

Increased Leptin Permeation across the Blood–Brain Barrier after Chronic Alcohol Ingestion

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Leptin, a polypeptide mainly produced in the periphery, crosses the blood–brain barrier (BBB) by receptor-mediated transport to exert multiple central nervous system actions including decreased food intake. The reciprocal interactions between leptin transport and alcohol drinking are not clear. In this study, we tested whether alcohol increases leptin entry into brain and, if this occurs, whether it is a consequence of a generalized increase in the permeability of the BBB. BBB permeability to albumin, the increased permeation of which indicates BBB disruption, as well as to leptin was measured after alcohol ingestion. CD1 and B6 mice ingested a 5% liquid alcohol diet or its isocaloric control for 2 weeks. Alcohol ingestion resulted in increased blood-alcohol levels, decreased blood-leptin concentrations, and increased permeation of radioactively labeled leptin across the BBB as shown by *in situ* perfusion. Although the increased influx of the vascular marker albumin into brain showed partial disruption of the BBB, the influx of ¹²⁵I-leptin still could be suppressed by excess unlabeled leptin, indicating persistence of its saturable transport system. When given a choice of either alcohol or control diet, even the alcohol-preferring B6 mice showed a significantly greater preference for the control liquid diet, and there was no evidence of BBB disruption or altered leptin transport. Furthermore, acute alcohol intoxication induced by intraperitoneal injection of 20% alcohol did not result in BBB disruption or increased leptin permeation 4 h later. Thus, partial disruption of the BBB and increased permeation of leptin in both CD1 and B6 mice were only induced by chronic alcohol ingestion. The results showing increased leptin permeation across the BBB lead to the speculation that leptin may serve as a homeostatic feeding signal in these mice.

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INTRODUCTION

Animal models of diet-induced alcoholism reveal a striking difference in alcohol (ethanol, EtOH) preference among different strains of mice (Vavrousek-Jakuba *et al*, 1991; Blednov *et al*, 2004). The C57BL/6J (B6) strain of alcohol-preferring mice showed the most consistently high intake of alcohol of 15 inbred mouse strains tested (Belknap *et al*, 1993). We examined whether the cardinal ingestive peptide leptin participates in the feeding behavior of alcohol preference, especially in the B6 strain.

Leptin is mainly produced by adipose tissue and can cross the blood–brain barrier (BBB) to act on the hypothalamus and other central nervous system (CNS) targets to reduce food intake (Kastin and Pan, 2006). There are complex interactions among alcohol ingestion, blood-leptin concentration, and the CNS response to leptin. Many other factors in addition to leptin can modulate alcohol ingestion, such as

methionine-enkephalin (Banks *et al*, 2006b; Urayama *et al*, 2006). Nevertheless, there is evidence that leptin participates in the metabolic disturbance associated with chronic alcoholism, shown by the effect of leptin in improving insulin resistance induced by chronic administration of high doses of alcohol in mice (Balasubramanian and Nalini, 2006).

Various studies have found an increase, decrease, or no change in blood concentrations of leptin after alcohol ingestion, depending upon test conditions. Circulating leptin levels do not correspond with the amount reaching the brain by BBB transport (Banks *et al*, 2000). The transport system is saturable and has limited capacity (Banks *et al*, 1996; Kastin *et al*, 2001). Thus, the apparent influx rate of leptin may be altered by its concentration in blood (Kastin *et al*, 1999; Banks *et al*, 2002). In this study, the direct effects of alcohol on the transport system for leptin at the BBB were determined after *in situ* perfusion in blood-free buffer.

Mild alcohol intoxication was induced in mice by a liquid diet containing 5% alcohol for 2 weeks after gradual adaptation. In contrast to this forced feeding regimen in which alcohol was the only source of energy intake, we also conducted a two-bottle choice feeding study. In addition,

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the effect of acute alcohol administration was tested to determine whether alcohol dependence is required for altered leptin influx. In this way, the experimental variables of duration of alcohol exposure, and voluntary compared with forced drinking, were examined.

MATERIALS AND METHODS

Young adult male mice (5–7-week-old) of either CD1 or B6 strains were used for the study. The animal protocol was approved by the Institutional Animal Care and Use Committee. Recombinant mouse leptin was obtained from Calbiochem (Temecula, CA). Bovine serum albumin, chloramine-T, sodium metabisulfite, and Sephadex G-10 were obtained from Sigma (St Louis, MO). ^{125}I was purchased from Amersham-GE Sciences (Piscataway, NJ) and ^{131}I was purchased from PerkinElmer (Boston, MA). Radioactive labeling (radiolabeling) and purification of ^{125}I -leptin and ^{131}I -albumin were performed by the chloramine-T method, and Sephadex G-10 columns as described previously. The 190 proof alcohol used for the feeding studies was obtained from Pharmco Products (Brookfield, CT). Powdered food to prepare the liquid diets was obtained from Bio-Serv (Frenchtown, NJ). The liquid diet offers 1000 kcal/l of calorie, with a distribution of 208 kcal/l from protein, 119 kcal/l from fat, and 673 kcal/l from carbohydrate.

In the forced alcohol-feeding studies, a group of mice 6–8-week-old were adapted to a nonalcoholic liquid diet for 3 days and then gradually adapted to an alcoholic liquid diet starting with 1.67% alcohol for 3 days, followed by 3.94% alcohol for another 3 days. The mice were then given a 5% alcohol-containing diet for 14 days. The control group was fed an isocaloric liquid diet. Study of both B6 and CD1 mice not only provides new data on strain differences, but it also facilitates comparison with previous reports as most leptin transport assays have been conducted on CD1 mice.

In the two-bottle pair-feeding studies, a 5% alcohol and a control isocaloric liquid diet were freely available to mice for 14 days after the 3 days initial adaptation period to liquid diet. To control for place preference, the alcohol and water bottles were alternated every 2 days. Alcohol and water consumption and body weight were monitored regularly. Control cages without mice were included in the study to exclude errors caused by unwanted motion or leakage. Only B6 mice were used for this study, as preliminary data showed that CD1 mice have an even lower preference for alcohol.

The acute alcohol model was generated by intraperitoneal (i.p.) injection of 20% alcohol (2 g/kg of body weight) 4 h before the BBB transport assay. The control group received vehicle (0.9% NaCl) injected at the same time ($n = 6/\text{group}$). This study design involved both B6 and CD1 mice to test for potential differences in their BBB transport of leptin to the influence of alcohol.

No significant adverse effects of alcohol ingestion, such as hypothermia, failure to thrive, unusual body tremor, fur muffling, or seizures, were observed in any of the studies. However, discoordination in beam walking and altered locomotor activity were noticed in mice 2 weeks after forced

alcohol feeding or within the first hour of i.p. injection of alcohol.

At the end of the feeding studies or after i.p. injection of alcohol, the mice were anesthetized by i.p. injection of a mixture of ketamine/xylazine/acepromazine and used either for measurement of blood concentrations of alcohol and leptin or for transport measurements. To avoid withdrawal, alcohol feed was present until the mice were anesthetized, and they were studied within 10 min of anesthesia.

The concentration of leptin in serum was measured by a quantitative sandwich enzyme immunoassay (MOB00, R & D Systems, Minneapolis, MN). The assay sensitivity was 22 pg/ml. The concentration of alcohol was measured by an alcohol dehydrogenase/reduced nicotinamide dinucleotide assay (Sigma) in serum stored in tightly sealed vials at -80°C until use. A standard solution of alcohol was used to generate the standard curve. Assays of leptin and alcohol concentrations involved samples obtained in different mice from those used for BBB transport assays, but were age- and sex-matched and fed at the same time.

Permeability of the BBB to the vascular marker albumin and leptin was determined by *in situ* brain perfusion of radiotracers as described previously (Pan *et al*, 1998). Both a single-time point study design (5 min of perfusion with radiotracers) and a multiple-time regression study design were used. In brief, 1% bovine serum albumin was added to the oxygenated perfusion buffer and the pH adjusted to 7.4. The perfusion was driven by a syringe pump at a speed of 2 ml/min. Each anesthetized mouse was transcardially perfused with buffer for 2 min to remove blood from the vasculature. This was followed by perfusion containing both ^{125}I -leptin and ^{131}I -albumin (0.5 $\mu\text{Ci}/\text{ml}$ of each) for the designated duration, after which the mice were again perfused with buffer for 1 min to remove any radiotracers remaining in the cerebral blood vessels. Two types of statistical analyses were performed: comparison of uptake at a single time point (mean \pm SE) by analysis of variance, and comparison of the difference in the influx rate (K_i) by multiple-time regression studies. The analyses were performed with SPSS and GraphPad Prism Statistical Analysis software.

To test the saturability of leptin influx at the end of the alcohol feeding, excess unlabeled leptin was included either in the *in situ* brain perfusion buffer or in the bolus intravenous (i.v.) injection, as specified in the Results. ^{125}I -leptin was injected i.v. into anesthetized mice in a bolus of 100 μl of lactated Ringer's solution containing 1% bovine serum albumin through the jugular vein at time 0. The mice were decapitated at various times, 1–20 min after i.v. injection. The linear regression correlation between the brain/blood ratio and the exposure time was used to calculate the influx rate K_i as described previously. In this assay, groups of mice (at least 7/group) were studied, each representing a different time point. The K_i values were compared by analysis of variance to determine whether there were differences within groups. This was followed by Newman–Keul's *post hoc* test to determine which values within a group differed. The standard deviation of the mean for the slope was taken as the standard error of the mean and, because two means (the slope and the intercept) were calculated from the data, $n-1$ was used as the n -value in the analysis of variance and range tests. Significant differences

between groups were determined with the aid of the GraphPad Prism statistical program (Kastin and Pan, 2003; Pan *et al*, 2004).

RESULTS

Feeding Behavior and Blood Concentrations of Alcohol and Leptin in Mice on a Forced Alcohol Diet

After 9 days of adaptation to the liquid diet and gradual increase of alcohol concentration, groups of mice ($n = 12-14/\text{group}$) were fed either a 5% alcohol diet or an isocaloric nonalcohol diet for 2 weeks. Despite the 9-day adaptation period, both B6 and CD1 mice showed reduced alcohol intake when the alcohol concentration was increased to 5%. Although the formula for the alcohol and control liquid diet groups contained similar composition and the same energy supply (kcal/ml), the alcohol groups drank less and thus had lower daily caloric intake. This reduced alcohol intake persisted for the entire 14 days in B6 mice, but only lasted a week in the CD1 mice (Figure 1a). Correlating with the reduced intake of liquid food, there was a significant reduction in body weight that was more pronounced in B6 mice than in CD1 mice (which weighed more than the age-matched B6 mice even in the control groups) (Figure 1b). The stepwise increase of blood-alcohol concentration was apparent in CD1 mice, as shown in the line graph (Figure 1c). The B6 mice, studied at a different time, appeared to show higher blood-alcohol concentrations after 14 days, which dropped precipitously 2 days after withdrawal (Figure 1c, inset).

There was no significance difference in water consumption in groups of mice fed with control diet or alcohol (Figure 1d). Nonetheless, concurrent with the significant increase of alcohol levels was a significant decrease in blood-leptin concentrations when measured at day 14 (Figure 1e). These lower blood-leptin concentrations were consistent with the lower body weights of the alcohol groups compared with the control groups fed an isocaloric diet. Thus, alcohol feeding significantly increased blood-alcohol and decreased blood-leptin concentrations in both strains of mice ($P < 0.001$).

The Effect of Forced Alcohol Diet on the Blood-to-Brain Influx of Albumin (Vascular Marker) and Leptin (Polypeptide with Specific Transport System) in Nonalcohol-Preferring CD1 Mice

As the groups of mice fed alcohol had significant differences in body weight and blood-leptin concentrations from the control groups, the potential use of i.v. injection to evaluate BBB permeability could be confounded by peripheral binding proteins and endogenous leptin levels. Therefore, *in situ* brain perfusion was performed to test the regression correlation between the brain/perfusate ratio and perfusion time (1–10 min). The influx rate of ^{125}I -leptin in CD1 mice was $0.47 \pm 0.18 \mu\text{l/gmin}$ in the control group ($n = 8$) and $1.24 \pm 0.26 \mu\text{l/gmin}$ in the alcohol group ($n = 7$). The difference between the groups was significant ($P < 0.05$). However, the influx of ^{131}I -albumin ($K_i = 0.47 \pm 0.16 \mu\text{l/gmin}$) in the alcohol group tended ($P = 0.055$) to be higher than in the control group ($0.03 \pm 0.13 \mu\text{l/gmin}$). After

subtraction of the vascular space to normalize the actual entry of leptin, the influx rate was $0.37 \pm 0.17 \mu\text{l/gmin}$ for the control group and $0.76 \pm 0.26 \mu\text{l/gmin}$ for the alcohol group. The difference between the two groups was not significant. Thus, alcohol feeding increased leptin influx, and this increase was associated with a partial disruption of the BBB as shown by a trend toward increased albumin influx (Figure 2a).

Despite an increase in the general permeability of the BBB to albumin associated with the influx of leptin, as shown above, the saturable transport system for leptin was still functional. This was evident after i.v. injection and multiple-time regression analysis in alcohol-fed mice receiving the ^{125}I -leptin tracer in the presence or absence of excess unlabeled leptin. As shown in Figure 2b, excess leptin ($1 \mu\text{g}/\text{mouse}$) significantly decreased the uptake of ^{125}I -leptin ($n = 9/\text{group}$), showing saturable transport.

The Effect of Forced Alcohol Diet on the Blood-to-Brain Influx of Albumin (Vascular Marker) and Leptin (Polypeptide with Specific Transport System) in Alcohol-Preferring B6 Mice

In situ brain perfusion was performed in groups of B6 mice 2 weeks after feeding of the alcohol or control liquid diet. Like the CD1 mice, the influx rates of both ^{125}I -leptin and ^{131}I -albumin in the B6 mice were higher in the alcohol-fed group. The value of [^{125}I -leptin- ^{131}I -albumin] was calculated to determine the amount of leptin completely crossing the BBB and taken up in the brain parenchyma by selective entry, without interference from the vascular space. The alcohol group had a corrected K_i of $0.31 \pm 0.07 \mu\text{l/gmin}$ ($n = 11$), whereas that in the control group was $0.15 \pm 0.04 \mu\text{l/gmin}$ ($n = 13$). The difference between the groups was significant ($P < 0.05$) (Figure 3a). As the BBB was partially disrupted, this differential permeability shows that the increase of leptin influx exceeded that of albumin influx, suggesting an upregulated transport system component to the increased leptin influx. Comparison of the K_i of leptin in the nonalcohol B6 and CD1 mice showed that the inbred B6 mice ($n = 10$) had a lower K_i than the outbred CD1 mice ($n = 6$) (Figure 3b). Regardless, the increase of leptin influx in the B6 mice after alcohol exceeded that of barrier disruption, probably reflecting upregulation of the transport system.

Paired Feeding Study Showed that a Lower Level of Alcohol Drinking did not Cause BBB Disruption or Increased Influx of Leptin

Two identical bottles of control diet were provided to the B6 mice, and spatial bias (preference for drinking from a particular location) was avoided by rotation of the bottles of food and water every other day. The 14-day study, the same duration as in the forced alcohol study, began when one of the two bottles in the randomly chosen alcohol group was switched to 5% alcohol at day 0. As shown in Figure 4a, on day 0, all mice were fed the control diet. The first preference for the alcohol or control diet was recorded on day 2. Thereafter, the mice in the alcohol-fed group showed significantly greater ($P < 0.005$) intake of the control liquid diet than the alcohol liquid diet on each study day. At the

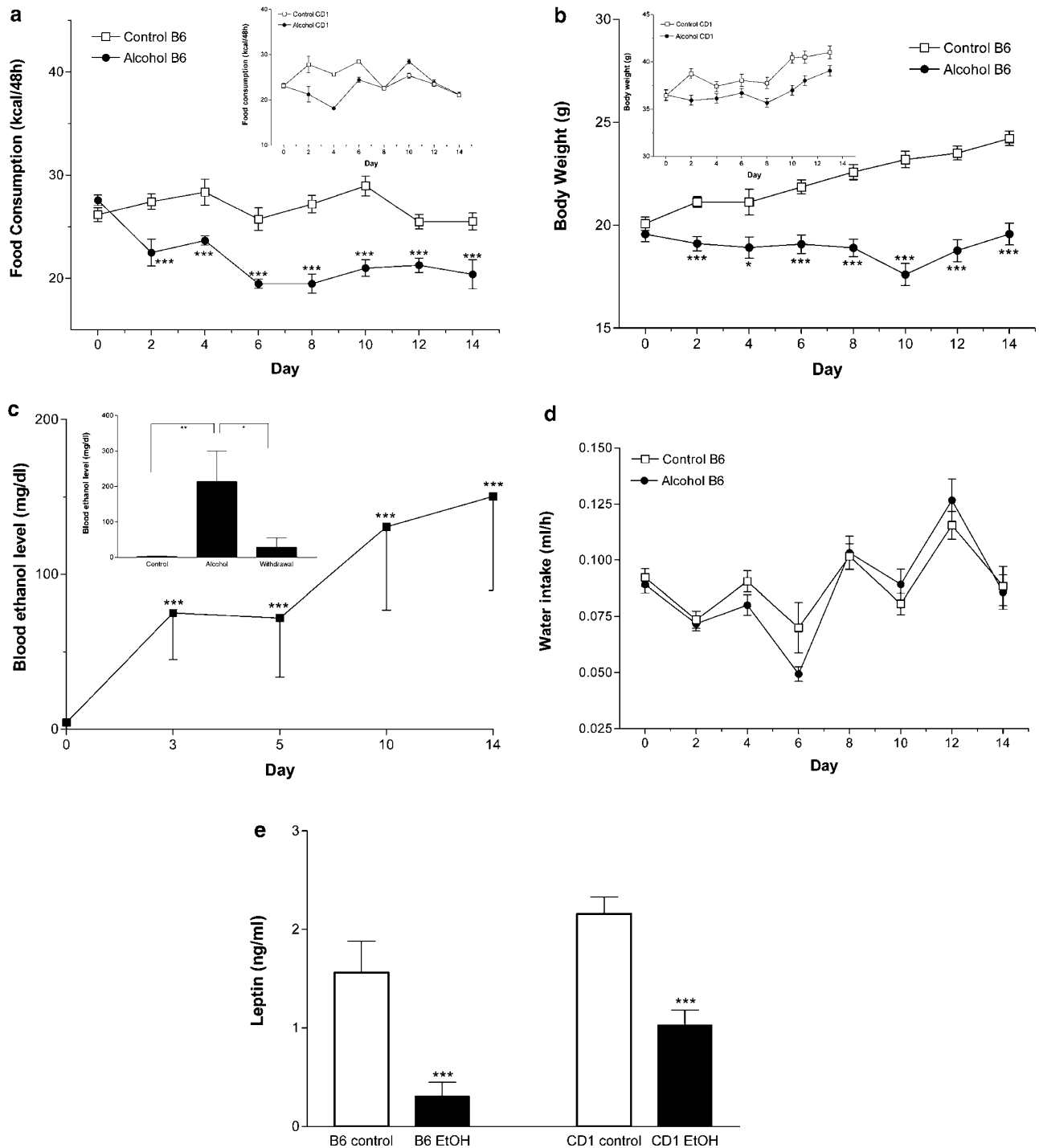


Figure 1 Effects of alcohol diet on food and water consumption, weight gain, blood-alcohol, and -leptin levels. (a) A significant decrease of food consumption was seen in alcohol-fed mice as compared with the control-fed mice. The study with B6 mice used 12–14 mice/group, and that with CD1 mice used 7/group (inset). Measurements were taken every other day. $***P < 0.005$. (b) The alcohol diet was associated with significantly lower body weights for both B6 and CD1 (inset) mice as compared with those fed the isocaloric control diet. Measurements were taken every other day ($n = 7$ –14/group). $*P < 0.05$; $***P < 0.005$. (c) In CD1 mice fed the alcohol diet, blood-alcohol concentrations showed a stepwise increase over the course of 2 weeks ($n = 5$ –10/time point). In B6 mice studied with the same diet, the alcohol levels were even higher but decreased rapidly after 2 days of withdrawal (inset). $*P < 0.05$; $**P < 0.01$; $***P < 0.005$ compared with the baseline. (d) Water consumption did not show differences in groups of mice fed with the control diet or alcohol diet. Measurements were taken every other day ($n = 14$ /group). (e) Blood-leptin concentrations measured at day 14 in mice on a forced alcohol diet showed a significant decrease ($***P < 0.005$) compared with the control groups in both B6 and CD1 strains of mice ($n = 6$ –8/group).

end of the study, the weight gain in the alcohol group (offered both control and alcohol diets) was similar to that in the control groups (offered two identical bottles of control diet) (Figure 4b). BBB transport assays at the end of

the study showed that the uptake of leptin and albumin was not significantly different between the two groups (Figure 4c). Because of anticipated large individual variations in alcohol drinking, 7 mice were included for each of

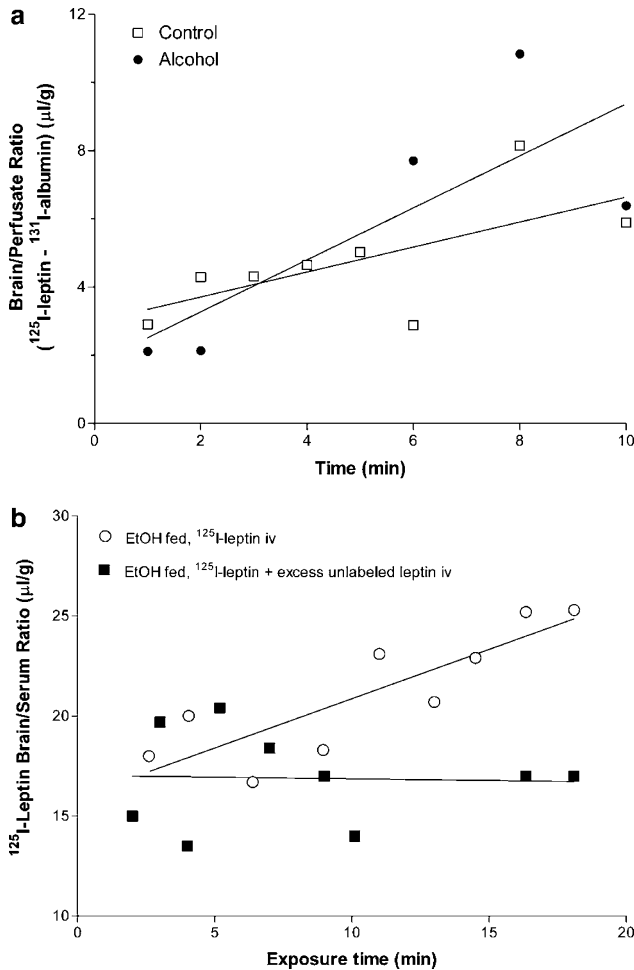


Figure 2 Effects of alcohol feeding on the BBB permeability in CD1 mice. (a) *In situ* brain perfusion showed that nonalcohol-preferring CD1 mice receiving a forced alcohol diet had a significantly ($P < 0.05$) higher influx rate of ^{125}I -leptin and a trend ($P = 0.055$) toward increased ^{131}I -albumin that was injected simultaneously ($n = 7$ – 8 /group). (b) After *in situ* injection of ^{125}I -leptin, its influx rate from blood to brain was significantly ($P < 0.05$) decreased by addition of excess unlabeled leptin in CD1 mice on the forced alcohol diet for 14 days, showing that saturability of the BBB transport system remained ($n = 9$ /group).

the control diet groups and 12 mice were used for each of the alcohol groups in this study.

Acute Alcohol Administration did not Modulate BBB Permeability to Leptin Nor did it Disrupt the BBB

Groups of mice were studied 4 h after *i.p.* injection of 20% alcohol (2 g/kg) or normal saline control ($n = 6$ /group). In each group, the brain/perfusate ratio of ^{125}I -leptin and ^{131}I -albumin after 5 min of *in situ* brain perfusion was calculated. In neither CD1 nor B6 mice, was there any significant change in the brain uptake of albumin (indicative of vascular space and potential BBB disruption) or leptin (polypeptide crossing the BBB with a specific transport system) (Figure 5). Studies of the CD1 and B6 mice were conducted on different days with different batches of radioisotopes.

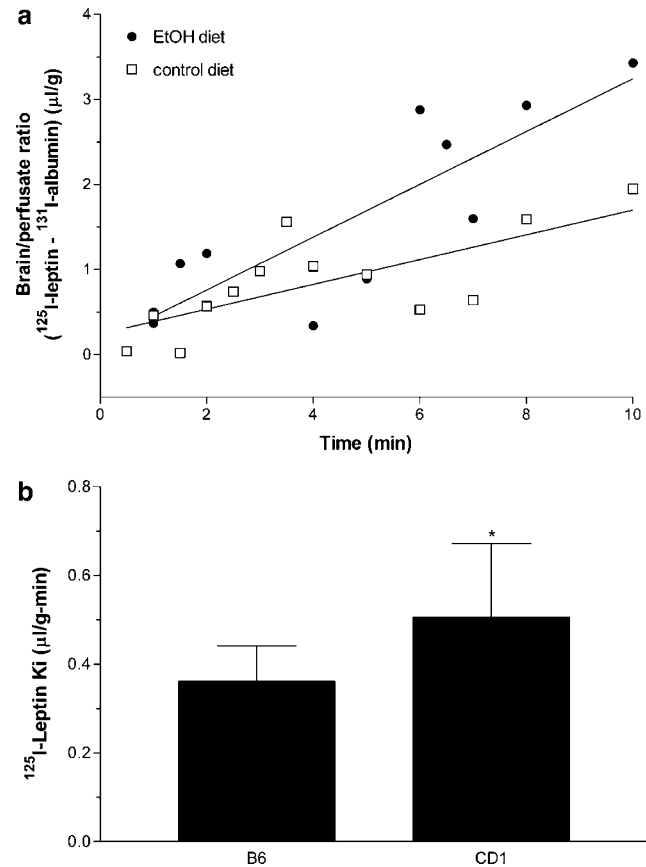


Figure 3 Effects of alcohol diet on BBB permeability of the B6 mice, which had a lower basal influx rate for leptin. (a) *In situ* brain perfusion showed that alcohol-preferring B6 mice fed a forced alcohol diet for 14 days had a significantly ($n = 11$, $P < 0.05$) higher ^{125}I -leptin influx (corrected by subtraction of albumin influx) than mice fed the isocaloric control diet ($n = 13$). (b) Comparison from different studies showed that CD1 mice had a higher K_i for leptin across the BBB than the B6 mice. Results were derived from six independent studies. * $P < 0.025$.

DISCUSSION

Alcohol readily crosses the BBB because of its high lipid solubility (Raichle *et al* 1976). However, the effects of alcohol on the permeability of the BBB in adult mammals have been inconsistent. Earlier studies, mostly in the 1980's, showed three types of results: increased permeability with evidence of structural damage (Karwacka, 1980; Gulati *et al*, 1985; Stewart *et al*, 1988; Borisenko, 1990); no change induced by alcohol (Hemmingsen *et al*, 1980; Hillbom and Tervo, 1981; Phillips, 1981; Phillips and Cragg, 1982); and even a protective and beneficial effect (Neilly and Lin, 1986). The conflicting results are not easily reconciled by dose, duration, or other pharmacological factors, whereas a detrimental effect of alcohol on tight junction proteins and alcohol-induced oxidative stress on endothelium was evident in a cell-culture system (Haorah *et al*, 2005b).

Therefore, we further tested the relationship between leptin permeation and alcohol drinking by use of different strains of mice, different doses of alcohol, and different durations of administration. Forced alcohol feeding for 2 weeks was associated with reduced food consumption and weight loss, even in alcohol-preferring B6 mice. This is similar to clinical observations in chronic alcoholic patients.

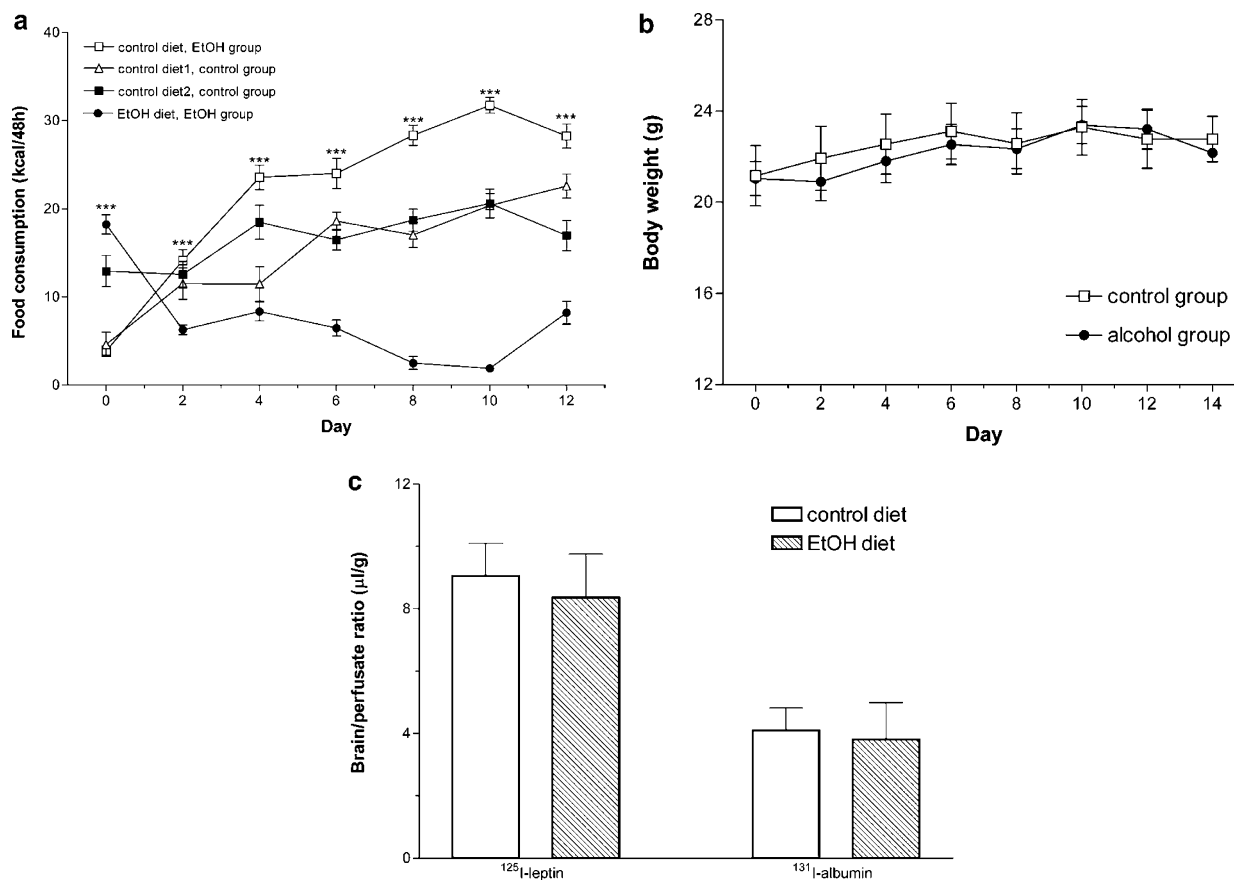


Figure 4 Effects of alcohol choice diet on food preference, body weight, and BBB permeability. (a) During the two-bottle choice study, the alcohol-prefering strain of B6 mice still preferred the control liquid diet ($n = 7$) more than the alcohol diet ($n = 12$). $***P < 0.005$. (b) The group of mice given the choice of consuming either alcohol or isocaloric liquid control diet had a similar weight gain as the group of control mice fed an isocaloric diet without alcohol. (c) Mice with the alcohol choice feeding showed no significant changes in leptin or albumin permeation across the BBB after 14 days.

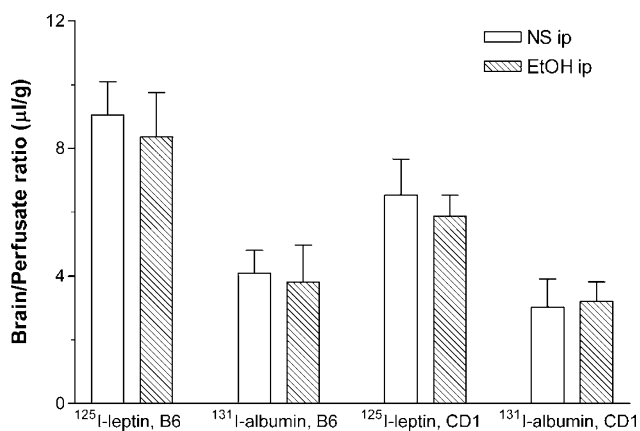


Figure 5 Acute treatment with alcohol (20% at 2 g/kg) vs normal saline (NS) by i.p. injection did not cause a significant change in the brain uptake of ^{125}I -leptin or ^{131}I -albumin. The radiotracers were administered simultaneously for 5 min by *in situ* brain perfusion in either B6 or CD1 mice ($n = 6/\text{group}$).

Although we cannot rule out taste aversive effects, the blood concentration of alcohol reached a level considered as intoxication, indicating that the chronic ingestion model was successful. Correlated with the lower body weight, the alcohol group had significantly decreased blood-leptin concentrations.

After *in situ* brain perfusion, the influx rate of ^{125}I -leptin was significantly higher in the alcohol-fed mice than the controls. This increased influx could indicate that: (1) the transport system functions at a higher capacity; (2) there was disruption of the BBB in the alcohol-fed mice; or (3) both processes are involved. By inclusion of excess unlabeled leptin to show saturability, we demonstrated that the transport system for leptin persisted in the alcohol-fed mice. By including simultaneous administration of the vascular marker ^{131}I -albumin in the studies, we also showed partial BBB disruption in mice when alcohol was the main supply of energy intake for 2 weeks. The inert protein albumin serves as a permeability marker as its basal penetration across the intact BBB is low. With these studies, we showed that the effects of alcohol were dose- and time-dependent, as increased BBB permeation was not seen in mice provided with two-bottle choice feeding for the same duration, or in mice receiving a large dose of alcohol 4 h before the end of the study. Thus, coexisting BBB disruption and persistent leptin transport contributed to the overall influx of leptin in alcohol-ingesting mice.

In support of the observation that alcohol increased the permeability of the BBB, electron microscopic studies of cerebral capillary endothelia after chronic alcohol administration showed increased micropinocytic vesicles, mitochondria, smooth endoplasmic reticulum, and the Golgi apparatus, whereas the astrocytic processes reinforcing the

BBB showed edema and abnormal mitochondria (Karwacka, 1980). Partial disruption of the BBB and increased nonspecific paracellular permeability are also consistent with the observation *in vitro* that alcohol treatment activates myosin light chain kinase, alters the distribution and function of the tight junction proteins occludin and claudin-5, and reduces transendothelial electrical resistance (Haorah et al, 2005a).

Both CD1 wild-type and B6 inbred mice adjusted to the diet smoothly during the adaptation phase involving a gradual increase of alcohol to 5% over 9 days. The blood concentration of leptin was lower in the B6 mice, and further reduction after alcohol ingestion was more pronounced in the B6 mice. The reduced serum leptin in the alcohol-ingesting mice was consistent with their lower body weight. A correlation between body weight and blood-leptin concentration has been well documented (McGregor et al, 1996). As both strains of mice showed increased uptake of leptin after ingestion of alcohol, the relatively smaller amount of leptin available to the brain in B6 mice might partially explain their reported higher tolerance to alcohol. Regardless, such preference appeared to be rather moderate, as B6 mice preferred to consume more of the control diet than the alcohol diet in the choice feeding study. The decreased serum leptin concentrations are probably explained by a lower production of leptin in the periphery, as the mice had slower weight gain than the controls. This is consistent with our observation from the transport assays that leptin permeation probably results from a combination of partial disruption of the BBB and persistent saturable transport. Transport of leptin across the BBB is unlikely to be the reason for the decreased serum leptin.

To determine whether the effect of alcohol on BBB permeability was immediate and direct or mediated by chronic adaptation, we tested the permeation of the BBB to albumin and leptin 4 h after alcohol administration by the i.p. route, an effective method to test acute alcohol actions (Redetzki et al, 1983; Broadbent et al, 2005). The lack of significant increase in the uptake of leptin and albumin within this short time suggests that the adaptive changes of the BBB might require a higher dose or longer duration of exposure.

The greater increased brain influx of leptin than albumin indicates the differential permeability of a partially disrupted BBB. The normalized influx rate of [¹²⁵I-leptin-¹³¹I-albumin] was significantly higher in mice after chronic alcohol ingestion. It suggests that both barrier disruption and an upregulated transport system were involved in the influx of leptin from blood to brain. Similar regulatory changes have been seen with other polypeptide cytokines in mouse models of autoimmune encephalomyelitis (Pan et al, 1996), brain and spinal cord injury (Pan et al, 1997, 2003, 2006a), and stroke (Pan et al, 2006b). As the blood concentration of leptin was significantly reduced in mice with chronic alcohol intoxication, the greater permeability of the BBB to leptin may serve to prevent further reduction of brain leptin concentrations to a pre-morbid level. Banks et al (1996, 2006a) has elegantly demonstrated the relevance of rodent studies to primates in terms of leptin transport (Farr et al, 2006; Rolland et al, 2006). Although the higher leptin influx may not improve the wasting syndrome seen in chronic alcoholism, homeostasis of the CNS feeding signal

would be better maintained by such regulatory changes at the BBB.

In summary, increased leptin permeation across the BBB occurred concomitantly with BBB disruption after 2 weeks of forced alcohol ingestion. This increase was not seen when the mice had a choice of a nonalcohol diet or when alcohol was given acutely as a single high dose. Thus, increased BBB permeability to leptin was a consequence of prolonged alcohol intake. The mechanisms involved include both BBB disruption and the persistence of a specific transport system.

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DISCLOSURE/CONFLICT OF INTEREST

The authors declare that, except for income received from my primary employer, no financial support or compensation has been received from any individual or corporate entity over the past 3 years for research or professional service and there are no personal financial holdings that could be perceived as constituting a potential conflict of interest.

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