Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

<table>
<thead>
<tr>
<th>FIGURE NUMBER</th>
<th>TEST USED</th>
<th>n</th>
<th>DESCRIPTIVE STATS (AVERAGE, VARIANCE)</th>
<th>P VALUE</th>
<th>DEGREES OF FREEDOM &amp; F/T/Z/R/ETC VALUE</th>
</tr>
</thead>
<tbody>
<tr>
<td>1a</td>
<td>one-way ANOVA</td>
<td>9, 9, 10, 15 mice from at least 3 litters/group</td>
<td>error bars are mean +/- SEM</td>
<td>Fig. legend</td>
<td>p = 0.044</td>
</tr>
<tr>
<td>results para 4</td>
<td>unpaired t-test</td>
<td>15 slices from 10 mice</td>
<td>error bars are mean +/- SEM</td>
<td>Results para 5</td>
<td>p = 0.0006</td>
</tr>
</tbody>
</table>

Nature Neuroscience: doi:10.1038/nn.4128
<table>
<thead>
<tr>
<th>FIGURE NUMBER</th>
<th>WHICH TEST?</th>
<th>SECTION &amp; PARAGRAPH #</th>
<th>TEST USED</th>
<th>n</th>
<th>EXACT VALUE</th>
<th>DEFINED?</th>
<th>DESCRIPTIVE STATS (AVERAGE, VARIANCE)</th>
<th>P VALUE</th>
<th>DEGREES OF FREEDOM &amp; F/T/Z/R/ETC VALUE</th>
</tr>
</thead>
<tbody>
<tr>
<td>a 1a</td>
<td></td>
<td></td>
<td>Methods/ Visual stimuli and presentation. Last paragraph.</td>
<td>20</td>
<td>Trials</td>
<td></td>
<td>Arithmetic mean +/- 68% confidence interval</td>
<td></td>
<td>Method/ Characterization of neuronal responses. First paragraph.</td>
</tr>
<tr>
<td>a 1b</td>
<td></td>
<td></td>
<td>Methods/ Visual stimuli and presentation. Paragraph 3.</td>
<td>270</td>
<td>Images</td>
<td></td>
<td>Geometric mean +/- standard deviation</td>
<td>Figure 1 caption</td>
<td></td>
</tr>
<tr>
<td>a 1b</td>
<td></td>
<td></td>
<td>Results/ Variability of surround modulation with natural images. Paragraph 2</td>
<td>207</td>
<td>Neurons</td>
<td></td>
<td>Spearman’s correlation coefficient</td>
<td></td>
<td>Results/ Variability of surround modulation with natural images. Paragraph 2</td>
</tr>
<tr>
<td>a 1c</td>
<td></td>
<td></td>
<td>Results/ Variability of surround modulation with natural images. Last Paragraph.</td>
<td>38,591</td>
<td>207 neurons times 270 images per neuron, minus cases not passing the inclusion criteria (Methods/ Characterization of neuronal responses)</td>
<td></td>
<td>Geometric mean +/- 68% confidence interval</td>
<td></td>
<td>Results/ Variability of surround modulation with natural images. Last paragraph.</td>
</tr>
</tbody>
</table>

Nature Neuroscience: doi:10.1038/nn.4128
| a | 3a | Bootstrap paired t-test for modulation ratio at high surround drive smaller than at low surround drive. | Results / Variability in surround suppression is not well captured by divisive normalization. Paragraph 5 | 126 | Neurons | Methods / Inclusion criteria. | Geometric mean | Result s / Variability in surround suppression is not well captured by divisive normalization. Paragraph 5 | p<4*10^-5 | Results / Variability in surround suppression is not well captured by divisive normalization. Paragraph 5 |
| a | 3b | Bootstrap paired t-test for modulation ratio at high surround drive smaller than at low surround drive. | Results / Surround suppression is stronger for homogeneous natural images. Paragraph 2 | 126 | Neurons | Methods / Inclusion criteria. | Geometric mean | Result s / Surround suppression is stronger for homogeneous natural images. Paragraph 2 | p<4*10^-5 | Results / Surround suppression is stronger for homogeneous natural images. Paragraph 2 |
| a | 6a | Bootstrap t-test for modulation ratio with homogeneous images smaller than with heterogeneous images. Across neurons. | Results / Surround suppression is stronger for homogeneous natural images. Paragraph 1. | 126 | Neurons | Methods / Inclusion criteria. | Geometric mean | Result s / Surround suppression is stronger for homogeneous natural images. Paragraph 1. | p<4*10^-5 | Results / Surround suppression is stronger for homogeneous natural images. Paragraph 1. |
### Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

   If so, what figure(s)?

   No representative figures

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

   If so, where is this reported (section, paragraph #)?

   N/A
### Statistics and general methods

1. **Is there a justification of the sample size?**
   - If so, how was it justified?
   - Where (section, paragraph #)?
   - Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

   - The effects reported are based on n=207 independent neurons recorded from 3 macaques. The sample size is well in line with the standards of the field when studying single-neuron response properties.

2. **Are statistical tests justified as appropriate for every figure?**
   - Where (section, paragraph #)?

   - The test used for figure 1b is a two-tailed permutation test; this is the standard choice for testing significance of a Spearman correlation.
   - All other statistical tests are non-parametric bootstrap t-tests, defined in the Methods section "Statistical Analysis".
   - When testing differences in Modulation Ratio and Normalized Modulation Ratio, logarithms were taken before testing. Besides improving normality, this ensures that the test appropriately evaluates the significance of differences in geometric means (rather than arithmetic means). Explained in Methods/ Statistical Analysis.

   - Tests are defined in the Methods section called Statistical Analysis.
   - The bootstrap t-test is nonparametric and was chosen to relax the assumption of normality. Methods/ Statistical Analysis.

   - Estimates of the variance are reported explicitly in the figures and throughout the Results.
   - For comparisons of prediction quality, bootstrap 68% confidence intervals are reported in figure 5b.
   - For comparisons of Modulation Ratio and Normalized Modulation Ratio, full scatter plots are provided in figures 1b; 3a; 6a; 7d. Bootstrap 68% confidence intervals are also reported throughout the Results section.

   - Unpaired, two-sided in Fig. 6b and Fig. 7e; paired, one-sided for all other comparisons. Methods/ Statistical Analysis.

   - No adjustments for multiple comparisons.

   - The exclusion criteria are discussed in detail in the Methods. Section "Characterization of neuronal responses" for images. Section "Inclusion criteria" for neurons.

   - The criteria were defined to ensure that there was measurable surround modulation for each neuron and image included.
4. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data. If no randomization was used, state so. Where does this appear (section, paragraph #)?

No randomization was used. Methods, last paragraph.

5. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included? If no blinding was done, state so. Where (section, paragraph #)?

No blinding was used. Methods, last paragraph.

6. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included? Where (section, paragraph #)?

A statement of compliance is included in the Methods section "Animal preparation and data collection".

7. Is the species of the animals used reported? Where (section, paragraph #)?

The species is first mentioned in the abstract, and reported in detail in the Methods section "Animal preparation and data collection".

8. Is the strain of the animals (including background strains of KO/transgenic animals used) reported? Where (section, paragraph #)?

N/A

9. Is the sex of the animals/subjects used reported? Where (section, paragraph #)?

Methods section "Animal preparation and data collection".

10. Is the age of the animals/subjects reported? Where (section, paragraph #)?

Methods section "Animal preparation and data collection".

11. For animals housed in a vivarium, is the light/dark cycle reported? Where (section, paragraph #)?

N/A

12. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported? Where (section, paragraph #)?

N/A

13. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)? Where (section, paragraph #)?

N/A

14. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported? Where (section, paragraph #)?

N/A
1. If multiple behavioral tests were conducted in the same group of animals, is this reported? Where (section, paragraph #)?

15. If any animals/subjects were excluded from analysis, is this reported? Where (section, paragraph #)?

a. How were the criteria for exclusion defined? Where is this described (section, paragraph #)?

b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study. Where is this described (section, paragraph #)?

Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?

   a. Is antibody catalog number given? Where does this appear (section, paragraph #)?

   b. Where were the validation data reported (citation, supplementary information, Antibodypedia)? Where does this appear (section, paragraph #)?

2. Cell line identity

   a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by ICLAC and NCBI Biosample? Where (section, paragraph #)?

   b. If yes, include in the Methods section a scientific justification of their use--indicate here in which section and paragraph the justification can be found.

   c. For each cell line, include in the Methods section a statement that specifies:
      - the source of the cell lines
      - have the cell lines been authenticated? If so, by which method?
      - have the cell lines been tested for mycoplasma contamination? Where (section, paragraph #)?
Data deposition

Data deposition in a public repository is mandatory for:
- Protein, DNA and RNA sequences
- Macromolecular structures
- Crystallographic data for small molecules
- Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available here. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad.

We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse.

1. Are accession codes for deposit dates provided?
   Where (section, paragraph #)?
   Data can be provided upon request.

Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.
   Custom scripts in C were used for stimulus presentation. Custom Matlab code was used to analyze the data and to train the Bayesian model.

2. If computer code was used to generate results that are central to the paper’s conclusions, include a statement in the Methods section under "Code availability" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.
   The custom code can be provided upon request.

Human subjects

1. Which IRB approved the protocol?
   Where is this stated (section, paragraph #)?

2. Is demographic information on all subjects provided?
   Where (section, paragraph #)?

3. Is the number of human subjects, their age and sex clearly defined?
   Where (section, paragraph #)?

4. Are the inclusion and exclusion criteria (if any) clearly specified?
   Where (section, paragraph #)?
5. How well were the groups matched?
   Where is this information described (section, paragraph #)?

6. Is a statement included confirming that informed consent was obtained from all subjects?
   Where (section, paragraph #)?

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?
   Where (section, paragraph #)?

fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

1. Were any subjects scanned but then rejected for the analysis after the data was collected?
   a. If yes, is the number rejected and reasons for rejection described?
      Where (section, paragraph #)?

2. Is the number of blocks, trials or experimental units per session and/or subjects specified?
   Where (section, paragraph #)?

3. Is the length of each trial and interval between trials specified?

4. Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized.

5. Is the task design clearly described?
   Where (section, paragraph #)?

6. How was behavioral performance measured?

7. Is an ANOVA or factorial design being used?

8. For data acquisition, is a whole brain scan used?
   If not, state area of acquisition.
   a. How was this region determined?
9. Is the field strength (in Tesla) of the MRI system stated?
   a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated?
   b. Are the field-of-view, matrix size, slice thickness, and TE/TR/flip angle clearly stated?

10. Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?

11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?

12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?

13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?

14. Were any additional regressors (behavioral covariates, motion etc) used?

15. Is the contrast construction clearly defined?

16. Is a mixed/random effects or fixed inference used?
   a. If fixed effects inference used, is this justified?

17. Were repeated measures used (multiple measurements per subject)?
   a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?

18. If the threshold used for inference and visualization in figures varies, is this clearly stated?

19. Are statistical inferences corrected for multiple comparisons?
   a. If not, is this labeled as uncorrected?
20. Are the results based on an ROI (region of interest) analysis?
   a. If so, is the rationale clearly described?
   b. How were the ROI’s defined (functional vs anatomical localization)?

21. Is there correction for multiple comparisons within each voxel?

22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?

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### Additional comments

Additional Comments