Supplementary Methods:

Generating DH10MultiBac\textsuperscript{Cre} cells.

Expression of Cre Recombinase Protein und generation of electro-competent DH10MultiBac\textsuperscript{Cre} cells:

1. Electroporate pBADZ-HisCre plasmid into DH10MultiBac cells (25 microFD, 2.0 kV, 200 Ohm).
2. Grow in 2xTY medium for 4h at 37\textdegree C.
3. Plate on LS plates (low salt medium agar) with antibiotics (kanamycin, tetracyclin, zeocin) and X-Gal/IPTG.
4. Grow a 500 ml LS (low salt) culture from one blue colony.
   Antibiotics: Kanamycin/tetracyclin/zeocin, Temp. 37\textdegree C or RT.
5. Grow to OD\textsubscript{600}=0.25 at 37\textdegree C or RT.
6. Take 500 µl ample („Minus“-probe)
7. Add L-arabinose to 0.1% (0.5 g in 500 ml).
8. Grow to OD\textsubscript{600}=0.5.
9. Take 250 µl sample („Plus“-probe)
10. Cool culture on ice for 15 min.
11. Resuspend in 500 ml ICE COLD STERILE 10% glycerol sol.
12. Centrifuge at 4000rpm, 4\textdegree C, 15 min.
13. Resuspend in 250 ml ICE COLD STERILE 10% glycerol sol.
14. Centrifuge at 4000rpm, 4\textdegree C, 15 min.
15. Resuspend in 10 ml ICE COLD STERILE 10% glycerol sol.
16. Centrifuge at 4000rpm, 4\textdegree C, 15 min.
17. Resuspend in 1 ml ICE COLD STERILE 10% glycerol sol.
18. Prepare 80 µl aliquots (sterile Eppendorfs).
19. Shock-freeze in liq. nitrogen, store at –70\textdegree C.
21. Resuspend in 150 ul protein gel loading buffer. Analyze by 15% SDS-PAGE (load 5-10 ul) (Fig. S3)
Cre expression in DH10MultiBac\textsuperscript{Cre} competent cells
(reprinted with permission from Ref. 1)

Strong Cre expression is mandatory for successful integration of pUCDM derivatives!

Low salt medium/Agar for zeocin cultures:
1. Combine 10 g tryptone, 5g NaCl, 5g yeast extract
2. Add water (dd) to 950 ml
3. Adjust pH to 7.5 with 1N NaOH
4. Add water (dd) to 1L (for plates add 15g/L agar) and autoclave.
5. Add zeocin to 25 \(\mu\)g/ml below 55\(^\circ\)C (same for other antibiotics)
   X-Gal to 500\(\mu\)g/ml from 1000x stock
   IPTG to 0.5 \(\mu\)M from 1000x stock

Store plates at 4\(^\circ\)C in the dark (X-Gal is light sensitive).

Purifying composite MultiBac virus and restriction mapping.
(adapted from Ref. 9):
1. Centrifuge 25ml viral supernatant for 75min in a SW28 rotor at 24,000 rpm through 3mls of a 25% sucrose cushion.
2. Resuspend pellet in 1ml of 10mM Tris (pH 7.5), 100mM NaCl, 10mM EDTA, 0.25%SDS.
3. Incubate at 50\(^\circ\)C for 4h.
4. Extract with phenol/chloroform, NaOAc/EtOH precipitate.
5. Digest with Sall according to manufacturers (NEB) instructions.