Table 2. BIOCHEMICAL DIFFERENTIATION OF P. mors-prunorum AND

 PEAR ISOLATES OF P. syringae

Species	Æsculin and arbutin hydro- lysis	Gelatin lique- faction	Casein hydro- lysis	Tyrosin- ase activity	Growth in nutrient broth + 5 per cent sucrose
Pmors- prunorum (a) Cherry					
strains	-		-	+	white
(b) Plum strains P. syringae	- +*	-+	+	+	white yellow

* One isolate failed to hydrolyse æsculin and arbutin.

without passage through the host and appear, therefore, to be relatively stable in vitro. The difference between the plum and cherry strains may merely reflect a different history of phage-infection in the The cherry strains, for example, may have field. failed to acquire resistance to A2 and A7 or related phages because these are absent on cherry or their activity is in some way inhibited. Alternatively, the phage types may be a result of selection by the host plants. This would imply some form of association between phage sensitivity and specific virulence factors in *P. mors-prunorum*.

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¹ Crosse, J. E., Trans. Brit. Mycol. Soc., 36, 38 (1953). ² Crosse, J. E., and Hingorani, M. K., Nature, 181, 60 (1958).

Serotonin in the Small Intestine of Conventional and Germ-free Chicks

THE function of serotonin (5-hydroxytryptamine) in the tissues of the small intestine is not understood even though it is found in relatively large quantities, particularly in the enterochromaffin cells¹. The most frequently repeated interpretation ascribes control of intestinal motility and vascular permeability to serotonin^{2,3}. Thus, if the tonus of the gut is dependent on serotonin-levels, germ-free animals, which possess poor tonus of the tract and distended cæca4, might be expected to have a lower serotonin content in the gut than conventional animals. On the contrary, however, this communication provides evidence that intestinal serotonin-levels are actually depressed by the presence of micro-organisms in the lumen of the gut.

White Leghorn chicks were raised in a germ-free environment by methods similar to those previously described⁵ on a complete starch-casein diet⁶. Fertile eggs were obtained from local hatcheries after 19 days of incubation, immersed in 2 per cent mercuric chloride solution for 10 min. at 39° C. and put into sterile 'Plexiglas' isolators where they hatched. Control chicks were treated in the same manner but were reared in a non-sterile isolator. After four weeks the chicks were decapitated; the small intestine was excised and washed in cold water. All germ-free animals were examined⁷ for bacterial contamination when killed. The various segments of the intestine were homogenized in 0.1 N hydrochloric acid in a Ten Brook apparatus at 5° C., and serotonin was assayed⁸ on fresh specimens in a Turner photofluorometer (G. K. Turner and Associates, Palo Alto, California). Table 1. SEROTONIN IN THE SMALL INTESTINE OF GERM-FREE AND CONVENTIONAL (CONTAMINATED) WHITE LEGHORN CHICKS AT FOUR WEEKS OF AGE

	11 20010								
Treatment	No. of chicks		n (µgm./ enum		n. fresh tissue) Ileum				
Germ-free	20	9.0	± 1.0*	11.0	± 0.7				
Conventional	20	8.3 ± 1.3		6.1	6.1 ± 0.8				
Significance of differences by the 't' test									
	-	t	Р	t	P				
Germ-free vs. c	0.84	< 0.4	8.8	< 0.01					
* Mean ± standard deviation.									

A significantly higher concentration of serotonin was found in the ileum of germ-free chicks than in control birds (Table 1). This is consistent with the observation⁹ that rats and mice given peroral antibiotics produce significant increases of serotonin in the small intestine. However, no such difference was apparent in the duodenum (Table 1). There is insufficient evidence to determine whether these results reflect differences between the microflora of the duodenum and ileum or an influence of the intense microbial activity of the cæca^{10,11} due to the proximity of the ileum to the latter. In any event, the presence of intestinal micro-organisms clearly affects serotonin in the ileum of chicks.

Several reports suggest alternative explanations for the microbial inhibition of serotonin. A few isolates of Aerobacter aerogenes and Alcaligenes faecalis appear to be capable of oxidizing serotonin in vitro¹² although it is not known if this event occurs in the lumen of the intestine. In addition, many species of lactobacilli which are present in large numbers in the chick¹³ require pyridoxine or its derivatives¹⁴; pyridoxal phosphate is required as a co-factor in the decarboxylation of 5-hydroxytryptophan to serotonin¹⁵. Thus bacterial removal of the coenzyme may inhibit serotonin synthesis. A similar effect might be achieved by the interaction of a bacterial product with the coenzyme such as has been reported for norepinephrine and 3,4-dihydroxyphenylalanine¹⁶.

Further work is required to determine the reason for the difference in intestinal serotonin between germ-free and conventional chicks and the physiological significance of this observation.

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- ¹ Benditt, E. P., and Wong, R. L., J. Exp. Med., 105, 509 (1957).
- ² Erspamer, V., Z. Vit. Hormon. Fermentforsch., 9, 74 (1957).
- ⁹ Page, I. H., Physiol. Rev., 38, 277 (1958).
- ¹ Gordon, H. A., Amer. J. Digest. Dis., 5, 841 (1960).
 ⁶ Gordon, H. A., Amer. J. Digest. Dis., 5, 841 (1960).
 ⁶ Phillips, A. W., Rupp, F. A., Smith, J. E., and Newcomb, H. B., in Proc. Second Symp. Gnotoiotic Techn. (Notre Dame Univ. Press, Notre Dame, Indiana, 1960).
 ⁶ Forbes, M., and Park, J. T., J. Nutrition, 67, 69 (1959).
- Wagner, M., Ann. N.Y. Acad. Sci., 78, 89 (1959).
- ⁸ Weissbach, H., Waalkes, T. P., and Udenfriend, S., J. Biol. Chem., 230, 865 (1958). ⁹ Sullivan, T. J., Brit. J. Pharmacol., 16, 90 (1961).
- ¹⁰ Lev, M., and Briggs, C. A. E., J. App. Bacteriol., 19, 224 (1956).
- ¹¹ Lev, M., Brig 364 (1957). Briggs, C. A. E., and Coates, M. E., Brit. J. Nutrition, 11,
- ¹² Sack, R. B., and Oginsky, E. L., Bacteriol. Proc., 32 (1959).
- ¹³ Wiseman, R. W., Bushnell, O. A., and Rosenberg, M. M., Poultry Sci., 35, 126 (1956).
- Spector, W. S. (ed.), Handbook of Biological Data (Saunders, Philadelphia and London, 1956).
 ¹⁵ Buzard, J. A., and Nytch, P. D., J. Biol. Chem., 229, 409 (1957).
- ¹⁶ Buzard, J. A., and Nytch, P. D., J. Biol. Chem., 234, 884 (1959).