosphate | NADH | 20 | 34 |
| Lactate | ∫NAD+ | 10 | 30 |
| Lactate | NADH | 20 | 37 |
| Alcohol | NAD+ | 10 | 19 |
| Malate (ref. 2) | NAD+ | 5 | 40 |
| Isocitrate (ref. 2) | NADP+ | 5 | 20 |

The effects of salicylate on the transfer of radiocarbon from labelled substrates into metabolic intermediates suggest that the drug inhibits dehydrogenase activities in more complex biological systems. The increased formation of radioactive malate and citrate in rat mitochondria incubated with (1 : 4-14C2) succinate in the presence of salicylate is explicable in terms of a direct inhibitory action of the drug on the mitochondrial malate and isocitrate dehydrogenases². Further work is in progress to determine if the in vitro inhibitory effects of salicylate on the other dehydrogenase enzymes contained in Table 1 may also be demonstrated in preparations of animal tissues.

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⁵ Euler, H. Von, and Ahlstrom, L., *Hoppe-Seyl. Z.*, 279, 175 (1943).
 ² Bryant, C., Smith, M. J. H., and Hines, W. J. W., *Biochem. J.*, 86, 391 (1963).

³ Gould, B. J., Huggins, A. K., and Smith, M. J. H., Biochem. J., 88, 346 (1963).

2,6,10,14-Tetramethylpentadecanoic Acid, a Constituent of Butterfat

In earlier work in this laboratory, on the trace and minor fatty acid constituents of butterfat, a C20 multibranched chain fatty acid was isolated^{1,2} which possessed physical and chemical properties corresponding with an acid afterwards isolated and conclusively identified by Bjurstam et al.³ and by Sonneveld et al.⁴ as 3,7,11,14tetramethylhexadecanoic acid. Continued investigations on the fatty acid constituents of New Zealand butterfat have now resulted in the isolation in trace amounts of a homologous C19 saturated fatty acid with similar properties and with four side-chain methyl groups. Examination of this fatty acid by techniques which included mass spectrometry, nuclear magnetic resonance, infrared spectrometry and gas-liquid chromatography has led to this previously unreported constituent being identified as 2,6,10,14-tetramethylpentadecanoic acid.

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¹ Hansen, R. P., and Shorland, F. B., Biochem. J., 50, 358 (1951).

² Hansen, R. P., and Shorland, F. B., Biochem. J., 55, 662 (1953).

^a Bjurstam, N., Hallgren, B., Ryhage, R., and Ställberg-Stenhagen, S., referred to by Stenhagen, E., Z. Anal. Chem., 181, 462 (1961).
^c Sonneveld, W., Haverkamp Begemann, P., van Beers, G. J., Keuning, R., and Schogt, J. C. M., J. Lipid Res., 3, 351 (1962).

A Reaction between Glucose and Egg White **Proteins in Incubated Eggs**

RECENT work in this laboratory¹ showed that incubation of infertile chicken eggs or separated chicken egg whites resulted in changes in the starch-gel electrophoretic We have now obtained results which show: patterns. (1) The major deteriorative changes observed on incubation of egg white, as seen in starch-gel electrophoretic patterns, can be prevented by dialysis of the egg white; the changes can be simulated by adding back glucose. (2) Glucose reacts with most of the proteins. (3) The reaction occurs similarly in the egg whites of the three species investigated in addition to chicken egg white.

The deteriorative changes previously seen in chicken egg white¹ were also found in the starch-gel electrophoretic patterns of egg whites from Japanese rice quail eggs incubated at 37° C and in the separated and blended egg whites of Japanese rice quail, Peking duck, and guinea fowl incubated at 40° -44° C as shown in Fig. 1A.B. Horizontal starch-gel electrophoresis was performed with