

Behavioral Sensitization, Alternative Splicing, and D3 Dopamine Receptor-Mediated Inhibitory Function

Neil M Richtand*^{1,2}

¹ Psychiatry Service (VI 16A), Cincinnati Veterans Affairs Medical Center, Cincinnati, OH, USA; ²Department of Psychiatry, University of Cincinnati College of Medicine, Cincinnati, OH, USA

Behavioral sensitization, the progressive and enduring augmentation of certain behaviors following repetitive drug use, alters rodent locomotion in a long-standing manner. The same dopamine pathways playing an important role in drug dependence and psychosis also play a critical role in sensitization. Individual dopamine receptor subtypes have markedly different functional responses to stimulation, with D3 dopamine receptor stimulation inhibiting rodent locomotion. The D3 receptor has highest affinity of the dopamine receptor subtypes for dopamine, and is occupied to a greater degree following stimulant drug administration. D3 receptor activity may be regulated through the expression of an alternatively spliced, truncated receptor isoform (termed 'D3nf') altering receptor localization and function via dimerization with the full-length subunit. The expected physiological response to repetitive drug administration is tolerance. Tolerance of D3 receptor inhibition of locomotion would contribute to sensitization to stimulant drugs. We hypothesize that repetitive D3 receptor stimulation contributes to the development of behavioral sensitization through decreased responsivity of D3-receptor-mediated locomotor inhibition. Increased D3nf expression may direct altered receptor localization and subsequent release of D3-receptor-mediated inhibition, contributing to the expression of sensitization. These hypotheses follow directly from the affinities of the receptor subtypes for dopamine; dopamine concentrations following stimulant administration; the effects of individual dopamine receptor subtype stimulation on locomotion; and the expected homeostatic response of the system to perturbation by drug. Clarifying these mechanisms underlying sensitization may suggest new interventions for neuropsychiatric conditions in which dopamine plays an important role, including psychosis, drug dependence, and Parkinson's disease. This information may also elucidate a previously unrecognized mechanism regulating receptor trafficking and desensitization.

Neuropsychopharmacology (2006) **31**, 2368–2375. doi:10.1038/sj.npp.1301163; published online 19 July 2006

Keywords: dopamine; D3nf; amphetamine; locomotor activity; animal model; drug dependence

INTRODUCTION

Although David Segal has sadly passed away, he leaves as one legacy to those who were fortunate to have had the opportunity to know and admire him, an enduring interest in the paradox of behavioral sensitization. Behavioral sensitization, the progressive and long-lasting augmentation of certain behaviors following repetitive stimulant drug administration (Segal and Mandell, 1974), is a well-studied model of behavioral plasticity. Following repeated, intermittent treatment with stimulant drugs such as amphetamine (AMPH) or cocaine, sensitized behaviors may occur more intensely, with shorter latency, and/or at lower dose than before sensitization (Segal *et al*, 1980). The paradox of

sensitization is that the expected response of a biological system to repetitive stimulation is tolerance, rather than augmentation of the behavioral response. David's view of this was that it captured the essence of science: that both the work, and the joy, of science begin with the unexpected result, in determining a context in which the unexpected finding fits logically and make sense with the other pieces of the puzzle, that is, in making the paradox unparadoxical. Here we describe a theoretical model of behavioral sensitization, which attempts to do that, by viewing sensitization as tolerance of inhibitory systems, rather than an increase following repeated drug exposure. Evidence for the role of the dopamine D3 receptor in this adaptation is presented, and RNA alternative splicing is suggested as a possible underlying mechanism.

DEVELOPMENT VS EXPRESSION OF BEHAVIORAL SENSITIZATION

Behavioral sensitization may be separated into two distinct components, both temporally and spatially, termed 'development' (or 'initiation'), and 'expression.' Initiation of

*Correspondence: Dr NM Richtand, Department of Psychiatry, University of Cincinnati College of Medicine, 231 Albert Sabin Way, ML0559, Cincinnati, OH 45267, USA, Tel: +1 513 558 6657, Fax: +1 513 558 0042, E-mail: neil.richtand@uc.edu

Received 13 January 2006; revised 31 March 2006; accepted 4 April 2006

Online publication: 9 June 2006 at <http://www.acnp.org/citations/Npp060906060026/default.pdf>

sensitization involves both D1 dopamine (DA) receptor and glutamate receptor activation within ventral tegmentum and substantia nigra (Bjijou *et al*, 1996; Wolf *et al*, 1995; Kalivas and Alesdatter, 1993). Projections from prefrontal cortex and amygdala also participate in the initiation of sensitization (Wolf, 1998). Expression of sensitization, in contrast, is believed to result from changes in the nucleus accumbens. In general, changes in the ventral tegmentum believed to be related to the initiation of sensitization are transient, such as decreased sensitivity of somatodendritic D2 subtype dopamine autoreceptors (Kamata and Rebec, 1984; White and Wang, 1984; Henry *et al*, 1989; Ackerman and White, 1990). In contrast, neurochemical changes in the nucleus accumbens accompanying sensitization are persistent (Wolf, 1998), such as increased D1 receptor electrophysiological responsiveness (Henry and White, 1991; Higashi *et al*, 1989) and upregulation of cAMP signal transduction (Nestler *et al*, 1990; Terwilliger *et al*, 1991). Some of the changes in the nucleus accumbens, such as augmentation of the increase in extracellular dopamine elicited by stimulants (Robinson *et al*, 1988; Kalivas and Duffy, 1990), require a withdrawal time interval to be manifest. The augmentation in behavioral response is an enduring alteration, persisting for at least 1 year in rats (Paulson *et al*, 1991). Evidence from several labs suggests sensitization is dependent on new protein synthesis (Karler *et al*, 1993; Sorg and Ulibarri, 1995).

BEHAVIORAL CHARACTERISTICS OF SENSITIZATION

In humans, sensitized behaviors following repetitive stimulant drug administration frequently include psychotic symptoms of paranoia, ideas of reference, and auditory and visual hallucinations in otherwise healthy individuals (Ellinwood, 1967; Griffith *et al*, 1972; Utena, 1966; Sato *et al*, 1983). Human symptoms following repetitive stimulant drug use evolve gradually from intense curiosity, progressing to intense exploration of the environment, which may be displayed in repetitive stereotyped searching, sorting, and examining behaviors. This curious 'suspiciousness' of the environment later evolves into paranoia and psychotic thought (Ellinwood *et al*, 1973; Segal *et al*, 1981; Segal and Schuckit, 1983). Because pretreatment with AMPH facilitates the later acquisition of drug self-administration in rodents (Piazza *et al*, 1989), it has also been theorized that sensitization may underlie the development of drug craving, and thus initiate addictive behaviors seen in human drug dependence (Robinson and Berridge, 1993).

Specific behaviors sensitized in rodents depend upon drug dose, with stimulant-induced locomotion enhanced following sensitization with low doses of stimulant drug. At higher drug doses, sensitization of stereotyped behaviors emerges. The initial increase in exploratory locomotion, and later repetitive-focused stereotyped movements observed in rats with progressively increasing drug dose, is in some respects analogous to the evolution of the human behavioral pattern of response to stimulants. Although this description highlights common features shared by rodent behavioral sensitization and the development of psychosis in man (Segal and Mandell, 1974; Segal *et al*, 1981; Bell, 1973; Segal and Schuckit, 1983; Sato *et al*, 1983; Lieberman *et al*, 1997),

the rodent brain and behavioral repertoire is far more limited and less complex than its human counterpart, and there is no direct correspondence between rodent behavioral sensitization and human psychiatric disease. There is, however, strong evidence that behavioral sensitization is related in large part to plasticity of limbic neurochemical systems, which also play an important role in a range of psychiatric and substance use disorders. Thus primitive, highly conserved structures modulating emotional homeostasis are likely shared by systems contributing to behavioral sensitization, psychosis, and drug dependence.

DOPAMINE AND BEHAVIORAL SENSITIZATION

Brain DA systems play an important role in behavioral sensitization, as demonstrated in lesion studies (Creese and Iversen, 1975; Creese and Iversen, 1972; Fog *et al*, 1970; Roberts *et al*, 1975; Pijnenburg *et al*, 1975; Costall and Naylor, 1974), and microinjection studies of both dopamine agonists (Hitzemann *et al*, 1980; Kalivas and Weber, 1988; Vezina and Stewart, 1990) and antagonists (Stewart and Vezina, 1989). Dopamine-mediated effects occur through interaction with two pharmacologically distinct receptor families, D1 family (including D1 and D5 receptors) and D2 family (D2, D3, and D4 receptors).

D3 dopamine receptor mRNA and protein expression are limited primarily to olfactory tubercle, nucleus accumbens, and islands of Calleja (located ventral to the ventral pallidum and nucleus accumbens), phylogenetically ancient limbic brain regions linked to motivated and emotional behaviors (Levesque *et al*, 1992; Bouthenet *et al*, 1991; Richtand *et al*, 1995; Landwehrmeyer *et al*, 1993; Gurevich and Joyce, 1999; Levant, 1998). Protein and mRNA expression are highly colocalized, suggesting that receptor expression occurs primarily on perikarya, proximal dendrites, and short axons as opposed to long axon terminals from other brain regions (Levesque *et al*, 1992).

Because D3 receptors exhibit functional as well as physical interactions with both D1 and D2 receptors (Scarselli *et al*, 2001; Zeng *et al*, 2006), it is of interest that receptor colocalization studies indicate that D1 and D3 receptors are coexpressed in islands of Calleja, nucleus accumbens, and caudate (Schwartz *et al*, 1998; Ridray *et al*, 1998; Surmeier *et al*, 1996; Le Moine and Bloch, 1996). In human brain, most D3 mRNA-expressing cells also express D2 mRNA (Gurevich and Joyce, 1999), whereas in rodent brain, in contrast, D2 and D3 receptors appear to have predominantly complementary rather than overlapping patterns of expression (Bouthenet *et al*, 1991). In the nucleus accumbens shell, colocalized D1 and D3 receptors exert synergistic effects on substance P gene expression (Ridray *et al*, 1998; Schwartz *et al*, 1998). In contrast, D1 and D3 receptor activation in the islands of Calleja exert opposite effects on *c-fos* expression, presumably through opposing actions on adenylate cyclase activity (Ridray *et al*, 1998; Schwartz *et al*, 1998). Similarly, D2 receptors in the nucleus accumbens core and D3 receptors in the nucleus accumbens shell region play opposing roles in neurotensin expression (Diaz *et al*, 1994).

D3 receptor occupation by agonist appears to inhibit neuronal firing (Liu *et al*, 1994a). Following stimulant drug

administration, D1 and D3 receptors exert opposing effects on intracellular signaling pathways and gene expression through opposite regulatory effects on extracellular signal-regulated kinase (ERK) activity and expression of Fos family genes (Zhang *et al*, 2004). Behaviorally, although acute concurrent D1/D2 dopamine receptor activation stimulates rodent locomotion (Dalia *et al*, 1998; Dreher and Jackson, 1989), D3 receptor stimulation inhibits both novelty-stimulated locomotion (Pritchard *et al*, 2003; Xu *et al*, 1997; Menalled *et al*, 1999; Ekman *et al*, 1998; Accili *et al*, 1996) and AMPH-stimulated locomotion (Waters *et al*, 1993). One model, as presented in this paper, suggests the loss of a D3 receptor-mediated 'brake' on these behaviors would be expressed as the emergence of augmented locomotor response to novelty and AMPH in rodents, and might also be expressed as paranoid psychotic symptoms in humans.

D3 DOPAMINE RECEPTOR THEORY OF SENSITIZATION

Dopamine receptor subtypes differ widely in affinity for dopamine and the dopamine concentration range over which receptor occupancy varies and alterations in cellular signaling occur. The D3 receptor has highest affinity for dopamine, with low-affinity state $K_i = 30$ nM (Sokoloff *et al*, 1992), close to basal extracellular (3–5 nM; Kalivas and Duffy, 1993; Parsons and Justice, 1992) and synaptic dopamine concentrations (50 nM; Ross, 1991). In contrast, D1 and D2 receptor affinity for dopamine are far lower: D2 K_i (nM) = 2000 and D1 K_i (nM) = 2300 (Sokoloff *et al*, 1992). Following stimulant drug administration, dopamine concentrations are elevated for prolonged periods (Jones

et al, 1996a,b), with average concentrations of 750 nM (Zetterstrom *et al*, 1983). At that dopamine concentration, calculated low-affinity state receptor occupancy is 96% for D3 receptor, compared to 25% for D1 and 27% for D2 receptor. These differences in receptor occupancy suggest the likelihood that tolerance would develop differentially for each of the three-receptor systems, with greatest tolerance of D3-receptor-mediated inhibitory effects. The D3 dopamine receptor behavioral action as a 'brake' on locomotion suggests that tolerance of this inhibition would contribute to sensitization (Richtand *et al*, 2001a,b).

D3 DOPAMINE RECEPTOR ALTERNATIVE SPLICING

The single gene coding for the D3 receptor is organized to allow for the production of different transcripts via alternative splicing of D3 receptor heteronuclear-RNA (hn RNA or pre-mRNA) in the cell nucleus, as illustrated in Figure 1 (Griffon *et al*, 1996; Fu *et al*, 1995; Giros *et al*, 1991; Park *et al*, 1995). At least seven distinct alternatively spliced D3 receptor variants are produced, including the full-length D3 receptor (called 'D3'), and a shorter receptor isoform, D3S, lacking 21 amino acids within the third cytoplasmic loop (Fishburn *et al*, 1993). Both D3 and D3S exhibit high-affinity dopamine binding. Five additional alternatively spliced variants have also been described, which do not bind dopamine, and are believed to function instead through the regulation of receptor dimerization (Snyder *et al*, 1991). These include D3 (TM3-del) (Giros *et al*, 1991; Snyder *et al*, 1991), D3 (TM4-del) (Nagai *et al*, 1993), D3 (O2-del) (Giros *et al*, 1991), rD3in (Pagliusi *et al*, 1993), and D3nf (Schmauss *et al*, 1993; Liu *et al*, 1994b). D3nf is the best characterized of the non-dopamine-binding splice

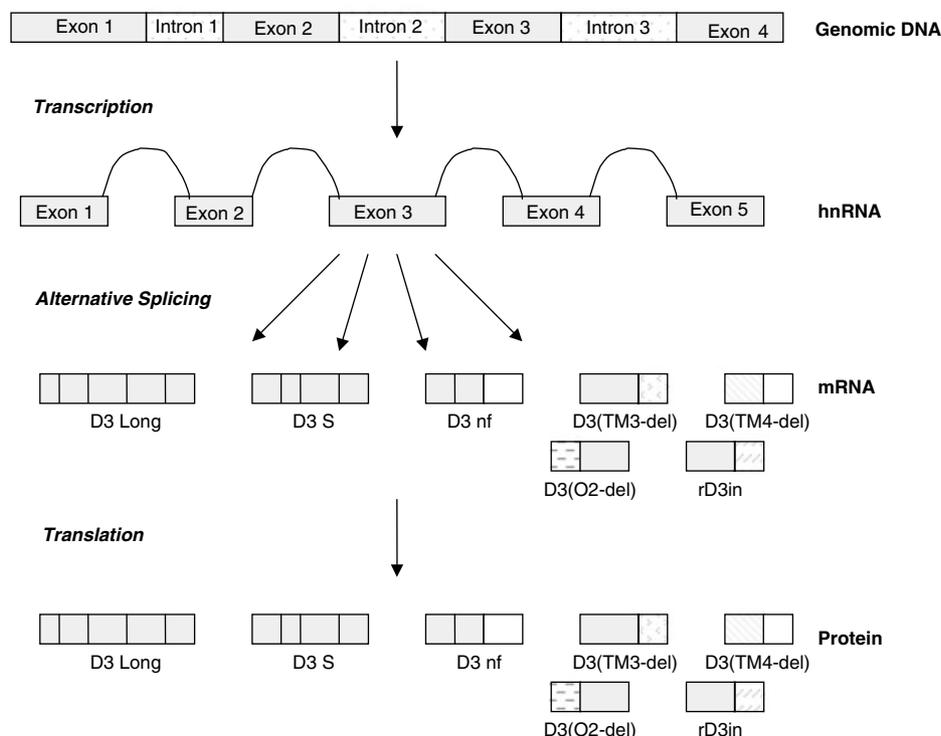


Figure 1 D3 receptor alternative splicing.

variants. D3nf is formed through a deletion of 98 bp in the third cytoplasmic loop, causing a coding frame shift resulting in the creation of a novel 55-amino-acid peptide and appearance of a new premature stop codon. The prematurely truncated protein thus lacks transmembrane domains 6 and 7 (Schmauss *et al*, 1993; Liu *et al*, 1994b), and does not bind dopamine (Elmhurst *et al*, 2000). D3nf mRNA and protein are expressed in rat, monkey, and human brain (Nimchinsky *et al*, 1997; Liu *et al*, 1994b). Importantly, the highly conserved nature of D3nf splicing and protein expression, from rat to human, suggest D3nf likely plays an important, although currently unknown, biological function. Evidence from three laboratories demonstrates that D3nf binds to the full-length D3 receptor subunit (Nimchinsky *et al*, 1997; Elmhurst *et al*, 2000; Karpa *et al*, 2000). D3nf expression inhibits dopamine binding to full-length D3 receptor (Elmhurst *et al*, 2000), and also redirects full-length D3 receptor localization away from the plasma membrane, and instead into an intracellular compartment (Karpa *et al*, 2000). Importantly, D3 mRNA expression is decreased in the cortex of schizophrenia patients (Schmauss *et al*, 1993), whereas increased D3nf splicing efficiency was observed in the cortex of post-mortem tissue from schizophrenia patients (Schmauss, 1996). These findings suggest that increased D3nf expression may contribute to functional states of altered dopaminergic activity. Collectively, these observations suggest that, in a manner analogous to the way that dimerization plays an important role in the modulation of cell signaling for the homologous insulin- and gonadotropin-releasing hormone receptors (Kahn *et al*, 1981; Gregory *et al*, 1982), D3nf dimerization with full-length D3, D2, or D1 receptor could regulate dopamine signaling. Thus, alterations in D3 receptor isoform expression could modulate the effects of dopamine stimulation in altered functional states such as behavioral sensitization.

D3 RECEPTOR ALTERNATIVE SPLICING AND BEHAVIORAL SENSITIZATION

The information described above suggests a sensitization model of response to repetitive stimulant drug exposure illustrated in Figure 2: (1) AMPH increases extracellular dopamine, activating opposing second messenger signaling systems downstream of D1 and D3 receptor stimulation. (2) D3 and D1 receptor stimulations interact at the second messenger level (Ridray *et al*, 1998; Schwartz *et al*, 1998), for example, through opposite regulatory effects on ERK activity and Fos gene family expression (Zhang *et al*, 2004). This interaction is depicted in Figure 2 as occurring within the same cell because D3 and D1 receptors are frequently coexpressed (Schwartz *et al*, 1998); however, the interaction could also occur at the systems level (Xu *et al*, 1997; Jung and Schmauss, 1999; Levavi-Sivan *et al*, 1998). The D3 receptor has highest dopamine affinity, and persistent D3 receptor stimulation following dopamine transporter blockade results in homeostatic mechanisms opposing receptor stimulation. This homeostatic response would include (3) a change in D3 hnRNA alternative splicing in the cell nucleus resulting in increased D3nf expression. D3nf is a receptor splice variant whose function opposes D3 receptor stimula-

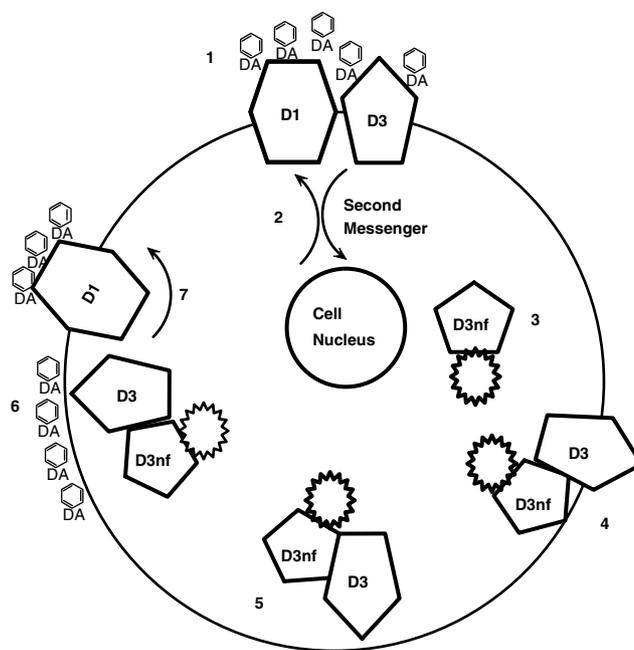


Figure 2 D3/ D3nf mechanism of behavioral sensitization to AMPH. (1) AMPH increases extracellular dopamine. (2) D1 and D3 receptor stimulation activate opposing intracellular signaling systems. D3 receptor has highest dopamine affinity. D3 stimulation results in homeostatic mechanisms opposing receptor stimulation, including (3) increased D3nf expression. (4) D3nf and D3 dimerize, directing the D3/D3nf dimer (5) toward intracytoplasmic trafficking pools and removing D3 receptor from the synaptic membrane. (6) At the next AMPH exposure, D3 receptor is not available to bind dopamine. (7) The result is release of D3 receptor-mediated opposition to D1 receptor stimulation of adenylate cyclase activity. For additional details see in the text.

tion. D3nf shares an NH₃-terminus 'dimerization domain' with the D3 receptor, resulting in (4) dimerization between D3nf and D3. D3nf also contains a unique protein sequence at the COOH-tail, which may direct the D3/D3nf dimer (5) toward intracytoplasmic trafficking pools, thereby removing the D3 receptor from a functional position at the synaptic membrane. In this manner, at the next AMPH exposure (6), the D3 receptor is localized in an intracytoplasmic pool, and is not available to bind dopamine at the synapse. The resulting loss of D3 receptor 'brake', at both the cellular and systems level, includes, for example (7), the loss of D3-receptor-mediated opposition to D1 receptor stimulation of adenylate cyclase activity.

EVIDENCE FOR D3 RECEPTOR THEORY OF BEHAVIORAL SENSITIZATION

Experimental evidence supporting the D3 receptor model of sensitization comes from three lines of data: (1) the effect of D3 receptor antagonists on the development of sensitization; (2) loss of D3 receptor function following sensitization; and (3) decreased D3 receptor mRNA and binding following sensitization.

If tolerance of persistent D3 receptor stimulation contributes to the development of sensitization, blockade with D3 receptor antagonist should inhibit the development of sensitization to dopamine agonists. Consistent with this

prediction, studies in different laboratories demonstrate inhibition of locomotor sensitization by pretreatment with the D3 receptor antagonists nafadotride (Richtand *et al*, 2000), U99194A, or GR103691 (Chiang *et al*, 2003). Additionally, the effect of U99194A was attenuated by pretreatment with the D3 receptor agonist 7-OH-DPAT (Chiang *et al*, 2003), providing further evidence in support of a model in which D3 receptor stimulation contributes to the development of sensitization.

BEHAVIORAL RESPONSE TO D3 RECEPTOR AGONIST IS DIMINISHED IN AMPH-SENSITIZED RATS

A second, independent line of evidence regarding the D3 hypothesis relates to D3 function in sensitized animals. If persistent release of D3-receptor-mediated inhibitory influence contributes to the expression of sensitization, the behavioral response to D3 agonists should be attenuated in sensitized animals. Consistent with this prediction, D3-receptor-mediated locomotor inhibition is dramatically reduced in sensitized rats (Richtand *et al*, 2003). We determined the behavioral response to D3 receptor agonists 7-OH-DPAT and PD 128907 1 week following repetitive AMPH treatment. Locomotor response to novelty is inhibited by the D3 agonists PD 128907 (shown in Figure 3) and 7-OH-DPAT in animals pretreated with a 5-day saline injection regimen, whereas this D3-receptor-mediated locomotor inhibition is not detectable following completion of a sensitizing AMPH regimen. This study also found, unexpectedly, that the locomotor response to AMPH, and locomotor inhibition by D3 agonist, are negatively correlated within individual animals, accounting for over 50% of the individual variance in locomotor response to AMPH, and that this relationship is abolished following repetitive AMPH treatment. In combination, these findings suggest that release of D3-receptor-mediated inhibition contributes to the expression of behavioral sensitization (Richtand *et al*, 2003).

LONG-TERM CONSEQUENCES OF STIMULANT DRUG EXPOSURE

The third line of evidence regarding the D3 hypothesis relates to underlying mechanism. If a persistent decline of D3-receptor-mediated inhibition contributes to sensitization, the underlying mechanism should be evident through analysis of the effects of repetitive stimulant exposure on D3 mRNA, binding activity, downstream signaling pathways, or other indices of receptor function. The evidence in this area to date, on balance, has not been conclusive, perhaps in part because direct measures of D3 receptor *function* have been elusive. In support of the D3 receptor hypothesis, multiple studies in different laboratories have observed decreases in measures of D3 receptor expression in sensitized animals. D3 receptor mRNA and protein expression are both decreased in AMPH-sensitized rats (Chiang *et al*, 2003), and D3 binding in the nucleus accumbens is also decreased following locomotor sensitization to cocaine (Wallace *et al*, 1996). D3 receptor binding is also decreased in the nucleus accumbens in rat offspring exposed to stress in late pregnancy, a manipulation causing increased locomotor

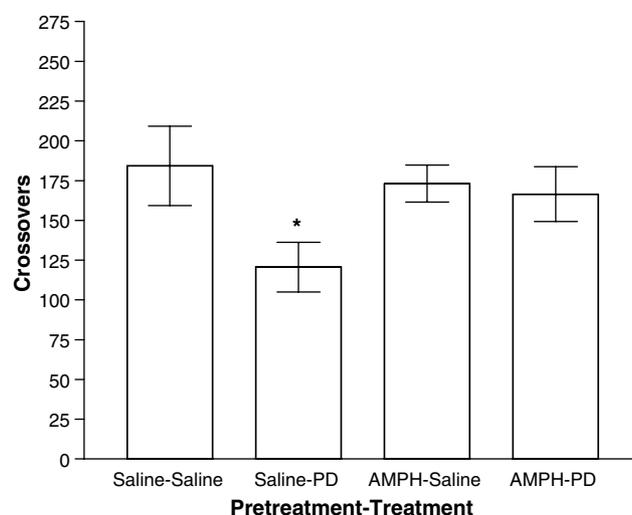


Figure 3 Behavioral response to PD 128907 is diminished in behaviorally sensitized rats. Locomotion following injection with saline and PD 128907 is shown following saline or AMPH pretreatment. *Saline pretreatment group:* *PD 128907 vs saline injection planned comparison $p=0.021$, one-tailed. *AMPH pretreatment group:* PD 128907 vs saline injection planned comparison $p=0.754$. *Saline vs AMPH pretreatment group response to saline injection mean comparison* $p=0.368$, two-tailed. *Methods:* Male Sprague-Dawley were treated in home cages for 5 consecutive days with either saline or AMPH (1 mg/kg). On days 12–14, rats were injected in random order with either saline or PD 128907 (10 μ g/kg) on a rotating basis, allowing each animal to be used as its own control to measure D3-agonist-mediated inhibitory effect. Data analysis by one-way ANOVA revealed no difference in order of injection (saline first or PD 128907 first). Crossover data from 0 to 180 min following injection with saline and PD 128907 were analyzed using ANOVA techniques for independent observations. Results are expressed as geometric mean number of crossovers, \pm SE, for each group. Planned *t*-tests were performed for hypotheses of interest. Adapted from Richtand *et al* (2003).

response to novelty, more rapid locomotor sensitization, and increased AMPH self-administration (Henry *et al*, 1995). D3 receptor binding is also decreased in adult rats following neonatal hippocampal lesion (Flores *et al*, 1996), suggesting hyperlocomotor behavioral responses in that model also result from decreased D3 receptor inhibitory function (Wan and Corbett, 1997).

In contrast to the studies described above, we have not detected significant alterations in full-length D3 mRNA expression (Hondo *et al*, 1999) or D3 receptor binding in ventral striatum of AMPH-treated rats (Richtand *et al*, 2003). Studies of D3 expression in human cocaine-dependent subjects have also had conflicting results (Staley and Mash, 1996; Mash and Staley, 1999; Segal *et al*, 1997; Meador-Woodruff *et al*, 1995). In one series studying a small number of subjects ($n=6$ /group), D3 receptor binding and mRNA were both increased in cocaine overdose victims in comparison to age-matched control subjects; however, comparable changes were not observed in cocaine delirium victims (Staley and Mash, 1996; Mash and Staley, 1999; Segal *et al*, 1997). In contrast, another study failed to detect any difference between control and cocaine-abusing subjects in D3 mRNA in autopsy tissue (Meador-Woodruff *et al*, 1995). None of the published studies to date have been designed to distinguish alternatively spliced D3 receptor isoforms. Future studies evaluating D3 receptor isoform expression, subcellular

localization, and intracellular signaling pathways may be able to more clearly elucidate the mechanism underlying the decline in D3 receptor-mediated inhibitory function accompanying behavioral sensitization.

CONCLUSIONS

Behavioral sensitization may be accounted for in part by tolerance of D3-receptor-mediated inhibitory systems. Perturbations in D3-receptor-mediated inhibitory function may contribute to several neuropsychiatric disorders, including drug dependence, schizophrenia, and Parkinson's disease. Because highly selective D3 receptor ligands have been synthesized (Newman *et al*, 2005), identifying the role of the D3 receptor in the pathophysiology of each of these conditions could make novel approaches to drug treatment feasible. Importantly, D3 receptor tissue distribution is relatively limited within limbic brain regions, suggesting the potential for focused treatments with few side effects. Additionally, elucidating the role of alternative splicing pathways in behavioral sensitization may identify a novel mechanism at which to intervene both to prevent sensitization, as well as to potentially reverse sensitization through restoration of D3-mediated inhibitory function.

ACKNOWLEDGEMENTS

This work was supported by the Department of Veterans Affairs Medical Research Service, and National Institute of Drug Abuse (DA016778-01)

REFERENCES

- Accili D, Fishburn CS, Drago J, Steiner H, Lachowicz JE, Park BH *et al* (1996). A targeted mutation of the D3 dopamine receptor gene is associated with hyperactivity in mice. *Proc Natl Acad Sci USA* **93**: 1945–1949.
- Ackerman JM, White FJ (1990). A10 somatodendritic dopamine autoreceptor sensitivity following withdrawal from repeated cocaine treatment. *Neurosci Lett* **117**: 181–187.
- Bell DS (1973). The experimental reproduction of amphetamine psychosis. *Arch Gen Psychiatry* **29**: 35–40.
- Bijou Y, Stinus L, Le Moal M, Cador M (1996). Evidence for selective involvement of dopamine D1 receptors of the ventral tegmental area in the behavioral sensitization induced by intraventricular injections of D-amphetamine. *J Pharmacol Exp Ther* **277**: 1177–1187.
- Bouthenet ML, Souil E, Martres MP, Sokoloff P, Giros B, Schwartz JC (1991). Localization of dopamine D3 receptor mRNA in the rat brain using in situ hybridization histochemistry: comparison with dopamine D2 receptor mRNA. *Brain Res* **564**: 203–219.
- Chiang YC, Chen PC, Chen JC (2003). D(3) dopamine receptors are down-regulated in amphetamine sensitized rats and their putative antagonists modulate the locomotor sensitization to amphetamine. *Brain Res* **972**: 159–167.
- Costall B, Naylor RJ (1974). Extrapyramidal and mesolimbic involvement with the stereotypic activity of D- and L-amphetamine. *Eur J Pharmacol* **25**: 121–129.
- Creese I, Iversen SD (1972). Amphetamine response in rat after dopamine neurone destruction. *Nat N Biol* **238**: 247–248.
- Creese I, Iversen SD (1975). The pharmacological and anatomical substrates of the amphetamine response in the rat. *Brain Res* **83**: 419–436.
- Dalia A, Uretsky NJ, Wallace LJ (1998). Dopaminergic agonists administered into the nucleus accumbens: effects on extracellular glutamate and on locomotor activity. *Brain Res* **788**: 111–117.
- Diaz J, Levesque D, Griffon N, Lammers CH, Martres MP, Sokoloff P *et al* (1994). Opposing roles for dopamine D2 and D3 receptors on neurotensin mRNA expression in nucleus accumbens. *Eur J Neurosci* **6**: 1384–1387.
- Dreher JK, Jackson DM (1989). Role of D1 and D2 dopamine receptors in mediating locomotor activity elicited from the nucleus accumbens of rats. *Brain Res* **487**: 267–277.
- Ekman A, Nissbrandt H, Heilig M, Dijkstra D, Eriksson E (1998). Central administration of dopamine D3 receptor antisense to rat: effects on locomotion, dopamine release and [³H]spiperone binding. *Naunyn Schmiedeberg's Arch Pharmacol* **358**: 342–350.
- Ellinwood EHJ, Sudilovsky A, Nelson LM (1973). Evolving behavior in the clinical and experimental amphetamine (model) psychosis. *Am J Psychiatry* **130**: 1088–1093.
- Ellinwood Jr EH (1967). Amphetamine psychosis: I. Description of the individuals and process. *J Nerv Ment Dis* **144**: 273–283.
- Elmhurst JL, Xie Z, O'Dowd BF, George SR (2000). The splice variant D3nf reduces ligand binding to the D3 dopamine receptor: evidence for heterooligomerization. *Brain Res Mol Brain Res* **80**: 63–74.
- Fishburn CS, Belleli D, David C, Carmon S, Fuchs S (1993). A novel short isoform of the D3 dopamine receptor generated by alternative splicing in the third cytoplasmic loop. *J Biol Chem* **268**: 5872–5878.
- Flores G, Barbeau D, Quirion R, Srivastava LK (1996). Decreased binding of dopamine D3 receptors in limbic subregions after neonatal bilateral lesion of rat hippocampus. *J Neurosci* **16**: 2020–2026.
- Fog R, Randrup A, Pakkenberg H (1970). Lesions in corpus striatum and cortex of rat brains and the effect on pharmacologically induced stereotyped, aggressive and cataleptic behaviour. *Psychopharmacologia* **18**: 346–356.
- Fu D, Skryabin BV, Brosius J, Robakis NK (1995). Molecular cloning and characterization of the mouse dopamine D3 receptor gene: an additional intron and an mRNA variant. *DNA Cell Biol* **14**: 485–492.
- Giros B, Martres MP, Pilon C, Sokoloff P, Schwartz JC (1991). Shorter variants of the D3 dopamine receptor produced through various patterns of alternative splicing. *Biochem Biophys Res Commun* **176**: 1584–1592.
- Gregory H, Taylor CL, Hopkins CR (1982). Luteinizing hormone release from dissociated pituitary cells by dimerization of occupied LHRH receptors. *Nature* **300**: 269–271.
- Griffith JD, Cavanaugh J, Held J, Oates JA (1972). Dextroamphetamine: evaluation of psychotomimetic properties in man. *Arch Gen Psychiatry* **26**: 97–100.
- Griffon N, Crocq MA, Pilon C, Martres MP, Mayerova A, Uyanik G *et al* (1996). Dopamine D3 receptor gene: organization, transcript variants, and polymorphism associated with schizophrenia. *Am J Med Genet* **67**: 63–70.
- Gurevich EV, Joyce JN (1999). Distribution of dopamine D3 receptor expressing neurons in the human forebrain: comparison with D2 receptor expressing neurons. *Neuropsychopharmacology* **20**: 60–80.
- Henry C, Guegant G, Cador M, Arnauld E, Arsaut J, Le Moal M *et al* (1995). Prenatal stress in rats facilitates amphetamine-induced sensitization and induces long-lasting changes in dopamine receptors in the nucleus accumbens. *Brain Res* **685**: 179–186.
- Henry DJ, Greene MA, White FJ (1989). Electrophysiological effects of cocaine in the mesoaccumbens dopamine system: repeated administration. *J Pharmacol Exp Ther* **251**: 833–839.
- Henry DJ, White FJ (1991). Repeated cocaine administration causes persistent enhancement of D1 dopamine receptor sensitivity within the rat nucleus accumbens. *J Pharmacol Exp Ther* **258**: 882–890.

- Higashi H, Inanaga K, Nishi S, Uchimura N (1989). Enhancement of dopamine actions on rat nucleus accumbens neurones *in vitro* after methamphetamine pretreatment. *J Physiol* **408**: 587–603.
- Hitzemann R, Wu J, Hom D, Loh H (1980). Brain locations controlling the behavioral effects of chronic amphetamine intoxication. *Psychopharmacology (Berlin)* **72**: 93–101.
- Hondo H, Spitzer RH, Grinius B, Richtand NM (1999). Quantification of dopamine D3 receptor mRNA level associated with the development of amphetamine-induced behavioral sensitization in the rat brain. *Neurosci Lett* **264**: 69–72.
- Jones SR, Lee TH, Wightman RM, Ellinwood EH (1996a). Effects of intermittent and continuous cocaine administration on dopamine release and uptake regulation in the striatum: *in vitro* voltammetric assessment. *Psychopharmacology (Berlin)* **126**: 331–338.
- Jones SR, O'Dell SJ, Marshall JF, Wightman RM (1996b). Functional and anatomical evidence for different dopamine dynamics in the core and shell of the nucleus accumbens in slices of rat brain. *Synapse* **23**: 224–231.
- Jung MY, Schmauss C (1999). Decreased c-fos responses to dopamine D(1) receptor agonist stimulation in mice deficient for D(3) receptors. *J Biol Chem* **274**: 29406–29412.
- Kahn CR, Baird KL, Flier JS, Grunfeld C, Harmon JT, Harrison LC *et al* (1981). Insulin receptors, receptor antibodies, and the mechanism of insulin action. *Recent Prog Horm Res* **37**: 477–538.
- Kalivas PW, Alesdatter JE (1993). Involvement of N-methyl-D-aspartate receptor stimulation in the ventral tegmental area and amygdala in behavioral sensitization to cocaine. *J Pharmacol Exp Ther* **267**: 486–495.
- Kalivas PW, Duffy P (1990). Effect of acute and daily cocaine treatment on extracellular dopamine in the nucleus accumbens. *Synapse* **5**: 48–58.
- Kalivas PW, Duffy P (1993). Time course of extracellular dopamine and behavioral sensitization to cocaine. I. Dopamine axon terminals. *J Neurosci* **13**: 266–275.
- Kalivas PW, Weber B (1988). Amphetamine injection into the ventral mesencephalon sensitizes rats to peripheral amphetamine and cocaine. *J Pharmacol Exp Ther* **245**: 1095–1102.
- Kamata K, Rebec GV (1984). Long-term amphetamine treatment attenuates or reverses the depression of neuronal activity produced by dopamine agonists in the ventral tegmental area. *Life Sci* **34**: 2419–2427.
- Karler R, Finnegan KT, Calder LD (1993). Blockade of behavioral sensitization to cocaine and amphetamine by inhibitors of protein synthesis. *Brain Res* **603**: 19–24.
- Karpa KD, Lin R, Kabbani N, Levenson R (2000). The dopamine D3 receptor interacts with itself and the truncated D3 splice variant d3nf: D3–D3nf interaction causes mislocalization of D3 receptors. *Mol Pharmacol* **58**: 677–683.
- Landwehrmeyer B, Mengod G, Palacios JM (1993). Differential visualization of dopamine D2 and D3 receptor sites in rat brain. A comparative study using *in situ* hybridization histochemistry and ligand binding autoradiography. *Eur J Neurosci* **5**: 145–153.
- Le Moine C, Bloch B (1996). Expression of the D3 dopamine receptor in peptidergic neurons of the nucleus accumbens: comparison with the D1 and D2 dopamine receptors. *Neuroscience* **73**: 131–143.
- Levant B (1998). Differential distribution of D3 dopamine receptors in the brains of several mammalian species. *Brain Res* **800**: 269–274.
- Levavi-Sivan B, Park BH, Fuchs S, Fishburn CS (1998). Human D3 dopamine receptor in the medulloblastoma TE671 cell line: cross-talk between D1 and D3 receptors. *FEBS Lett* **439**: 138–142.
- Levesque D, Diaz J, Pilon C, Martres MP, Giros B, Souil E *et al* (1992). Identification, characterization, and localization of the dopamine D3 receptor in rat brain using 7-[3H]hydroxy-N,N-dipropyl-2-aminotetralin. *Proc Natl Acad Sci USA* **89**: 8155–8159.
- Lieberman JA, Sheitman BB, Kinon BJ (1997). Neurochemical sensitization in the pathophysiology of schizophrenia: deficits and dysfunction in neuronal regulation and plasticity. *Neuro-psychopharmacology* **17**: 205–229.
- Liu JC, Cox RF, Greif GJ, Freedman JE, Waszczak BL (1994a). The putative dopamine D3 receptor agonist 7-OH-DPAT: lack of mesolimbic selectivity. *Eur J Pharmacol* **264**: 269–278.
- Liu K, Bergson C, Levenson R, Schmauss C (1994b). On the origin of mRNA encoding the truncated dopamine D3-type receptor D3nf and detection of D3nf-like immunoreactivity in human brain. *J Biol Chem* **269**: 29220–29226.
- Mash DC, Staley JK (1999). D3 dopamine and kappa opioid receptor alterations in human brain of cocaine-overdose victims. *Ann NY Acad Sci* **877**: 507–522.
- Meador-Woodruff JH, Little KY, Damask SP, Watson SJ (1995). Effects of cocaine on D3 and D4 receptor expression in the human striatum. *Biol Psychiatry* **38**: 263–266.
- Menalled LB, Dziewczapolski G, Garcia MC, Rubinstein M, Gershanik OS (1999). D3 receptor knockdown through antisense oligonucleotide administration supports its inhibitory role in locomotion. *Neuroreport* **10**: 3131–3136.
- Nagai Y, Ueno S, Saeki Y, Soga F, Yanagihara T (1993). Expression of the D3 dopamine receptor gene and a novel variant transcript generated by alternative splicing in human peripheral blood lymphocytes. *Biochem Biophys Res Commun* **194**: 368–374.
- Nestler EJ, Terwilliger RZ, Walker JR, Sevarino KA, Duman RS (1990). Chronic cocaine treatment decreases levels of the G protein subunits Gi alpha and Go alpha in discrete regions of rat brain. *J Neurochem* **55**: 1079–1082.
- Newman AH, Grundt P, Nader MA (2005). Dopamine D3 receptor partial agonists and antagonists as potential drug abuse therapeutic agents. *J Med Chem* **48**: 3663–3679.
- Nimchinsky EA, Hof PR, Janssen WGM, Morrison JH, Schmauss C (1997). Expression of dopamine D3 receptor dimers and tetramers in brain and in transfected cells. *J Biol Chem* **272**: 29229–29237.
- Pagliusi S, Chollet-Daemerius A, Losberger C, Mills A, Kawashima E (1993). Characterization of a novel exon within the D3 receptor gene giving rise to an mRNA isoform expressed in rat brain. *Biochem Biophys Res Commun* **194**: 465–471.
- Park BH, Fishburn CS, Carmon S, Accili D, Fuchs S (1995). Structural organization of the murine D3 dopamine receptor gene. *J Neurochem* **64**: 482–486.
- Parsons LH, Justice JBJ (1992). Extracellular concentration and *in vivo* recovery of dopamine in the nucleus accumbens using microdialysis. *J Neurochem* **58**: 212–218.
- Paulson PE, Camp DM, Robinson TE (1991). Time course of transient behavioral depression and persistent behavioral sensitization in relation to regional brain monoamine concentrations during amphetamine withdrawal in rats. *Psychopharmacology (Berlin)* **103**: 480–492.
- Piazza PV, Deminiere JM, Le Moal M, Simon H (1989). Factors that predict individual vulnerability to amphetamine self-administration. *Science* **245**: 1511–1513.
- Pijnenburg AJ, Honig WM, Van Rossum JM (1975). Inhibition of *d*-amphetamine-induced locomotor activity by injection of haloperidol into the nucleus accumbens of the rat. *Psychopharmacology* **41**: 87–95.
- Pritchard LM, Logue AD, Hayes S, Welge JA, Xu M, Zhang J *et al* (2003). 7-OH-DPAT and PD 128907 selectively activate the D3 dopamine receptor in a novel environment. *Neuropsychopharmacology* **28**: 100–107.
- Richtand NM, Goldsmith RJ, Nolan JE, Berger SP (2001a). The D3 dopamine receptor and substance dependence. *J Addict Dis* **20**: 19–32.
- Richtand NM, Kelsoe JR, Segal DS, Kuczenski R (1995). Regional quantification of D1, D2, and D3 dopamine receptor mRNA in rat brain using a ribonuclease protection assay. *Brain Res Mol Brain Res* **33**: 97–103.

- Richtand NM, Logue AD, Welge JA, Perdue J, Tubbs LJ, Spitzer RH *et al* (2000). The dopamine D3 receptor antagonist nafadotride inhibits development of locomotor sensitization to amphetamine. *Brain Res* **867**: 239–242.
- Richtand NM, Welge JA, Levant B, Logue AD, Hayes S, Pritchard LM *et al* (2003). Altered behavioral response to dopamine D3 receptor agonists 7-OH-DPAT and PD 128907 following repetitive amphetamine administration. *Neuropsychopharmacology* **28**: 1422–1432.
- Richtand NM, Woods SC, Berger SP, Strakowski SM (2001b). D3 dopamine receptor, behavioral sensitization, and psychosis. *Neurosci Biobehav Rev* **25**: 427–443.
- Ridray S, Griffon N, Mignon V, Souil E, Carboni S, Diaz J *et al* (1998). Coexpression of dopamine D1 and D3 receptors in islands of Calleja and shell of nucleus accumbens of the rat: opposite and synergistic functional interactions. *Eur J Neurosci* **10**: 1676–1686.
- Roberts DC, Zis AP, Fibiger HC (1975). Ascending catecholamine pathways and amphetamine-induced locomotor activity: importance of dopamine and apparent non-involvement of norepinephrine. *Brain Res* **93**: 441–454.
- Robinson TE, Berridge KC (1993). The neural basis of drug craving: an incentive-sensitization theory of addiction. *Brain Res Brain Res Rev* **18**: 247–291.
- Robinson TE, Jurson PA, Bennett JA, Bentgen KM (1988). Persistent sensitization of dopamine neurotransmission in ventral striatum (nucleus accumbens) produced by prior experience with (+)-amphetamine: a microdialysis study in freely moving rats. *Brain Res* **462**: 211–222.
- Ross SB (1991). Synaptic concentration of dopamine in the mouse striatum in relationship to the kinetic properties of the dopamine receptors and uptake mechanism. *J Neurochem* **56**: 22–29.
- Sato M, Chen CC, Akiyama K, Otsuki S (1983). Acute exacerbation of paranoid psychotic state after long-term abstinence in patients with previous methamphetamine psychosis. *Biol Psychiatry* **18**: 429–440.
- Scarselli M, Novi F, Schallmach E, Lin R, Baragli A, Colzi A *et al* (2001). D2/D3 dopamine receptor heterodimers exhibit unique functional properties. *J Biol Chem* **276**: 30308–30314.
- Schmauss C (1996). Enhanced cleavage of an atypical intron of dopamine D3-receptor pre-mRNA in chronic schizophrenia. *J Neurosci* **16**: 7902–7909.
- Schmauss C, Haroutunian V, Davis KL, Davidson M (1993). Selective loss of dopamine D3-type receptor mRNA expression in parietal and motor cortices of patients with chronic schizophrenia. *Proc Natl Acad Sci USA* **90**: 8942–8946.
- Schwartz JC, Diaz J, Bordet R, Griffon N, Perachon S, Pilon C *et al* (1998). Functional implications of multiple dopamine receptor subtypes: the D1/D3 receptor coexistence. *Brain Res Brain Res Rev* **26**: 236–242.
- Segal DM, Moraes CT, Mash DC (1997). Up-regulation of D3 dopamine receptor mRNA in the nucleus accumbens of human cocaine fatalities. *Brain Res Mol Brain Res* **45**: 335–339.
- Segal DS, Geyer MA, Schuckit MA (1981). Stimulant-induced psychosis: an evaluation of animal models. In: Youdim MBH, Lovenberg W, Sharman DF, Lagnado JR (eds). *Essays in Neurochemistry and Neuropharmacology*. John Wiley & Sons, Ltd: New York. Vol. 5, pp 95–129.
- Segal DS, Mandell AJ (1974). Long-term administration of *d*-amphetamine: progressive augmentation of motor activity and stereotypy. *Pharmacol Biochem Behav* **2**: 249–255.
- Segal DS, Schuckit MA (1983). Animal models of stimulant-induced psychosis. In: Creese I (ed). *Stimulants: Neurochemical, Behavioral and Clinical Perspectives*. Raven Press: New York. pp 131–167.
- Segal DS, Weinberger SB, Cahill J, McCunney SJ (1980). Multiple daily amphetamine administration: behavioral and neurochemical alterations. *Science* **207**: 905–907.
- Snyder LA, Roberts JL, Sealfon SC (1991). Alternative transcripts of the rat and human dopamine D3 receptor. *Biochem Biophys Res Commun* **180**: 1031–1035.
- Sokoloff P, Martres MP, Giros B, Bouthenet ML, Schwartz JC (1992). The third dopamine receptor (D3) as a novel target for antipsychotics. *Biochem Pharmacol* **43**: 659–666.
- Sorg BA, Ulibarri C (1995). Application of a protein synthesis inhibitor into the ventral tegmental area, but not the nucleus accumbens, prevents behavioral sensitization to cocaine. *Synapse* **20**: 217–224.
- Staley JK, Mash DC (1996). Adaptive increase in D3 dopamine receptors in the brain reward circuits of human cocaine fatalities. *J Neurosci* **16**: 6100–6106.
- Stewart J, Vezina P (1989). Microinjections of Sch-23390 into the ventral tegmental area and substantia nigra pars reticulata attenuate the development of sensitization to the locomotor activating effects of systemic amphetamine. *Brain Res* **495**: 401–406.
- Surmeier DJ, Song WJ, Yan Z (1996). Coordinated expression of dopamine receptors in neostriatal medium spiny neurons. *J Neurosci* **16**: 6579–6591.
- Terwilliger RZ, Beitner-Johnson D, Sevarino KA, Crain SM, Nestler EJ (1991). A general role for adaptations in G-proteins and the cyclic AMP system in mediating the chronic actions of morphine and cocaine on neuronal function. *Brain Res* **548**: 100–110.
- Utena H (1966). Behavioral aberrations in methamphetamine-intoxicated animals and chemical correlates in the brain. *Prog Brain Res* **21**: 192–207.
- Vezina P, Stewart J (1990). Amphetamine administered to the ventral tegmental area but not to the nucleus accumbens sensitizes rats to systemic morphine: lack of conditioned effects. *Brain Res* **516**: 99–106.
- Wallace DR, Mactutus CF, Booze RM (1996). Repeated intravenous cocaine administration: locomotor activity and dopamine D2/D3 receptors. *Synapse* **23**: 152–163.
- Wan RQ, Corbett R (1997). Enhancement of postsynaptic sensitivity to dopaminergic agonists induced by neonatal hippocampal lesions. *Neuropsychopharmacology* **16**: 259–268.
- Waters N, Svensson K, Haadma-Svensson SR, Smith MW, Carlsson A (1993). The dopamine D3-receptor: a postsynaptic receptor inhibitory on rat locomotor activity. *J Neural Transm Gen Sect* **94**: 11–19.
- White FJ, Wang RY (1984). Electrophysiological evidence for A10 dopamine autoreceptor subsensitivity following chronic *D*-amphetamine treatment. *Brain Res* **309**: 283–292.
- Wolf ME (1998). The role of excitatory amino acids in behavioral sensitization to psychomotor stimulants. *Prog Neurobiol* **54**: 679–720.
- Wolf ME, Dahlin SL, Hu XT, Xue CJ, White K (1995). Effects of lesions of prefrontal cortex, amygdala, or fornix on behavioral sensitization to amphetamine: comparison with N-methyl-D-aspartate antagonists. *Neuroscience* **69**: 417–439.
- Xu M, Koeltzow TE, Santiago GT, Moratalla R, Cooper DC, Hu XT *et al* (1997). Dopamine D3 receptor mutant mice exhibit increased behavioral sensitivity to concurrent stimulation of D1 and D2 receptors. *Neuron* **19**: 837–848.
- Zeng C, Wang Z, Li H, Yu P, Zheng S, Wu L *et al* (2006). D3 dopamine receptor directly interacts with D1 dopamine receptor in immortalized renal proximal tubule cells. *Hypertension* **47**: 573–579.
- Zetterstrom T, Sharp T, Marsden CA, Ungerstedt U (1983). In vivo measurement of dopamine and its metabolites by intracerebral dialysis: changes after *d*-amphetamine. *J Neurochem* **41**: 1769–1773.
- Zhang L, Lou D, Jiao H, Zhang D, Wang X, Xia Y *et al* (2004). Cocaine-induced intracellular signaling and gene expression are oppositely regulated by the dopamine D1 and D3 receptors. *J Neurosci* **24**: 3344–3354.